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# Sugarcane Bagasse Hydrolysis Enhancement by Microwave-Assisted Sulfolane Pretreatment

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**Abstract:** Sugarcane bagasse is the major by-product of the sugarcane industry and, due to its abundant availability, it has been extensively studied for lignocellulosic bioconversion in the production of bioethanol and other value-added commercial products. In the study presented herein, a combined pretreatment using sulfolane, TiO<sub>2</sub> and alkali microwave irradiation (MW-A) was assessed for the dissolution of lignin prior to enzymatic saccharification of holocellulose. Total reducing sugars (TRS) and saccharinic acid yields were investigated. The increase in NaOH concentration up to 5% and in temperature from 120 °C to 140 °C were found to have a positive influence on both yields. While increasing the reaction time from 5 to 60 min only led to an increase in TRS yield <2%, a reaction time of 30 min almost doubled the saccharinic acids production. TRS yields and saccharinic acid production were approximately 5% and 33% higher when the sulfolane-TiO<sub>2</sub> reaction medium was used, as compared to MW-A in water, reaching up to 64.8% and 15.24 g/L of saccharinic acids, respectively. The proposed MW-A pretreatment may hold promise for industrial applications, given the good TRS yields obtained, and the associated enzyme and time/energy savings. The use of sulfolane-TiO<sub>2</sub> reaction medium is encouraged if saccharinic acids are to be recovered too.

**Keywords:** enzymatic hydrolysis; microwave irradiation; NaOH; pretreatment; sulfolane; total reducing sugars

## 1. Introduction

In recent decades, several strategies for the valorization of biomass and its associated waste have been extensively researched for the production of energy and bioproducts. In this context, second-generation (2G) bioethanol from lignocellulosic biomass as a raw material has gained interest in the development and application of sustainable technologies [1].

Lignocellulosic materials are complex structures formed mainly by cellulose, hemicellulose and lignin. For lignocellulosic ethanol production, three steps are required: (i) a pretreatment, which converts the recalcitrant lignocellulosic structure into cellulosic and hemicellulosic intermediates; (ii) an enzymatic hydrolysis, in which enzymes hydrolyze cellulose and hemicellulose polymers to fermentable sugars such as glucose and xylose, respectively; and (iii) a fermentation stage, in which ethanol is produced by microorganisms.

Thus, the goal of the pretreatment is to disrupt the polymeric components of the lignocellulosic matrix, increasing availability of cellulose and hemicellulose so that they can be converted into

monosaccharides and bioethanol in subsequent steps. Nonetheless, lignin is a recalcitrant structure that hinders the enzymes accessibility to aforementioned polysaccharides [2]. Hence, one of the most challenging stages in the second-generation ethanol production process is the pretreatment (dissolution of lignin) prior to enzymatic saccharification of holocellulose [3].

A number of different strategies have been assayed to disrupt the macromolecular structure that binds the matrix of cellulose, hemicellulose and lignin [4]. The choice of an appropriate pretreatment—according to the physical and chemical characteristics of the biomass to be converted into ethanol—is essential for the fractionation of lignocellulosic biomass, lignin removal, the reduction of cellulose crystallinity and the increase in material porosity, resulting in an increase in the amount of fermentable sugars released to the medium [5].

Numerous technologies involving physical and chemical pretreatments—applied separately, simultaneously, or in a sequential manner—have been investigated [6]. Physical pretreatments include: mechanical comminution [7], thermal methods (pyrolysis, steam explosion and hydrothermal) [8,9], microwave irradiation [10] and ultrasonic treatment [11]. On the other hand, chemical pretreatments that make use of acidic catalysts [12], alkalis [13], ionic liquids [14] and organosolvs [15] have also been received extensive attention in the literature.

Combined physical-chemical pretreatments have been reported to have a synergistic effect on the dissolution of lignin, improving subsequent enzymatic saccharification [16,17]. In this regard, microwave irradiation combined with chemical catalysts (acids, alkalis, organosolvs and ionic liquids) holds particular promise in comparison with other heating systems [18,19]. The main advantage of microwave irradiation (MWI) pretreatments over those based on conventional heating systems is that a homogeneous distribution of heat in the substrate can be achieved, while the latter generate temperature gradients in the biomass. In the MWI-based approach, the molecular collisions generated by dielectric polarization and rapid heat transfer break the complex structure of lignocellulose, resulting in shorter reaction times and in a higher energy efficiency [7,20].

Apropos of the chemical pretreatments, several studies have reported on the efficiency of organosolvs for the conversion of lignocellulosic biomass into hydrolyzed cellulose and high-purity lignin, allowing for the recovery of valuable byproducts and improving the economic viability of the overall cellulosic ethanol process [21]. In this type of pretreatment, the lignocellulosic material is subjected to an organic solvent, with or without catalysts [22]. Low-boiling alcohols, such as ethanol and methanol, have been widely used as organosolvs due to their low cost and easy recovery in the process, but they require operating at a high pressure and hence special equipment that is expensive to purchase and operate has to be used [23]. High-boiling alcohols, such as ethylene glycol or glycerol, can also be used at low temperatures and pressure, but their recovery requires more energy [24].

The search for alternative solvents amenable to be used in lignocellulose pretreatments has led to the consideration of sulfolane, which features several interesting properties: it has a high chemical and thermal stability, and presents solvent properties for the extraction of aromatic hydrocarbons [25]. Thus, sulfolane can be used for the delignification of lignocellulosic materials, given the fact that lignin is a complex molecule composed mainly by aromatic alcohols such as p-coumaryl alcohol, coniferyl alcohol and sinapyl alcohol.

Further, during the pretreatment stage, other platform chemicals can be recovered [26]. For instance, saccharinic acids can be used for the preparation of various fine chemicals; as complexing agents for heavy metals [27], radionuclides and other materials; or as energy sources for aerobic and anaerobic bacteria [28].

In the study presented herein, the effect of a combined pretreatment of sugarcane bagasse (SCB) using organosolv (sulfolane) and alkali microwave irradiation was assessed, with the aim of finding the best pretreatment conditions for a more efficient subsequent enzymatic hydrolysis. Secondarily, the production of saccharinic acids during the pretreatment was also monitored.

## 2. Materials and Methods

### 2.1. Raw Materials and Reagents

Sugarcane bagasse was supplied by a local sugar production company (Ingenio Azucarero del Norte, Imbabura, Ecuador) and was used as the lignocellulosic raw material. The lignocellulosic material was dried, physically processed (cut, milled) and sieved ( $<100\ \mu\text{m}$ ). Titanium dioxide ( $\text{TiO}_2$ ; CAS 1317-70-0; anatase, nanopowder, 99.7%), sulfolane ( $\text{C}_4\text{H}_8\text{O}_2\text{S}$ ; CAS 126-33-0; 99%), 3,5-dinitrosalicylic acid (DNS; CAS 609-99-4; 98%), barbituric acid ( $\text{C}_4\text{H}_4\text{N}_2\text{O}_3$ ; CAS 67-52-7; 99%), 2,4,5,6-tetrahydroxyhexanoic acid ( $\text{C}_6\text{H}_{12}\text{O}_6$ ; CAS 1518-59-8), sodium hydroxide ( $\text{NaOH}$ ; CAS 1310-73-2;  $\geq 97\%$ ), sodium (meta)periodate ( $\text{NaIO}_4$ ; CAS 7790-28-5;  $\geq 99.0\%$ ) and sodium (meta)arsenite ( $\text{NaAsO}_2$ ; CAS 7784-46-5;  $\geq 90\%$ ) were purchased from Sigma Aldrich Química S.A. (Madrid, Spain). Phosphoric acid ( $\text{H}_3\text{PO}_4$ , CAS 7664-38-2; 85%) was supplied by Panreac Química SLU (Castellar del Vallès, Spain). Cellic<sup>®</sup> Ctec2 y Cellic<sup>®</sup> Htec2 enzymes were donated by Novozymes (Bagsværd, Denmark).

### 2.2. Vibrational and SEM Characterization

The vibrational spectra were characterized with a Nicolet iS50 (Thermo Scientific, Waltham, MA, USA) Fourier-Transform Infrared (FTIR) spectrometer equipped with a diamond attenuated total reflection (ATR) module. Spectra were collected in the  $400\text{--}4000\ \text{cm}^{-1}$  region with a  $1\ \text{cm}^{-1}$  spectral resolution, averaging 64 scans.

Scanning electron microscopy (SEM) images were obtained with a FlexSEM 1000 (Hitachi, Chiyoda, Tokyo, Japan) apparatus.

### 2.3. Organosolv on Alkali Microwave-Assisted Pretreatment

SCB pretreatments were carried out in a Milestone Srl (Sorisole, Italy) Ethos-One microwave oven, with and without sulfolane as an organosolv medium and with  $\text{TiO}_2$  as a co-catalyst of the reaction. The effect of the organosolv was tested in alkali conditions, for which 0.5%, 1%, 3% and 5%  $\text{NaOH}$  solutions were tested. The experiment was carried out by mixing 10% ( $w/v$ ) of SCB (dry biomass, particles  $< 100\ \mu\text{m}$ ), 0.2% ( $w/v$ ) of  $\text{TiO}_2$ , and a 1:1 ( $v/v$ )  $\text{NaOH}$  solution to sulfolane ratio. The final concentration of sulfolane in the reaction was 50% ( $v/v$ ) [29]. In pretreatments without organosolv medium, sulfolane and  $\text{TiO}_2$  were replaced by distilled water. Finally, this mixture was subjected to sonication (with a probe-type UIP1000hdT ultrasonicator; Hielscher, Teltow, Germany; 1000 W, 20 kHz) for 1 min and to irradiation in a microwave oven at different temperatures (120, 130 and  $140\ ^\circ\text{C}$ ) and for various reaction times (5, 15, 30, 45 and 60 min). After pretreatment, the solid residue was separated by centrifugation, at 5000 rpm for 10 min. The solid fraction was repeatedly washed with distilled water until a neutral pH was reached, and was dried at  $105\ ^\circ\text{C}$  for 24 h.

### 2.4. Enzymatic Hydrolysis

The enzymatic hydrolysis of the solid residues obtained from the pretreatments was carried out using Cellic<sup>®</sup> Ctec2 and Cellic<sup>®</sup> Htec2 enzymes. For the reactions, a loading of Cellic<sup>®</sup> Ctec 2 of 10 filterpaper units (FPU)/g of substrate and 20% Cellic<sup>®</sup> Htec2 (based on the amount of Cellic<sup>®</sup> Ctec2 loading) was added to a 50 mM sodium citrate buffer (pH 4.8), and then the solid substrate was mixed up to 15% ( $w/v$ ). Enzymatic hydrolysis experiments were performed in triplicate at  $50\ ^\circ\text{C}$ , 100 rpm for 72 h.

### 2.5. Total Reducing Sugars Measurements

The total amount of reducing sugars (TRS) in the enzymatic hydrolysates was determined spectrometrically according to the DNS method (Miller, 1959). The hydrolysate was filtered and 2 mL of the filtrate were mixed in a test tube with 2 mL of DNS reagent. The test tube was incubated in a boiling water bath for 10 min and immediately immersed in an ice bath to stop the reaction. The solution of the sample and the DNS reagent was calibrated with distilled water to a final volume of 15 mL and the absorbance of the solution was measured at 540 nm. Glucose was used for the calibration curve. The TRS yield was obtained with Equation (1):

$$\text{TRS (\%)} = \frac{\text{Concentration of TRS (g}\cdot\text{L}^{-1}\text{)}}{\text{Initial biomass concentration (g}\cdot\text{L}^{-1}\text{)}} \times 100 \quad (1)$$

### 2.6. Saccharinic Acid Measurements

Saccharinic acids were spectrometrically quantified according to the method proposed by Ko [30]. In a tube, 0.2 M sodium (meta)periodate (0.1 mL) and the hydrolysate resulting from SCB alkaline pretreatment (0.2 mL) were mixed. The mixture was left in incubation at room temperature for 20 min, and then sodium arsenite 5% (1 mL) and barbituric acid (1.5 mL) were added. The solution was heated at 100 °C for 30 min, cooled to room temperature, and the absorbance was determined at 486 nm. The same procedure was applied to the standard (2,4,5,6-tetrahydroxyhexanoic acid) to determine the calibration curve.

### 2.7. Statistical Analysis

The data was statistically processed by using IBM (Armonk, NY, USA) SPSS Statistics v.25 software, performing an analysis of variance (ANOVA). The means of experiments were compared using Tukey's HSD (honestly significant difference) test with a significance level of 0.05.

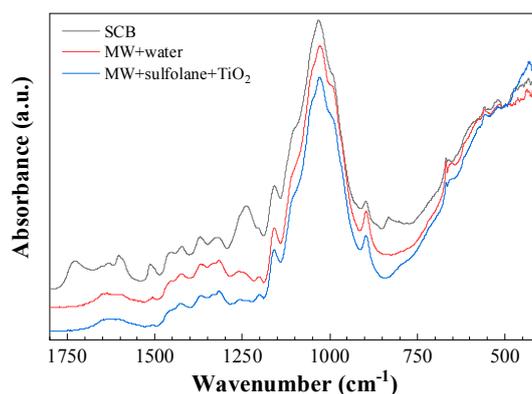
## 3. Results and Discussion

### 3.1. Vibrational and SEM Characterization

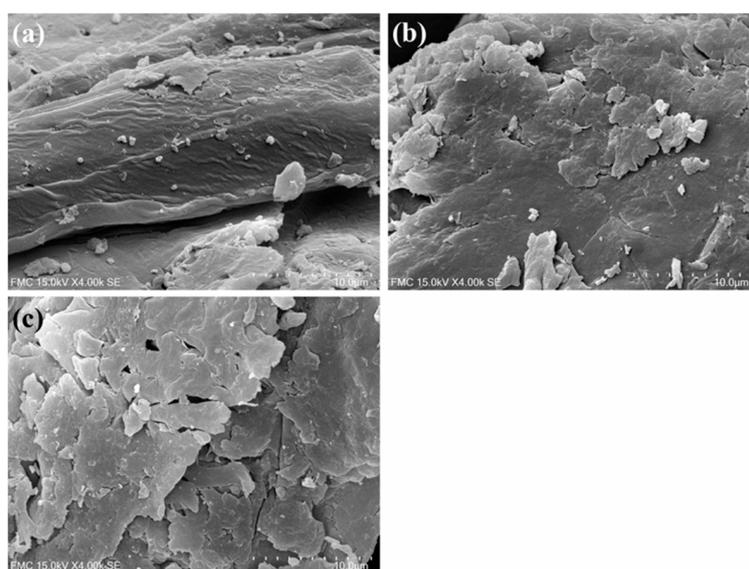
The ATR-FTIR spectra of the untreated and treated SCB samples are depicted in Figure 1. Both pretreated and treated residues exhibited a peak at around 2900  $\text{cm}^{-1}$ , which indicates  $-\text{CH}_2$  stretching; a peak at 897  $\text{cm}^{-1}$ , which indicates  $\beta$ -glucosidic linkages (present in cellulose and xylan); and a strong band at 1031  $\text{cm}^{-1}$ , attributed to C–O–C stretching from cellulose and hemicelluloses [31].

The treated bagasse presented a notable decrease in the absorbance of peaks attributed to lignin, such as the one at 1729  $\text{cm}^{-1}$ , attributed to ferulate and p-coumarate esters [32]; those at 1603  $\text{cm}^{-1}$  and 1513  $\text{cm}^{-1}$ , ascribed to the vibration of the aromatic ring [33]; the one at 1374  $\text{cm}^{-1}$ , associated with syringyl group [34]; and the one at 832  $\text{cm}^{-1}$ , attributed to the C-H out of plane bending vibrations in the p-hydroxyphenyl propane units [35]. While the increased intensity of the band at 898  $\text{cm}^{-1}$  showed an increase in cellulose content due to the delignification, the decrease in intensity of the band at around 1369  $\text{cm}^{-1}$  indicated an increase in amorphous cellulose [36]. Therefore, all these alterations in the FTIR-ATR spectra are consistent with the effectiveness of the two MWI treatments for lignin removal.

Representative SEM micrographs of both untreated and treated SCB are shown in Figure 2. Changes in morphology could be observed after MWI treatment in both media, resulting in a more flaky structure. Nonetheless, they were slightly more evident in the sulfolane-based medium than in the water-based medium, suggesting a higher degree of attack for the former.



**Figure 1.** Attenuated total reflection-Fourier-transform infrared (ATR-FTIR) spectra of sugarcane bagasse (SCB) samples: before treatment (black); after microwave irradiation (MWI) treatment with NaOH 3% at 140 °C for 60 min in the water-based medium (red), and in the sulfolane-based medium (blue).



**Figure 2.** SEM micrographs of SCB samples at 4000× magnification: (a) before treatment; after MWI treatment with NaOH 3% at 140 °C for 60 min in the (b) water-based medium and in the (c) sulfolane-based medium.

### 3.2. Influence of Sodium Hydroxide Concentration, Temperature and Reaction Time on TRS Yield

#### 3.2.1. NaOH Concentration

An increase in TRS yields was observed when the alkali concentration was increased from 0.5% to 5%, both with and without organosolv (Table 1). This can be ascribed to the fact that NaOH is one of the strongest base catalysts, and its effectiveness for the pretreatment of lignocellulosic biomass has been evidenced by a higher degree of enzymatic hydrolysis compared to other alkaline pretreatments in the literature [37,38].

The alkaline pretreatment reactions include the dissolution of lignin and hemicellulose, and the de-esterification (saponification) of intermolecular ester bonds cross-linking hemicellulose and lignin. During the NaOH pretreatment reaction, NaOH is dissociated into hydroxide ion ( $\text{OH}^-$ ) and sodium ion ( $\text{Na}^+$ ) [13], and the increase in the concentration of hydroxide ions increases the rate of the hydrolysis reaction. This is consistent with the results obtained in this study, in which the increase in TRS production was evident when NaOH concentration was increased to 3% and 5%, reaching yields

of up to 61.1% and 64.8%, respectively. Significant differences were found for both pretreatments, with sulfolane-TiO<sub>2</sub> (Table 2) and without sulfolane-TiO<sub>2</sub> (Table A1).

In an investigation carried out by Hoşgün [39], pretreatments at different NaOH concentrations in the 2 to 10% (*w/v*) range were tested on lignocellulosic material (hazelnut shells). The results evidenced that the increase in NaOH concentration led to greater lignin removal. TRS yields improved up to a NaOH concentration of 6%, but further increase resulted in a decrease in TRS production.

**Table 1.** Total reducing sugars (TRS) production from enzymatic hydrolysis of SCB for different temperatures, NaOH concentrations and reaction times, with and without sulfolane-TiO<sub>2</sub>.

Pretreatment	T (°C)	NaOH Concentration (% <i>w/v</i> )	TRS Yield (%)				
			5 min	15 min	30 min	45 min	60 min
MW-A only (without sulfolane-TiO <sub>2</sub> )	120	0.5	29.7 ± 1.08	29.5 ± 2.13	30.0 ± 0.92	30.7 ± 1.28	31.0 ± 2.43
		1	35.2 ± 1.21	35.3 ± 0.33	36.4 ± 1.02	36.9 ± 1.34	37.4 ± 1.11
		3	49.0 ± 2.47	49.3 ± 0.51	50.9 ± 1.50	52.0 ± 0.58	52.2 ± 1.04
		5	53.5 ± 1.99	53.0 ± 2.36	54.4 ± 0.19	55.8 ± 0.92	56.1 ± 0.51
	130	0.5	31.5 ± 0.42	31.5 ± 1.81	32.1 ± 0.58	32.6 ± 0.92	32.9 ± 1.02
		1	39.1 ± 0.63	39.5 ± 0.98	40.7 ± 1.36	41.8 ± 0.73	42.0 ± 0.67
		3	52.4 ± 1.79	53.0 ± 1.51	53.3 ± 1.79	54.7 ± 1.06	55.0 ± 2.39
		5	56.1 ± 1.58	56.0 ± 2.37	57.0 ± 0.54	57.2 ± 1.51	57.4 ± 1.42
	140	0.5	35.0 ± 1.27	35.0 ± 1.20	35.4 ± 1.67	35.7 ± 0.19	36.2 ± 0.26
		1	40.8 ± 0.73	40.6 ± 0.70	41.9 ± 0.98	42.6 ± 0.33	43.3 ± 1.04
		3	53.3 ± 0.51	53.0 ± 0.17	54.2 ± 0.75	55.7 ± 0.60	56.3 ± 1.44
		5	58.0 ± 0.26	57.6 ± 0.29	59.0 ± 0.35	60.0 ± 0.98	59.5 ± 1.60
MW-A + Sulfolane-TiO <sub>2</sub>	120	0.5	30.4 ± 1.75	30.6 ± 1.94	31.2 ± 0.67	31.6 ± 0.77	32.0 ± 2.07
		1	36.9 ± 0.70	36.8 ± 0.42	37.9 ± 0.19	38.3 ± 0.79	38.5 ± 0.26
		3	52.4 ± 0.77	52.0 ± 0.51	53.4 ± 0.60	54.7 ± 0.88	55.0 ± 0.17
		5	55.3 ± 0.67	56.7 ± 0.48	57.8 ± 0.33	58.0 ± 0.96	57.8 ± 0.88
	130	0.5	33.9 ± 0.84	33.6 ± 0.35	34.7 ± 1.44	35.0 ± 0.92	35.3 ± 0.86
		1	42.3 ± 0.88	42.0 ± 1.17	42.7 ± 0.75	44.1 ± 2.81	44.4 ± 1.64
		3	55.3 ± 1.18	55.7 ± 0.92	56.8 ± 1.21	57.0 ± 1.11	57.6 ± 1.18
		5	61.1 ± 0.44	60.8 ± 1.07	62.1 ± 0.48	63.6 ± 0.79	63.9 ± 1.37
	140	0.5	35.9 ± 0.95	36.2 ± 0.92	36.5 ± 0.60	37.1 ± 0.77	37.8 ± 1.34
		1	44.2 ± 0.92	43.7 ± 0.63	44.1 ± 0.63	45.1 ± 0.39	44.9 ± 0.86
		3	56.1 ± 0.19	56.5 ± 0.77	57.6 ± 0.93	58.0 ± 0.60	61.1 ± 0.79
		5	62.9 ± 0.77	62.8 ± 0.77	64.1 ± 0.58	64.6 ± 0.87	64.8 ± 0.35

### 3.2.2. Temperature

In the enzymatic hydrolysis of lignocellulosic biomass (sweet sorghum stalks) pretreated with organosolv using an acid catalyst, Ostovareh et al. [40] reported that an increase in temperature (100–160 °C) improved enzymatic conversion. However, Koo et al. [41] showed that enzymatic hydrolysis was barely affected beyond 140 °C, thus suggesting that the pretreatment would not need to be conducted at higher temperatures.

In the study presented herein, in line with Ostovareh et al. [40], temperature was found to be an important parameter, resulting in significant differences (Tables 2 and A1): the increase in temperature from 120 °C to 140 °C improved the formation of TRS from enzymatic hydrolysis of SCB by approximately 5%.

### 3.2.3. Reaction Time

Although significant differences were also found among the five reaction times assayed (Tables 2 and A1), it is worth noting that it would be parameter that had the smallest influence: the increase in reaction time from 5 to 60 min only resulted in a 1.9% increase in TRS yield.

Wang et al. [42] reported that pretreatments with organosolv (2-propanol) at higher temperatures (ranging from 200 to 220 °C) and longer reaction times (60 and 120 min) in an autoclave facilitated enzymatic hydrolysis. In contrast, the use of MWI—even under milder conditions (in terms of

temperatures and reaction times, such as those used as in this study)—would noticeably speed up the process, resulting in TRS yields of up to 62.9% in only 5 min.

**Table 2.** Tukey’s HSD test results for the MW-A + sulfolane-TiO<sub>2</sub> pretreatment in relation to the TRS yield (%). Analysis of the differences between the categories with a confidence interval of 95% for: temperature (top), NaOH concentration (center) and time (bottom).

T (°C)	LS Means	Standard Error	Lower Bound (95%)	Upper Bound (95%)	Groups
120	44.865	0.139	44.590	45.140	A
130	49.094	0.139	48.819	49.369	B
140	50.702	0.139	50.427	50.977	C
NaOH (% w/v)	LS Means	Standard Error	Lower Bound (95%)	Upper Bound (95%)	Groups
0.5	34.116	0.161	33.799	34.434	A
1	41.729	0.161	41.412	42.047	B
3	55.951	0.161	55.633	56.268	C
5	61.085	0.161	60.768	61.403	D
Time (min)	LS Means	Standard Error	Lower Bound (95%)	Upper Bound (95%)	Groups
5	47.226	0.180	46.871	47.581	A
15	47.285	0.180	46.930	47.640	A
30	48.241	0.180	47.886	48.596	B
45	48.924	0.180	48.568	49.279	B
60	49.427	0.180	49.072	49.782	C

LS means: least-squares means.

In view of aforementioned results, NaOH 5%, 140 °C and 5 min would be the most appropriate conditions for the pretreatment of lignocellulosic biomass with sulfolane in alkaline conditions under MWI, from the point of view of sugar yield.

### 3.3. Influence of Sodium Hydroxide Concentration, Temperature and Reaction Time on Saccharinic Acids Yield

#### 3.3.1. NaOH Concentration

In a similar fashion to the TRS yield, saccharinic acids production systematically increased as NaOH concentration was increased (Table 3). Significant differences were found between the four assayed concentrations for both pretreatment media: TiO<sub>2</sub> + sulfolane (Table 4, top) and water (Table A2). Improvement factors of up to 3.24 and 2.65 were obtained for the water and sulfolane media, respectively, depending on the other two parameters under study.

#### 3.3.2. Temperature

Statistically significant differences in the saccharinic acids production were also detected as a function of temperature (Table 4, center; Table A2): Depending on the reaction time and NaOH concentration, improvement factors when the temperature was increased from 120 °C to 140 °C ranged from 1.15 to 2.33 for the water pretreatment medium, and from 1.18 to 1.88 for the sulfolane medium.

#### 3.3.3. Reaction Time

In relation to the effect of reaction time, the behavior differed from that reported above for the TRS yield. In this case, a significant improvement in saccharinic acids production occurred when the reaction time was increased from 5 to 30 min (Table 4, bottom; Table A2). The enhancement factor was in the 1.2–2.9 range for the water-based medium, and in the 1.4–2.3 range for the sulfolane medium, depending on temperature and NaOH concentration. Further increase of the reaction time (to 45 and 60 min) did not lead to additional saccharinic acids production.

Taking all parameters into account, the best combination in terms of saccharinic acid production was NaOH 5%, 140 °C and 30 min. In these conditions, 11.31 g/L and 15.08 g/L were obtained for the water-based and the sulfolane-based pretreatments, respectively (i.e., the sulfolane medium resulted in a 33% higher production).

**Table 3.** Saccharinic acids production from enzymatic hydrolysis of SCB for different temperatures, NaOH concentrations and reaction times, with and without sulfolane-TiO<sub>2</sub>.

Pretreatment	T (°C)	NaOH Concentration (% w/v)	Saccharinic Acids (g/L)				
			5 min	15 min	30 min	45 min	60 min
MW-A only (without sulfolane-TiO <sub>2</sub> )	120	0.5	1.19 ± 0.04	1.41 ± 0.10	2.44 ± 0.17	2.51 ± 0.17	2.42 ± 0.12
		1	1.75 ± 0.18	2.12 ± 0.09	2.93 ± 0.22	3.01 ± 0.04	3.05 ± 0.16
		3	2.18 ± 0.14	2.34 ± 0.07	3.44 ± 0.07	3.52 ± 0.14	3.56 ± 0.08
		5	3.86 ± 0.04	3.82 ± 0.14	6.05 ± 0.04	5.99 ± 0.07	6.20 ± 0.04
	130	0.5	2.02 ± 0.10	2.16 ± 0.06	3.95 ± 0.10	4.07 ± 0.00	4.15 ± 0.05
		1	2.10 ± 0.11	2.32 ± 0.13	5.57 ± 0.10	5.65 ± 0.14	5.61 ± 0.07
		3	2.30 ± 0.07	2.77 ± 0.05	6.60 ± 0.13	6.38 ± 0.06	6.64 ± 0.12
		5	6.34 ± 0.15	6.30 ± 0.06	7.64 ± 0.18	7.70 ± 0.10	7.76 ± 0.12
	140	0.5	2.34 ± 0.04	2.26 ± 0.10	4.35 ± 0.14	4.47 ± 0.09	4.43 ± 0.03
		1	2.50 ± 0.15	2.44 ± 0.08	6.08 ± 0.10	6.12 ± 0.10	6.16 ± 0.10
		3	2.93 ± 0.04	3.09 ± 0.05	8.02 ± 0.09	8.10 ± 0.06	8.06 ± 0.10
		5	7.05 ± 0.14	7.19 ± 0.11	11.31 ± 0.10	11.39 ± 0.09	11.29 ± 0.29
MW-A + sulfolane-TiO <sub>2</sub>	120	0.5	2.81 ± 0.21	2.91 ± 0.48	4.57 ± 0.12	4.37 ± 0.18	4.49 ± 0.02
		1	3.03 ± 0.17	3.17 ± 0.15	4.98 ± 0.31	5.08 ± 0.22	4.94 ± 0.32
		3	3.22 ± 0.13	3.36 ± 0.10	5.61 ± 0.18	5.67 ± 0.19	5.71 ± 0.12
		5	5.73 ± 0.12	5.85 ± 0.20	8.04 ± 0.09	8.18 ± 0.18	8.14 ± 0.30
	130	0.5	2.89 ± 0.21	3.22 ± 0.08	5.20 ± 0.10	5.22 ± 0.00	5.32 ± 0.19
		1	3.13 ± 0.22	3.30 ± 0.16	6.54 ± 0.16	6.64 ± 0.31	6.68 ± 0.14
		3	3.54 ± 0.28	3.88 ± 0.28	7.45 ± 0.25	7.60 ± 0.21	7.56 ± 0.29
		5	7.66 ± 0.15	7.86 ± 0.34	10.52 ± 0.22	10.48 ± 0.21	10.68 ± 0.10
	140	0.5	3.32 ± 0.26	3.56 ± 0.19	5.85 ± 0.08	5.81 ± 0.12	5.93 ± 0.20
		1	3.64 ± 0.08	3.88 ± 0.20	7.13 ± 0.16	7.27 ± 0.16	7.17 ± 0.14
		3	3.88 ± 0.09	3.95 ± 0.12	8.93 ± 0.15	9.00 ± 0.13	9.14 ± 0.08
		5	8.25 ± 0.14	8.41 ± 0.26	15.08 ± 0.11	14.92 ± 0.07	15.24 ± 0.19

**Table 4.** Tukey's HSD test results for the MW-A + sulfolane-TiO<sub>2</sub> pretreatment in relation to the saccharinic acids yield (g/L). Analysis of the differences between the categories with a confidence interval of 95% for: temperature (top), NaOH concentration (center) and time (bottom).

T (°C)	LS Means	Standard Error	Lower Bound (95%)	Upper Bound (95%)	Groups
120	4.992	0.025	4.942	5.042	A
130	6.268	0.025	6.218	6.318	B
140	7.518	0.025	7.468	7.568	C
NaOH (% w/v)	LS Means	Standard Error	Lower Bound (95%)	Upper Bound (95%)	Groups
0.5	4.364	0.029	4.306	4.421	A
1	5.104	0.029	5.046	5.162	B
3	5.901	0.029	5.843	5.958	C
5	9.669	0.029	9.612	9.727	D
Time (min)	LS Means	Standard Error	Lower Bound (95%)	Upper Bound (95%)	Groups
5	4.259	0.033	4.194	4.323	A
15	4.446	0.033	4.382	4.511	B
30	7.490	0.033	7.426	7.555	C
45	7.520	0.033	7.455	7.584	C
60	7.582	0.033	7.518	7.647	C

LS means: least-squares means.

### 3.4. Effect of the Pretreatment Reaction Medium

#### 3.4.1. TRS Yield

The statistical analysis showed significant differences between the two reaction media (Table 5), indicating that sulfolane-TiO<sub>2</sub> promoted the production of TRS, attaining yields of up to 64.8% (vs. yields of up to 59.5% for the pretreatment without the addition of sulfolane-TiO<sub>2</sub>). This relatively small difference points at the very positive impact of MWI, regardless of the reaction medium.

In the studies carried out by Peng et al. [43,44] in which MWI pretreatment was assayed on microcrystalline cellulose, they reported that noticeably shorter times were needed in order to reach a favorable saccharides yield. Moreover, in the assays conducted by Lai et al. [45], and Podschun et al. [46], pretreatments with different heating systems that included a steam-alkali-chemical combination, conventional thermo-chemical (NaOH) and microwave-alkali (MW-A) pretreatments on lignocellulosic biomass were compared. They demonstrated that the MW-A was the most effective method for the disruption of lignocellulosic materials and for improving enzyme saccharification, suggesting that it may be the best alternative due to its lower energy consumption and shorter reaction times, which make it economically viable in industrial scale processes.

**Table 5.** Tukey's HSD test results for the TRS yield (%). Analysis of the differences between the two reaction media with a confidence interval of 95%. "1" stands for the pretreatment medium with sulfolane-TiO<sub>2</sub> and "2" for the medium without it.

Contrast	Difference	Standardized Difference	Critical Value	Pr > Diff	Significant
2 vs. 1	-2.762	-20.155	1.967	<0.0001	Yes
Tukey's d critical value:		2.781			
Category	LS Means	Standard Error	Lower Bound (95%)	Upper Bound (95%)	Groups
2	45.458	0.097	45.268	45.649	A
1	48.220	0.097	48.030	48.411	B

LS means: least-squares means.

### 3.4.2. Saccharinic Acids

Apropos of saccharinic acids production, statistically significant differences were also detected between the two reaction media (Table 6), with a higher production of these byproducts for sulfolane-TiO<sub>2</sub>-based pretreatment.

**Table 6.** Tukey's HSD test results for the saccharinic acids yield (g/L). Analysis of the differences between the two reaction media with a confidence interval of 95%. "1" stands for the pretreatment medium with sulfolane-TiO<sub>2</sub> and "2" for the medium without it.

Contrast	Difference	Standardized Difference	Critical Value	Pr > Diff	Significant
2 vs. 1	-1.570	-63.626	1.969	<0.0001	Yes
Tukey's d critical value:		2.785			
Category	LS Means	Standard Error	Lower Bound (95%)	Upper Bound (95%)	Groups
2	4.690	0.017	4.655	4.724	A
1	6.259	0.017	6.225	6.294	B

LS means: least-squares means.

### 3.5. Comparison with Other Pretreatment Conditions Reported in the Literature

Table 7 shows a comparison of the pretreatment efficiency attained through the proposed procedure vs. those achieved with other pretreatments reported in the literature for SBC. It should be stressed that the lack of a common agreement in relation to the units (results may be expressed in terms of glucose yield, TRS production -in mg/g or in g/L-, etc.) jeopardizes a direct comparison among studies. Comparisons of the yields below should therefore be taken with caution.

In view of Table 7, it may be inferred that higher yields may be attained with other pretreatments, but at the expense of higher enzyme loadings and longer pretreatment times. As noted by Mithra et al. [47], the cost of enzymatic hydrolysis stage is critical in the low-cost production of 2G ethanol. Thus, the proposed MW-A approach, with much shorter processing times, regardless of the chosen reaction medium, may be regarded as an interesting option for industrial applications from the cost-efficiency point of view. Moreover, the system may be supplemented with detoxification chemicals to further reduce cellulase dosage [47–49]. Further research to explore this possibility is under way.

**Table 7.** Comparison of the yield results presented herein with the reported literature.

Pretreatment				Saccharification Conditions	Yield		Reference
Reaction Medium and Catalysts	T (°C)	Time (min)	Glucose (%)		TRS (mg/g)		
Methanol (65% <i>w/v</i> ) + soda-green liquor (1.5 mL/g substrate)	120	180	Enzymatic hydrolysis with 18 FPU cellulases/g biomass, at 50 °C for 72 h	95.7	747.4 *	[50]	
Ethanol:H <sub>2</sub> O (60:40, <i>v/v</i> ) + FeCl <sub>3</sub> (0.025 mol·L <sup>-1</sup> )	160	60	Enzymatic hydrolysis with 20 FPU cellulase/g dried substrate, at 50 °C for 72 h	93.8	732.6 *	[49]	
			Enzymatic hydrolysis with 20 FPU cellulase/g dried substrate + Tween-80 (150 mg/g substrate), at 50 °C for 6 h	92.5	722.4 *		
Ethanol:water (50:50, <i>v/v</i> ) + green liquor (mainly sodium carbonate and sodium hydroxide, 0.5–1.5 mL/g dry substrate) + anthraquinone (0.1% <i>w/v</i> )	120	180	Enzymatic hydrolysis with 12 FPU cellulases/g biomass + 18 CBU cellobiase/g biomass, 48 °C, 72 h	94.6	738.8 *	[51]	
Biphasic solution of n-propanol (588 g/kg), water (400 g/kg) and H <sub>2</sub> SO <sub>4</sub> (12 g/kg) + (NH <sub>4</sub> ) <sub>2</sub> SO <sub>4</sub> (400 g/kg) to promote formation of a biphasic solution	140	90	Enzymatic hydrolysis with 20 FPU cellulases/g biomass, at 50 °C for 72 h	86.1	672.4 *	[52]	
Water/ethanol/ethyl acetate/formic acid (43:20:16:21% <i>v/v</i> )	160	40	Enzymatic hydrolysis with 15 FPU cellulases/g biomass, at 50 °C for 72 h	84.5	659.9 *	[53]	
Water:sulfolane (1:1 <i>v/v</i> ) + TiO <sub>2</sub> (0.2% <i>w/v</i> ), NaOH 5% Water, NaOH 5%	140	5	Enzymatic hydrolysis with 10 FPU cellulase/g dried substrate, at 50 °C for 72 h	80.5 *	629.0	This study	
			74.3 *	580.0			
Ethanol: Water (1:1, <i>v/v</i> ) + high-pressure CO <sub>2</sub> (6.8 MPa) + additional liquid hot water pretreatment at 180 °C for 20 min.	112	45	Enzymatic hydrolysis with 10 FPU of cellulases/g biomass + 20 IU β-glucosidase/g biomass, at 50 °C for 72 h	63.7	497.5 *	[54]	
Methyl isobutyl ketone (MIBK): Methanol: Water (16:68:16, <i>v/v</i> ) + H <sub>3</sub> PO <sub>4</sub> (5% <i>w/w</i> ) activated carbon	180	60	Hot compressed water at 225 °C, 10 min	58.3	455.3 *	[55]	
H <sub>2</sub> SO <sub>4</sub> (0.1 M)	120	120	Enzymatic hydrolysis with 10 FPU cellulases/g biomass, at 50 °C for 48 h	57.9 *	452.3	[56]	
CH <sub>3</sub> ONa (0.6% <i>w/w</i> ) + 80% glycerol	130	60	Enzymatic hydrolysis with 6 FPU cellulases/g biomass + 6.3 U of xylanase/g biomass, at 45 °C for 72 h	48.0	374.9 *	[57]	
Ethanol: Water (40:60, <i>v/v</i> )	195	30	Enzymatic hydrolysis with 10 FPU cellulases/g biomass, at 50 °C for 60 h	40.3	314.7 *	[58]	

\* A conversion factor of 7.81 was used to transform glucose yield (%) into TRS (mg/g), taking as a reference the average values reported by Rabelo et al. [59].

#### 4. Conclusions

The use of sulfolane-TiO<sub>2</sub> in combination with alkali MWI pretreatment was assessed on SCB. The use of sulfolane as a solvent resulted in 5% higher TRS yields and 33% higher saccharinic acids production than the water-based medium. The former led to TRS yields of up to 64.8% and 15.24 g/L of saccharinic acids, while the water-based treatment led to TRS and saccharinic acid yields of up to 59.5% and 11.31 g/L, respectively. Regardless of the reaction medium, the best results were attained for a NaOH concentration of 5% at a temperature of 140 °C. In relation to the reaction time, 5 min was found to be the best choice for TRS production, while 30 min would optimize saccharinic acids production. While the MW-A in water approach may be preferable with a view to industrial applications due to environmental and cost factors, the sulfolane-TiO<sub>2</sub> medium clearly outperforms it if saccharinic acids are to be recovered as high-added value byproducts. Regardless of the chosen reaction medium, the proposed MW-A pretreatment results in high glucose yields with short reaction times and enzyme-associated savings.

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#### Appendix A

**Table A1.** Tukey's HSD test results for the MW-A pretreatment in water medium in relation to the TRS yield (%). Analysis of the differences between the categories with a confidence interval of 95% for: temperature (top), NaOH concentration (center) and time (bottom).

T (°C)	LS Means	Standard Error	Lower Bound (95%)	Upper Bound (95%)	Groups
120	42.921	0.159	42.608	43.235	A
130	45.797	0.159	45.484	46.111	B
140	47.656	0.159	47.343	47.969	C
NaOH (% w/v)	LS Means	Standard Error	Lower Bound (95%)	Upper Bound (95%)	Groups
0.5	32.602	0.183	32.240	32.964	A
1	39.566	0.183	39.204	39.928	B
3	52.956	0.183	52.594	53.318	C
5	56.709	0.183	56.347	57.071	D
Time (min)	LS Means	Standard Error	Lower Bound (95%)	Upper Bound (95%)	Groups
15	44.471	0.205	44.066	44.875	A
5	44.436	0.205	44.031	44.841	A
30	45.453	0.205	45.049	45.858	B
45	46.321	0.205	45.916	46.726	C
60	46.610	0.205	46.205	47.015	C

LS means: least-squares means.

**Table A2.** Tukey’s HSD test results for the MW-A pretreatment in water medium in relation to the saccharinic acids yield (g/L). Analysis of the differences between the categories with a confidence interval of 95% for: Temperature (top), NaOH concentration (center) and time (bottom).

T (°C)	LS Means	Standard Error	Lower Bound (95%)	Upper Bound (95%)	Groups
120	3.188	0.015	3.160	3.217	A
130	4.902	0.015	4.874	4.931	B
140	5.978	0.015	5.949	6.007	C
NaOH (% w/v)	LS Means	Standard Error	Lower Bound (95%)	Upper Bound (95%)	Groups
0.5	2.945	0.017	2.911	2.978	A
1	3.827	0.017	3.794	3.860	B
3	4.661	0.017	4.627	4.694	C
5	7.326	0.017	7.293	7.359	D
Time (min)	LS Means	Standard Error	Lower Bound (95%)	Upper Bound (95%)	Groups
5	3.046	0.019	3.008	3.083	A
15	3.184	0.019	3.147	3.221	B
30	5.699	0.019	5.661	5.736	C
45	5.743	0.019	5.706	5.780	C
60	5.777	0.019	5.740	5.815	

LS means: least-squares means.

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