



## Original article

# Mechanisms of pro-arrhythmic abnormalities in ventricular repolarisation and anti-arrhythmic therapies in human hypertrophic cardiomyopathy



Elisa Passini<sup>a,b</sup>, Ana Mincholé<sup>a</sup>, Raffaele Coppini<sup>c</sup>, Elisabetta Cerbai<sup>c</sup>, Blanca Rodriguez<sup>a</sup>, Stefano Severi<sup>b</sup>, Alfonso Bueno-Orovio<sup>a,\*</sup>

<sup>a</sup> Department of Computer Science, University of Oxford, Oxford OX13QD, United Kingdom

<sup>b</sup> Department of Electrical, Electronic and Information Engineering, University of Bologna, Cesena 47521, Italy

<sup>c</sup> Department NEUROFARBA, University of Florence, Florence 50139, Italy

## ARTICLE INFO

## Article history:

Received 7 June 2015

Received in revised form 5 August 2015

Accepted 11 September 2015

Available online 16 September 2015

## Keywords:

Hypertrophic cardiomyopathy

Pro-arrhythmic mechanisms

Repolarization reserve

Inter-subject variability

In silico drug testing

## ABSTRACT

**Introduction:** Hypertrophic cardiomyopathy (HCM) is a cause of sudden arrhythmic death, but the understanding of its pro-arrhythmic mechanisms and an effective pharmacological treatment are lacking. HCM electrophysiological remodelling includes both increased inward and reduced outward currents, but their role in promoting repolarisation abnormalities remains unknown. The goal of this study is to identify key ionic mechanisms driving repolarisation abnormalities in human HCM, and to evaluate anti-arrhythmic effects of single and multichannel inward current blocks.

**Methods:** Experimental ionic current, action potential (AP) and  $\text{Ca}^{2+}$ -transient (CaT) recordings were used to construct populations of human non-diseased and HCM AP models ( $n = 9118$ ), accounting for inter-subject variability. Simulations were conducted for several degrees of selective and combined inward current block.

**Results:** Simulated HCM cardiomyocytes exhibited prolonged AP and CaT, diastolic  $\text{Ca}^{2+}$  overload and decreased CaT amplitude, in agreement with experiments. Repolarisation abnormalities in HCM models were consistently driven by L-type  $\text{Ca}^{2+}$  current ( $I_{\text{CaL}}$ ) re-activation, and  $I_{\text{CaL}}$  block was the most effective intervention to normalise repolarisation and diastolic  $\text{Ca}^{2+}$ , but compromised CaT amplitude. Late  $\text{Na}^+$  current ( $I_{\text{NaL}}$ ) block partially abolished repolarisation abnormalities, with small impact on CaT.  $\text{Na}^+/\text{Ca}^{2+}$  exchanger ( $I_{\text{NCX}}$ ) block effectively restored repolarisation and CaT amplitude, but increased  $\text{Ca}^{2+}$  overload. Multichannel block increased efficacy in normalising repolarisation, AP biomarkers and CaT amplitude compared to selective block.

**Conclusions:** Experimentally-calibrated populations of human AP models identify  $I_{\text{CaL}}$  re-activation as the key mechanism for repolarisation abnormalities in HCM, and combined  $I_{\text{NCX}}$ ,  $I_{\text{NaL}}$  and  $I_{\text{CaL}}$  block as effective anti-arrhythmic therapies also able to partially reverse the HCM electrophysiological phenotype.

© 2015 The Authors. Published by Elsevier Ltd. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

## 1. Introduction

Hypertrophic cardiomyopathy (HCM) is the most common monogenic cardiac disorder and the main cause of sudden cardiac death in children and young adults [1], with a reported prevalence of 1 in 500 worldwide [2]. Usually asymptomatic, it is characterised by an unexplained thickening (hypertrophy) of the left ventricle, and occasionally of the right, with predominant involvement of the inter-ventricular septum. However, the ejection fraction is usually preserved in HCM patients [3,4], with only a minority of the subjects developing enlarged ventricular cavities, hence pointing towards a different aetiology of

the disease compared to acquired heart failure. In addition, the first manifestation of HCM is often arrhythmic sudden death, caused by ventricular tachyarrhythmias [2], but the underlying electrophysiological mechanisms remain unclear.

HCM is still lacking of a disease-specific pharmacological treatment [5–7]. To date, implantable cardioverter-defibrillator therapy prevails as the only effective prevention of sudden cardiac death in HCM [8,9]. Different mutations are associated with different outcome in HCM patients, but the strength of the genotype-phenotype correlation is weak to warrant recommendation in risk management based on the genotype, due to significant inter-subject variability in disease expression, even among carriers of the same variant [10]. Therefore, a better understanding of the ionic mechanisms underlying arrhythmic risk and inter-subject variability in HCM is required to guide the development of specific pharmacological treatments and risk stratification.

\* Corresponding author at: Department of Computer Science, University of Oxford, Wolfson Building, Parks Road Oxford OX13QD, United Kingdom.  
E-mail address: [alfonso.bueno@cs.ox.ac.uk](mailto:alfonso.bueno@cs.ox.ac.uk) (A. Bueno-Orovio).

Recently, Coppini et al. characterised the electrophysiological profile of human HCM by measuring alterations in the action potential (AP),  $\text{Ca}^{2+}$  subsystem, sarcolemmal ionic currents and mRNA expression in donor HCM cardiomyocytes [3]. Particularly distinct to the ionic remodelling associated to heart failure [11], HCM cardiomyocytes exhibited a significant overexpression of the L-type  $\text{Ca}^{2+}$  current ( $I_{\text{CaL}}$ ), which together with an increase in the Late  $\text{Na}^+$  ( $I_{\text{NaL}}$ ) and a reduction in  $\text{K}^+$  repolarising currents, contributed to a prolonged AP and  $\text{Ca}^{2+}$  transient (CaT). Experimental recordings also provided evidence of HCM facilitating the occurrence of repolarisation abnormalities such as early after-depolarisations (EADs), which may act as triggers for ventricular arrhythmias [12,13].

Repolarisation abnormalities and in particular EADs are known to be facilitated by abnormalities in the  $\text{Ca}^{2+}$  subsystem and  $\text{Ca}^{2+}$  overload [14–16]. Genetic mutations in HCM commonly enhance  $\text{Ca}^{2+}$ -sensitivity and energy requirements of myosin ATPase, leading to altered force production and impairment of intracellular  $\text{Ca}^{2+}$  and intracellular  $\text{Ca}^{2+}$  load [6,8]. Indeed, experimental data in human HCM provided evidence of a highly impaired  $\text{Ca}^{2+}$  subsystem, owing to a reduction of  $\text{Ca}^{2+}$  uptake by SERCA and an altered  $\text{Na}^+/\text{Ca}^{2+}$  exchanger function, contributing to the slower kinetics of CaT and the elevated diastolic  $\text{Ca}^{2+}$  [3]. Further investigations are therefore required to advance our understanding of the ionic mechanisms underlying pro-arrhythmic repolarisation abnormalities in HCM.

The goals of this study are to investigate the ionic mechanisms driving repolarisation abnormalities in human HCM cardiomyocytes, and to evaluate the efficacy of selective and combined inward currents block as potential anti-arrhythmic strategy. Building on the comprehensive dataset provided by Coppini et al. [3] and on the methodology proposed by Britton et al. [17], we construct two populations of human ventricular AP models in range with the experimental data and accounting for cell-to-cell variability in non-diseased and HCM conditions. Based on our analysis in these populations, we propose different single and multi-channel strategies for the pharmacological management of cellular repolarisation abnormalities in HCM, evaluating their efficacy in abolishing repolarisation abnormalities and in reversing the electrophysiological phenotypic characteristics of human HCM cardiomyocytes.

## 2. Materials and methods

### 2.1. Experimental data

The experimental dataset, previously published in [3], consists of recordings from  $n = 80$  HCM cells and  $n = 31$  non-failing non-hypertrophic control cells (CTRL). HCM cardiomyocytes were hypertrophic, as indicated by an increased cell volume (+90%) compared with CTRL ( $33.5 \pm 4.3$  vs  $17.6 \pm 3.2$  pL,  $P < 0.05$ ). Single cell patch-clamp measurements and intracellular  $\text{Ca}^{2+}$  studies produced an extensive set of AP and  $\text{Ca}^{2+}$ -transient (CaT) biomarkers at 1 Hz pacing: AP duration (APD), computed at 20%, 50% and 90% of repolarisation (APD<sub>20</sub>, APD<sub>50</sub> and APD<sub>90</sub>, respectively), AP amplitude (AP<sub>amp</sub>), mean upstroke velocity ( $dV/dt_{\text{MEAN}}$ , computed as the mean  $dV/dt$  value during the upstroke phase), resting membrane potential (RMP), CaT time to peak (CaT<sub>tp</sub>), CaT relaxation time from peak, computed at 50% and 90% of CaT decay (T<sub>50</sub> and T<sub>90</sub>), CaT amplitude (CaT<sub>amp</sub>) and diastolic  $\text{Ca}^{2+}$  concentration ( $[\text{Ca}^{2+}]_{\text{i,dias}}$ ). In addition, voltage clamp experiments, together with mRNA and protein expression studies, were considered.

### 2.2. Baseline model

As baseline for our investigations, the endocardial version of the O'Hara-Rudy (ORd) model was used [18]. This constitutes the most sophisticated human ventricular AP model to date, developed from and extensively validated against experimental data from more than 100 non-diseased human hearts. Minor modifications were performed to

the original model in order to better reproduce the experimental non-diseased data considered in this study. A detailed description of these changes is provided in the extended methods and Supplemental Table S1.

### 2.3. Building the control population

As in Britton et al. [17], a population of non-diseased AP models accounting for biological variability was constructed, by assuming that variability is mostly caused by cell-to-cell differences in ion channel density rather than kinetics (which may be altered instead by abnormalities such as channelopathies [19]).

An initial population of 30,000 human endocardial AP models was generated, by varying a total of 11 parameters in the original model. These included the maximal conductances ( $g$ ) of the main ionic currents/pumps/exchangers characterising the human ventricular AP:  $g_{\text{Kr}}$ ,  $g_{\text{Ks}}$ ,  $g_{\text{K1}}$ ,  $g_{\text{to}}$ ,  $g_{\text{CaL}}$ ,  $g_{\text{NaL}}$ ,  $g_{\text{Na}}$ ,  $g_{\text{NCX}}$ ,  $g_{\text{NaK}}$  ( $\text{Na}^+/\text{K}^+$  pump),  $g_{\text{Rel}}$  (RyR) and  $g_{\text{Up}}$  (SERCA). All the parameters were probabilistically sampled in the [50%-150%] range with respect to their original values, by using Latin Hypercube Sampling [20].

From this initial pool of 30,000 candidate models, a calibration process was performed to select those models in agreement with the experimental non-diseased data [17]. Calibration ranges were extracted from each of the experimental AP and CaT biomarkers, by considering their minimum and maximum values. Only models within these experimental bounds, i.e. satisfying all the experimental constraints, were accepted in the final CTRL population, while the others were discarded.

Absolute  $[\text{Ca}^{2+}]_{\text{i,dias}}$  values were not used within the calibration process, since these are extracted from the conversion of fluorescence emission after removal of the background level signal [21]. Cell-to-cell offsets in background emission can hence significantly affect the indirect estimation of  $[\text{Ca}^{2+}]_{\text{i,dias}}$ , whereas other CaT biomarkers such as CaT amplitude and duration are insensitive to diastolic  $\text{Ca}^{2+}$  concentrations. We therefore reserved  $[\text{Ca}^{2+}]_{\text{i,dias}}$  data to compare diastolic  $\text{Ca}^{2+}$  levels in HCM vs CTRL cardiomyocytes, by normalising them with respect to the mean CTRL values.

### 2.4. Building the HCM population

Based on the experimental data described above, we constructed the human HCM population by applying the electrical remodelling measured in human HCM to the CTRL population. Remodelling in the different ionic currents was accounted for by scaling their corresponding conductances, based on the ratio between HCM and non-diseased data, as reported by Coppini et al. [3]. This is equivalent to shifting in mean values the distributions of peak intensities for the different affected ionic currents, as experimentally reported. Based on the voltage clamp experiments, we up-regulated  $I_{\text{NaL}}$  (+165%) and  $I_{\text{CaL}}$  (+40%), and down-regulated  $I_{\text{to}}$  (-70%) and  $I_{\text{K1}}$  (-30%), together with an increase of the fast and slow time constants of both voltage- and  $\text{Ca}^{2+}$ -dependent  $I_{\text{CaL}}$  inactivation (+35% and +20%, fast and slow respectively). Based on mRNA expression data, we modulated the  $\text{K}^+$  repolarizing currents ( $I_{\text{Kr}}$  and  $I_{\text{Ks}}$ , -45%), SERCA pump ( $J_{\text{up}}$ , -25%), RyRs release ( $J_{\text{rel}}$ , -20%) and  $\text{Na}^+/\text{Ca}^{2+}$  exchanger ( $I_{\text{NCX}}$ , +30%). Finally, we modified cell radius to reproduce the +90% increase in cell volume reported in the experiments. In the absence of specific ultrastructural analysis of the human HCM cardiomyocytes, we assumed an equal volume increase of all subcellular compartments, in agreement with the increased cell radius and marked enlargement of the SR in murine models of HCM [22].

Three additional remodelling elements were considered: i) an increased affinity of Troponin for  $\text{Ca}^{2+}$  ( $K_{\text{TRPN}}$ , -50%) [23,24]; ii)  $\text{Na}^+/\text{K}^+$  pump inhibition due to energy deficiency of ATP-consuming processes ( $I_{\text{NaK}}$ , -30%) [24,25], which markedly regulates intracellular  $\text{Ca}^{2+}$  load by  $\text{Na}^+$  regulation of the  $\text{Na}^+/\text{Ca}^{2+}$  exchanger [26]; and iii)

an increase of background  $\text{Na}^+$  current ( $I_{\text{Nab}}$ , +165% as for  $I_{\text{NaL}}$ ) [27]. A sensitivity analysis of model biomarkers for all the ionic parameters sampled in the CTRL and HCM human populations is provided in Supplemental Figure S2. For comparative purposes, the main differences in ionic remodelling between HCM and heart failure are highlighted in Supplemental Table S2.

### 2.5. Repolarisation abnormalities

All AP traces were automatically checked for repolarisation abnormalities, e.g. EADs. HCM models with abnormal repolarisation were identified, based on the following conditions: i) failure of repolarisation at the end of diastole (define as  $V_m > -65$  mV); and/or ii) showing a positive derivative of transmembrane voltage after 150 ms from AP peak. The final trace of models exhibiting repolarisation abnormalities was recorded for a full extent of 3 s. In accordance with experiments, these were subdivided into single/multiple EADs (based on the number of voltage peaks after the AP upstroke, for EADs lasting less than 3 s), and repolarisation failure (RF; EADs longer than 3 s). This latter group of abnormalities is also in agreement with independent experimental observations of long-lasting membrane potential oscillations after EADs onset due to slow repolarisation [28]. Matching experiments, models showing repolarisation abnormalities were not considered in the computation of mean AP and CaT biomarkers.

### 2.6. Current blocks

Based on our analysis of ionic mechanisms underlying repolarisation abnormalities in human HCM models, *in silico* studies were conducted to investigate the effects of inhibition of inward currents during the phase 2 of the human AP ( $I_{\text{NaL}}$ ,  $I_{\text{CaL}}$  and  $I_{\text{NCX}}$ ). This was achieved by reducing their respective maximal ion channel conductances, for all models within the HCM population. Single and multi-channel strategies, from 10% to 60% current block, were considered. Larger current blocks were not considered in order to avoid secondary effects due to almost complete repression of the respective ionic currents.

### 2.7. Simulation details

All models were paced at 1 Hz until steady state (500 s), to ensure that intracellular  $\text{Na}^+$  and  $\text{K}^+$  concentrations were stable over time. As in the experiments, the last ten AP and CaT traces for each model were stored and used to compute average AP and CaT biomarkers. Numerical simulations and biomarkers evaluation were performed using Chaste [29]. Post-processing of AP and CaT traces, together with data analysis, were performed in Matlab (Mathworks, Inc.).

## 3. Results

### 3.1. The ionic remodelling and abnormal $\text{Ca}^{2+}$ handling recover the main hallmarks of the human HCM electrophysiological phenotype

As a result of the experimentally-driven calibration, the human ventricular CTRL population consists of 9118 models out of the initial pool of 30,000, qualitatively and quantitatively in agreement with the experimental recordings in human non-diseased cells, and accounting for biological variability. Fig. 1A illustrates the distribution of AP and CaT biomarkers in the non-diseased population: only models for which all AP and CaT biomarkers were within the experimental bounds were accepted in the final population, while others were discarded. Fig. 1B shows representative AP and CaT traces for both accepted (blue) and discarded (light grey) models. The baseline model used to generate the population has been highlighted in white.

By applying the experimentally-recorded remodelling in HCM to the CTRL population (see Methods), the human HCM population was obtained. This population qualitatively and quantitatively reproduces the

HCM electrophysiological phenotype, as shown by the AP and CaT biomarkers comparison in Fig. 2A. In agreement with the experiments, HCM models are characterised by prolonged AP, reduced upstroke velocity and slightly increased  $\text{AP}_{\text{amp}}$ . CaT is prolonged as well, together with a decrease in  $\text{CaT}_{\text{amp}}$  and an increase in  $[\text{Ca}^{2+}]_{\text{i,dias}}$ .

The main differences between HCM (pink) and CTRL (blue) are summarised in Fig. 2B, showing representative AP and CaT traces from the two populations. To facilitate the comparison, the baseline CTRL model and its corresponding HCM counterpart have been highlighted in white and black, respectively. Within the HCM population, approximately 8% of the models exhibited repolarisation abnormalities. As in the experiments, AP and CaT biomarkers of these models were not considered for the biomarkers comparison presented above, and are analysed in detail below.

### 3.2. $I_{\text{CaL}}$ re-activation underlies repolarisation abnormalities in human HCM

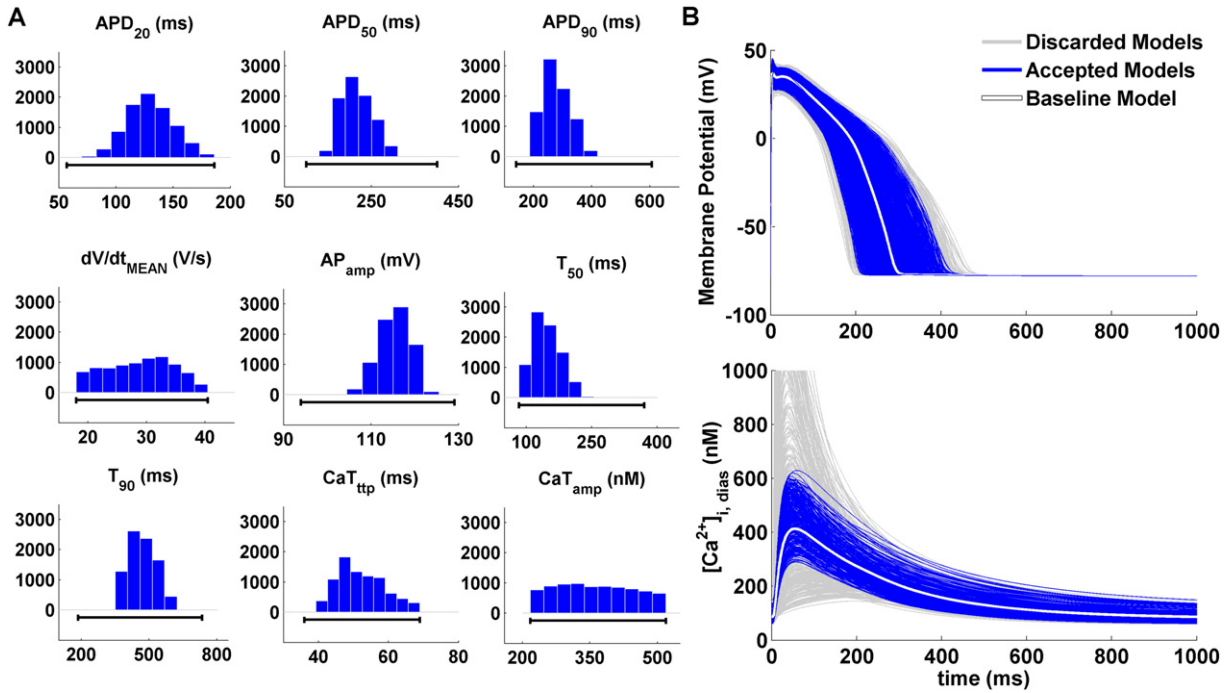
Repolarisation abnormalities were detected in 752 out of 9118 HCM models, and classified as single/multiple EADs (480 and 201 models, respectively) and repolarisation failure (RF, 71 models). Representative experimental and *in silico* AP traces from each of these subgroups are presented in Fig. 3A, highlighting the similarities between simulations and experimental recordings. Fig. 3B shows the distributions of ionic properties for each of the HCM repolarisation abnormalities subgroups, compared to the models with a normal AP, i.e. exhibiting a regular repolarisation phase. All models affected by repolarisation abnormalities were characterised by markedly low  $I_{\text{Kr}}$  conductances. In these subgroups, models were also characterised by decreasing conductances of  $I_{\text{Ks}}$ ,  $I_{\text{K1}}$  and  $I_{\text{NaK}}$  currents, which further contributed to compromise the repolarisation reserve [30]. In addition, all inward currents during the phase 2 of the AP ( $I_{\text{CaL}}$ ,  $I_{\text{NaL}}$  and  $I_{\text{NCX}}$ ) were up-regulated in HCM models displaying repolarisation abnormalities. Finally,  $\text{Ca}^{2+}$  uptake through SERCA and  $\text{Ca}^{2+}$  release through RyRs were moderately reduced compared to HCM models with normal repolarisation.

Fig. 4 further illustrates the ionic mechanisms underlying repolarisation abnormalities in HCM models. Single/multiple EADs and RF were consistently led by  $I_{\text{CaL}}$  re-activation in all 752 abnormal HCM models (Fig. 4, left column). The compromised repolarisation reserve kept the membrane depolarised long enough to allow for the re-opening of the L-type  $\text{Ca}^{2+}$  channels, as confirmed by the  $I_{\text{CaL}}$  activation gate traces (Fig. 4, left column, second row). Simulation results did not identify any re-activation of the  $\text{Na}^+$  gates or any significant release of  $\text{Ca}^{2+}$  from the sarcoplasmic reticulum (Fig. 4, left column, fourth and last rows).

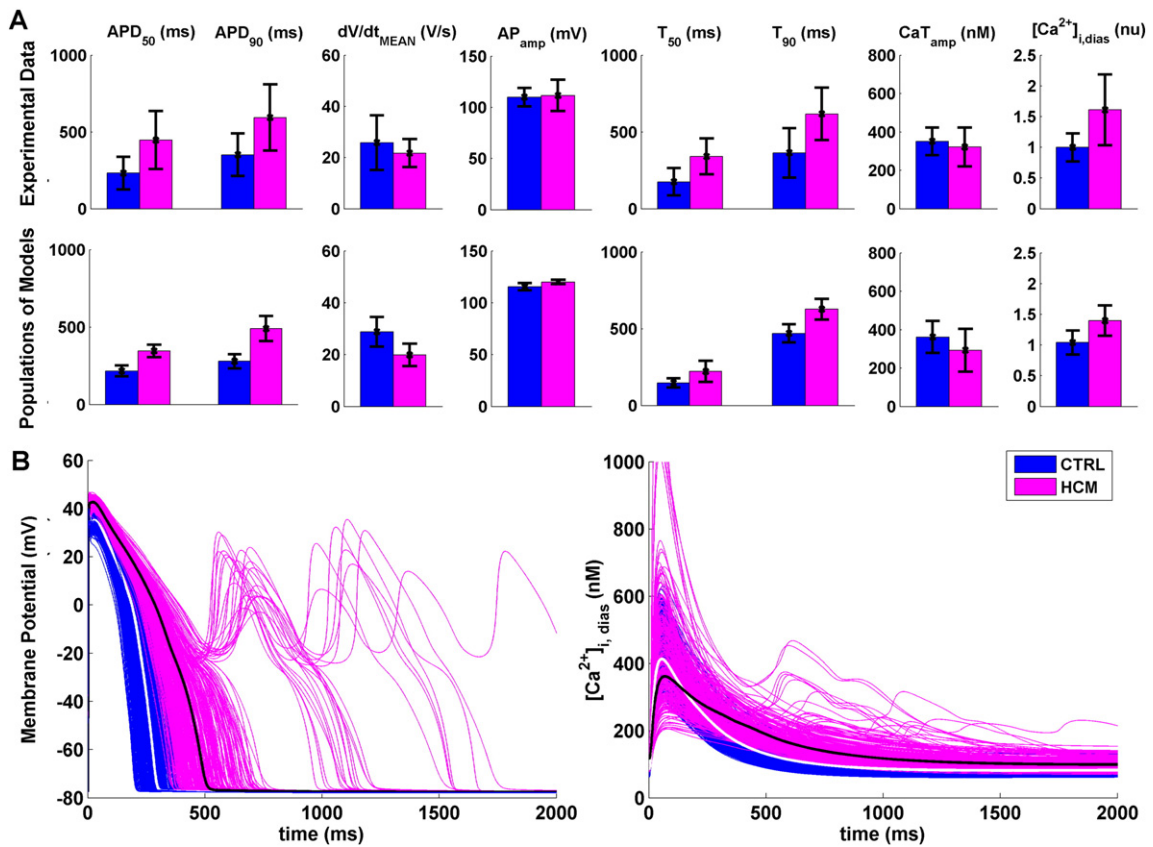
In order to corroborate these findings, additional simulations were performed artificially impeding the re-activation of L-type  $\text{Ca}^{2+}$  channels. This consistently eliminated all repolarisation abnormality types in all 752 abnormal HCM models (Fig. 4, middle column). Finally, to confirm the role played by the weak repolarisation reserve in leading to  $I_{\text{CaL}}$  re-activation, we considered additional simulation results obtained when eliminating the  $I_{\text{Kr}}$  remodelling associated to the human HCM phenotype. This again abolished all the observed repolarisation abnormalities within the remodelled HCM population (Fig. 4, right column).

### 3.3. Selective block of inward currents as a potential therapy in human HCM

Based on the analysis of the ionic mechanisms underlying repolarisation abnormalities in human HCM, we identified the selective blockade of the three main inward currents during the AP plateau phase ( $I_{\text{CaL}}$ ,  $I_{\text{NaL}}$  and  $I_{\text{NCX}}$ ) as potential anti-arrhythmic strategies in this disease. Indeed, the HCM models exhibiting repolarisation abnormalities were characterised by high conductances for these inward currents. This contributed to further reduce the repolarisation reserve, already compromised as a consequence of the reduced  $I_{\text{Kr}}$  associated to HCM



**Fig. 1.** Experimentally-calibrated population of human ventricular non-diseased cell models. A: AP and CaT biomarker distributions for the 9118 models accepted in the CTRL population, all in agreement with the experimental ranges (black lines). Histograms show the number of models for each of the bins. B: Representative AP and CaT traces for cell models accepted in the final CTRL population (blue) and discarded ones (light grey). The baseline model traces have been highlighted in white.



**Fig. 2.** Comparison of the HCM population of human ventricular cell models (pink) vs CTRL (blue). A: Bar plots show AP and CaT biomarkers distribution in HCM vs CTRL for experiments (first row; AP biomarkers:  $n = 19$  for CTRL,  $n = 57$  for HCM; CaT biomarkers:  $n = 12$  for CTRL,  $n = 22$  for HCM) and populations of models (second row,  $n = 8366$ ). Data are presented as mean  $\pm$  SD. B: Representative AP and CaT traces of HCM and CTRL models. The action potential for the baseline CTRL and HCM models have been highlighted in white and black, respectively.

remodelling, leading to a large APD prolongation and thus favouring  $I_{CaL}$  re-activation.

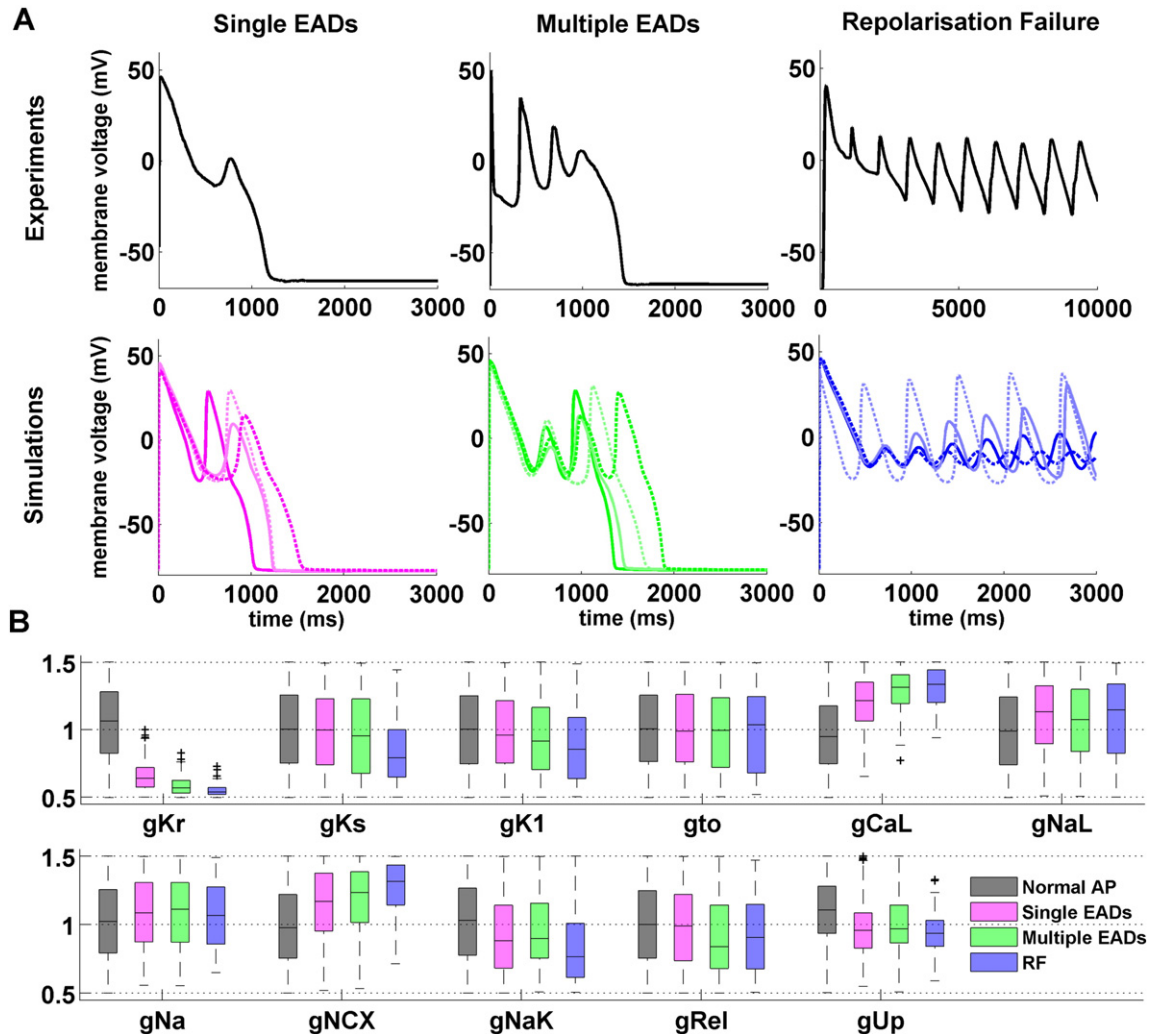
Simulation studies were conducted to investigate the effect of the selective block of these currents. Their therapeutic potency was evaluated by monitoring the abolishment of repolarisation abnormalities in the human HCM population, as well as their capability of normalising AP and CaT biomarkers in the rest of the HCM population (Fig. 5A and B, respectively).

The three current blocks were effective in reducing repolarisation abnormalities, especially single EADs (Fig. 5A, solid lines). Their efficacy was however lower in suppressing abnormalities under conditions of a highly compromised repolarisation reserve (Fig. 5A, dashed and thin solid lines for multiple EADs and RF, respectively).  $I_{CaL}$  block proved as the most effective option: indeed, 30% of  $I_{CaL}$  block sufficed to suppress all single EADs, and with a 60%  $I_{CaL}$  block all repolarisation abnormalities are abolished.  $I_{NaL}$  block significantly reduced the occurrence of single EADs (more than 95% with 60% block), but its effect was smaller for multiple EADs (20%) and null for RF. The efficacy of  $I_{NCX}$  block was higher than the one of  $I_{NaL}$ , succeeding in suppress more than 95% of single EADs as well, but up to 60% of multiple EADs and 20% of RF instances. None of the three selective inward current blocks elicited additional repolarisation abnormalities in the rest of the human HCM population.

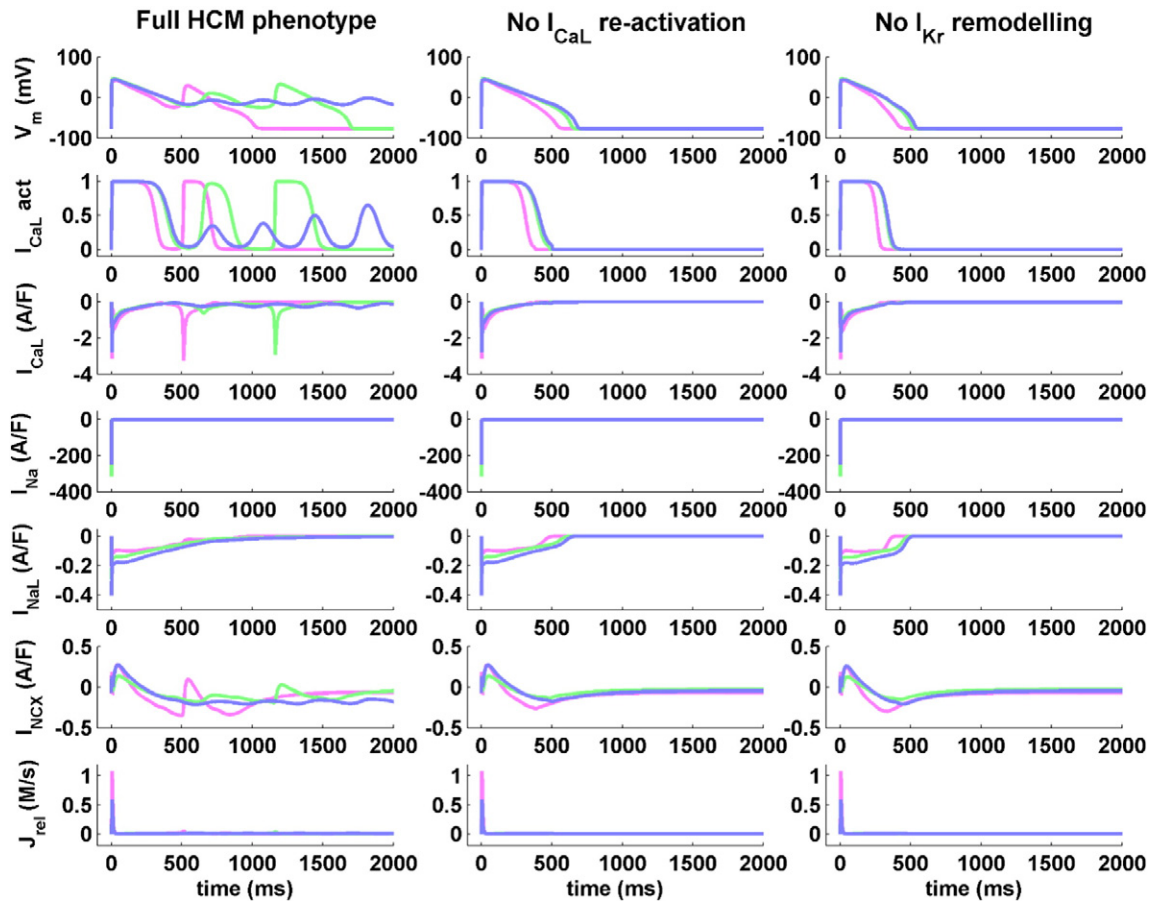
Regarding the normalisation of AP biomarkers, all the selective current blocks partially reversed the AP prolongation occurring in HCM cardiomyocytes (Fig. 5B, first column). The magnitude of APD reduction relative to the HCM population was similar for the three considered current blocks (i.e., APD<sub>90</sub> decrease of -7.8%, -9.2% and -8.6% for 60% of  $I_{CaL}$ ,  $I_{NaL}$  and  $I_{NCX}$  block, respectively).

Both  $I_{CaL}$  and  $I_{NaL}$  blocks reduced  $[Ca^{2+}]_{i,dias}$  (Fig. 5B, second column), one of the hallmarks induced by HCM remodelling. However,  $I_{CaL}$  blockade yielded a more remarkable reduction in diastolic  $Ca^{2+}$  load (-19.3% with 60%  $I_{CaL}$  block) than  $I_{NaL}$ , which had a smaller effect (-1.8% with 60%  $I_{NaL}$  block). On the contrary, blocking  $I_{NCX}$  led to an increase in  $[Ca^{2+}]_{i,dias}$  (+27% with 60%  $I_{NCX}$  block), thus potentially aggravating the HCM phenotype. A similar trend was observed for the CaT amplitude (Fig. 5B, third column), which decreased when blocking  $I_{CaL}$  and  $I_{NaL}$  (-82% and -14%, with 60%  $I_{CaL}$  and  $I_{NaL}$  block, respectively), and highly increased when blocking  $I_{NCX}$  (+105% with 40%  $I_{NCX}$  block). Therefore, both  $I_{NaL}$  and  $I_{CaL}$  blocks have a negative effect on CaT<sub>amp</sub>, already reduced in HCM, while  $I_{NCX}$  block seems to counteract this aspect of HCM remodelling.

To summarise our findings,  $I_{CaL}$  block was identified as the most effective strategy to suppress repolarisation abnormalities in HCM, as well as to revert the increase in APD and  $[Ca^{2+}]_{i,dias}$  induced by HCM



**Fig. 3.** Repolarisation abnormalities in human HCM models ( $n = 752$ ). A: Representative experimental (top) and simulated (bottom) HCM action potential traces, showing different types of repolarisation abnormalities. From left to right: single EADs, multiple EADs and repolarisation failure (RF). B: Normalised distributions of ionic properties for the 11 conductances varied within the population, for models displaying normal AP ( $n = 8366$ ), single/multiple EADs ( $n = 480/201$ ) and RF ( $n = 71$ ). On each box, the central mark is the median, box limits are the 25th and 75th percentiles, and whiskers extend to the most extreme data points not considered outliers, plotted individually as separate crosses.



**Fig. 4.**  $I_{CaL}$  re-activation underlies repolarisation abnormalities in human HCM models. From top to bottom, representative traces for AP,  $I_{CaL}$  activation, transmembrane inward currents ( $I_{CaL}$ ,  $I_{Na}$ ,  $I_{NaL}$  and  $I_{NCX}$ ) and  $Ca^{2+}$  release from the sarcoplasmic reticulum ( $J_{rel}$ ) are shown (left column). Selected HCM models are presented from the three repolarisation abnormalities subgroups: single EADs (pink), multiple EADs (green) and RF (blue). Simulation results are shown for the same models when inhibiting L-type  $Ca^{2+}$  channels re-opening (middle column), as well as when eliminating the  $I_{Kr}$  remodelling induced by HCM (right column). In both cases, repolarisation abnormalities are completely abolished.

remodelling. However, large  $I_{CaL}$  inhibition may severely compromise  $CaT_{amp}$ .  $I_{NaL}$  block is able to successfully suppress single EADs, but has a very low efficacy on multiple EADs and RF. It also has a positive effect by reducing both APD and  $[Ca^{2+}]_{i,dias}$ , with a small impact on  $CaT_{amp}$ . Finally,  $I_{NCX}$  block reduces repolarisation abnormalities, especially single EADs, and also shortens APD. However, it has a high impact on CaT, especially on  $CaT_{amp}$ , and further increases the already elevated  $[Ca^{2+}]_{i,dias}$  observed in human HCM cells.

### 3.4. $Na^+/Ca^{2+}$ exchanger block improves the efficacy of $I_{NaL}$ and $I_{CaL}$ selective blocks

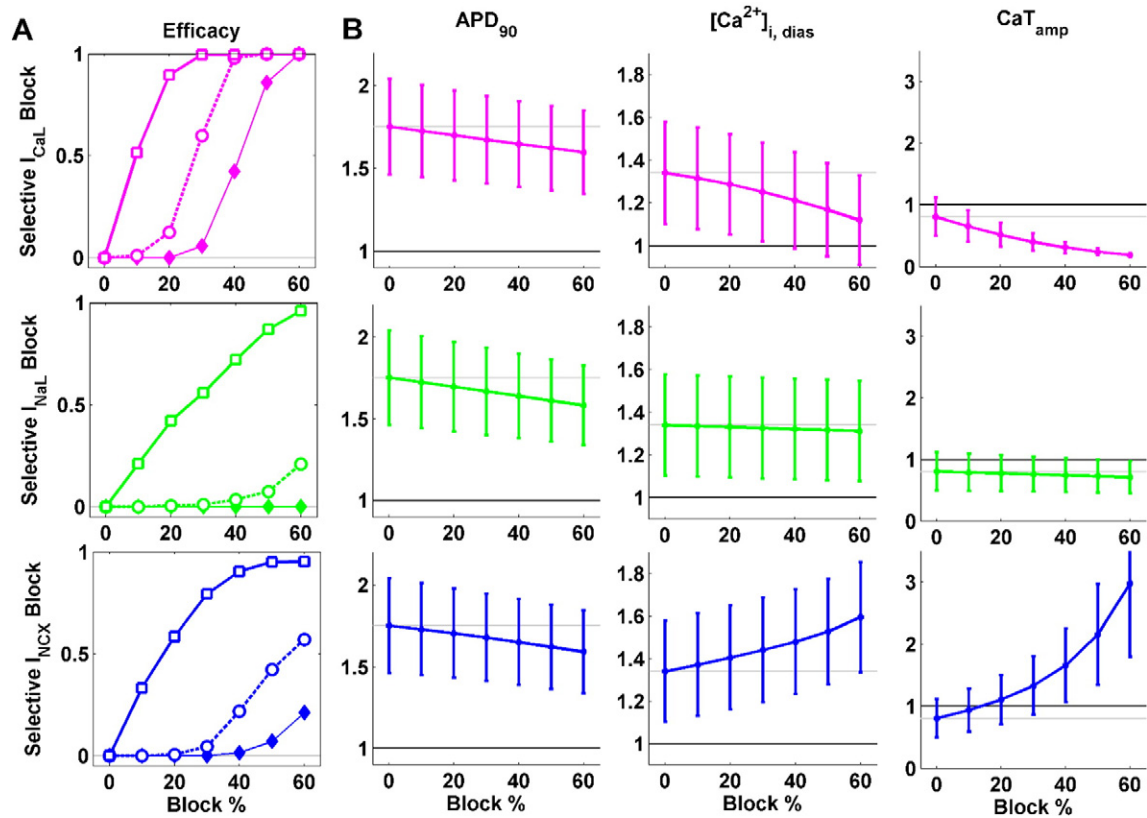
Simulation results for selective inward current blocks pointed out  $CaT_{amp}$  decrease as the main drawback when blocking  $I_{CaL}$  and  $I_{NaL}$ . Since  $I_{NCX}$  block produces instead an increase in  $CaT_{amp}$ , the potential benefit of multichannel block therapies were additionally investigated, by adding  $I_{NCX}$  blockade to both the selective  $I_{CaL}$  and  $I_{NaL}$  blocks. Due to the high number of possible combinations and number of models within the human HCM population ( $n > 9000$ ), results are presented for a double multichannel block strategy for each pair of currents, based on the findings on selective current block results presented in the previous section. As for  $I_{CaL}$ , we considered the 40% current block, which yields positive efficacy (100/98% for single/multiple EADs) without excessively compromising the CaT amplitude. As for  $I_{NaL}$ , as  $CaT_{amp}$  reduction was overall of small magnitude, we considered the maximum block (60%). We combined these two scenarios with  $I_{NCX}$  block (40% and

20%, respectively), aiming at balancing the changes in both  $CaT_{amp}$  and  $[Ca^{2+}]_{i,dias}$ .

Both multichannel approaches increased the efficacy of the single channel blocks in abolishing repolarisation abnormalities, including single/multiple EADs and RF. Blockade of 40% and 20% of  $I_{NCX}$  augmented the efficacy of  $I_{CaL}$  and  $I_{NaL}$  blocks in +5% and +18%, respectively (Fig. 6A). In terms of the normalisation of AP biomarkers,  $I_{NCX}$  block further contributed to AP shortening in both multichannel combinations (Fig. 6B, first column), even under already significant APD decrease due to large  $I_{NaL}$  inhibition. As for  $[Ca^{2+}]_{i,dias}$  (Fig. 6B, second column), the increase in diastolic  $Ca^{2+}$  load induced by  $I_{NCX}$  block was not fully compensated by the other single channel block actions: as a result,  $[Ca^{2+}]_{i,dias}$  distributions were comparable to those in the baseline HCM population. As for the CaT amplitude (Fig. 6B, last column), the large reduction in  $CaT_{amp}$  due to  $I_{CaL}$  block was not fully counteracted by additional  $I_{NCX}$  block: the final  $CaT_{amp}$  was still ~100 nM lower than for the HCM population without any current block. On the contrary, when combining  $I_{NaL}$  and  $I_{NCX}$ , the final  $CaT_{amp}$  was increased, and almost restored to that for the non-diseased population.

## 4. Discussion

In this study, we unravel the key ionic mechanisms underlying repolarization and  $Ca^{2+}$  handling abnormalities in human ventricular cardiomyocytes in HCM, and we identify potentially-efficacious anti-arrhythmic strategies specific to the HCM phenotype. Our synergistic



**Fig. 5.** *In silico* evaluation of selective inward current block in human HCM. Each row shows the effect of a single current block:  $I_{CaL}$  (top, magenta),  $I_{NaL}$  (middle, green) and  $I_{NCX}$  (bottom, blue). A: Anti-arrhythmic efficacy of single channel blocks, evaluated as the fraction of repolarisation abnormalities successfully suppressed in the human HCM population. The efficacy is shown separately for each subgroup of repolarisation abnormalities: single EADs (thick solid, white squares), multiple EADs (thick dashed, white circles) and RF (thin solid, full diamonds). B: Effects of blocking each ionic current in reversing APD<sub>90</sub>, [Ca<sup>2+</sup>]<sub>i,dias</sub> and CaT<sub>amp</sub> changes in the human HCM phenotype. Data are presented as mean  $\pm$  SD, normalised with respect to the corresponding CTRL means. CTRL and HCM mean values are shown as black and grey lines, respectively, to facilitate comparison.

approach tightly couples populations of models of human ventricular electrophysiology with a rich experimental dataset, including action potential, calcium transient, ionic current and mRNA measurements obtained in human HCM cardiomyocytes. We consistently found the reactivation of the overexpressed L-type Ca<sup>2+</sup> current, favoured by a decreased repolarisation reserve, as the key mechanism responsible for repolarisation abnormalities in HCM. These findings were subsequently integrated in *in silico* studies of single and multichannel ion block, aiming at suggesting potential strategies to ameliorate the main hallmarks of the electrophysiological phenotype of human HCM. In spite of exhibiting a high efficacy in the suppression of pro-arrhythmic abnormalities, selective  $I_{CaL}$  block also markedly compromised Ca<sup>2+</sup> transient amplitude in human HCM. Multichannel  $I_{NCX}$ ,  $I_{NaL}$  and  $I_{CaL}$  blocks, previously unexplored in the pharmacological management of the disease, exhibited increased efficacy than the respective single blocks, also mitigating adverse effects on systolic Ca<sup>2+</sup> function.

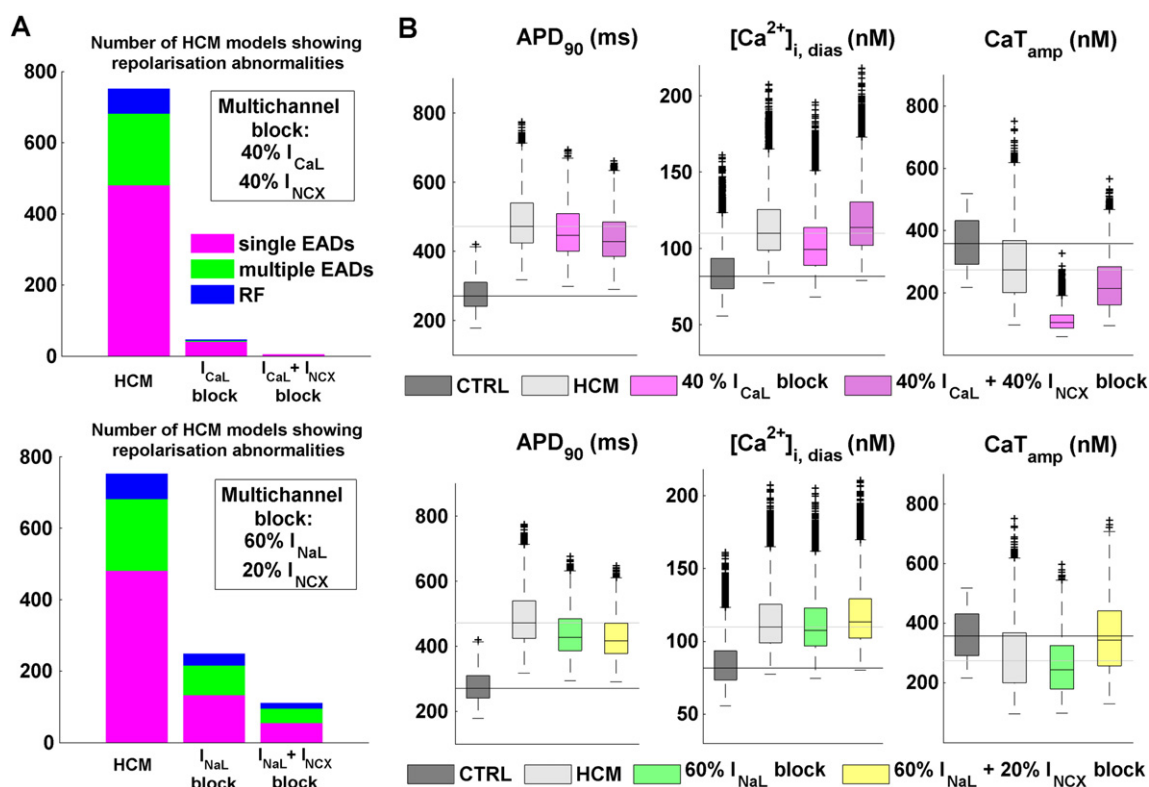
In order to address the significant inter-subject variability and disease expression exhibited in the disease, experimentally-calibrated populations of models of human ventricular electrophysiology under non-diseased and HCM conditions were constructed based on the most comprehensive electrophysiological and molecular characterisation of HCM to date in human [3]. The two populations qualitatively and quantitatively reproduce the distinctive characteristics of the non-diseased and HCM phenotypes, and in particular the distinctive characteristics associated to human HCM, including the drastic prolongation of AP duration at different levels of cellular repolarisation, increased diastolic Ca<sup>2+</sup> load and decreased Ca<sup>2+</sup> transient amplitude with slower kinetics, together with a propensity to develop repolarisation abnormalities.

The population of models approach [17,31–33] is particularly suitable for exploring the effect of inter-subject variability in the disease, especially when evaluating potential anti-arrhythmic therapies, whose efficacy may depend on individual responses to drug actions [34]. Furthermore, this is the first time that this methodology has been applied to the study of a genetic cardiomyopathy.

Here, instead of a single AP model representative of average cell behaviour, cell-to-cell differences are accounted for by varying ion channel parameters around their nominal values. These parameters are the maximal conductances of the main ionic currents, based on the assumption that variability is mostly depending on the difference in the number of ionic channels from cell to cell, as recently suggested also by others [19,34–36]. Calibration of model outputs against experimental data is then performed in order to only retain models within physiological range. In this regard, each model hence becomes a representation of a viable cell within the plausible bounds of biological variability observed in the population [17,31–33].

The HCM phenotype was recovered in this study based on the available experimental data, by modifying the conductances of the main ionic channel affected by the disease together with the inactivation of the L-type Ca<sup>2+</sup> current. Additional elements of the electrophysiological remodelling associated to HCM can be easily accommodated to the presented approach, shall the experimental evidence becomes available.

The experimental recordings also provided confirmation of increased proneness to repolarisation abnormalities in HCM diseased cardiomyocytes [3,37], and in particular to the development of EADs as a well-established pro-arrhythmic mechanism [12,13]. Our findings suggest that the concomitant decrease of the K<sup>+</sup> repolarising currents and the increase of inward currents during the plateau phase of the



**Fig. 6.** *In silico* evaluation of combined inward channel blocks in human HCM. Simulation results are shown for two different multichannel alternatives:  $I_{CaL} + I_{NCX}$  (top row), and  $I_{NaL} + I_{NCX}$  (bottom row). A: Efficacy of multichannel strategies in suppressing repolarisation abnormalities, compared to their respective selective current blocks. Bar plots illustrate the number of models exhibiting single/multiple EADs and RF in each considered scenario. B: Effects of multichannel block in reversing APD<sub>90</sub>,  $[Ca^{2+}]_{i,dias}$  and CaT<sub>amp</sub> changes in the human HCM phenotype. CTRL and HCM reference values are also presented to facilitate comparison. Boxplot descriptions as in Fig. 3.

AP, i.e. an impaired repolarisation reserve [30], is the provenance of such repolarisation abnormalities in human HCM. The marked AP prolongation induced by the combined action of these two factors keeps the membrane depolarised long enough to allow for the re-opening of the L-type  $Ca^{2+}$  channels. The mechanistic insights on EADs generation presented in this contribution, specific of human ventricular electrophysiology under HCM diseased conditions, are therefore in agreement with previous experimental and theoretical studies in different animal species [38,39]. Other possible factors, such as the reopening of  $Na^+$  channels, has been also associated with EADs triggering [12,40]. Our *in silico* results, analysed in ~800 human ventricular models exhibiting different severities of repolarisation abnormalities, consistently identified  $I_{CaL}$  re-activation as the only source of triggered activity in human HCM.

Emerging from these results, we identified the targeting of net inward currents ( $I_{CaL}$ ,  $I_{NaL}$  and  $I_{NCX}$ ) as potential therapeutic strategies to counteract AP prolongation in HCM. We evaluated *in silico* their selective block, considering both their anti-arrhythmic implications and their contribution in reversing the HCM phenotype. The selective block of  $I_{CaL}$  proved as highly effective in suppressing all repolarisation abnormalities, and also in reducing APD and lowering the diastolic  $Ca^{2+}$  load. However, our simulation results also show how  $I_{CaL}$  block markedly affects the already compromised  $Ca^{2+}$  transient amplitude in human HCM (Fig. 5). This provides additional mechanistic and quantitative insights into the interpretation of the existing guidelines for the management of HCM [41], which suggest caution in the use of  $Ca^{2+}$  blockers (e.g. verapamil) in the treatment of the disease [5].

Selective  $I_{NaL}$  block successfully suppressed single EADs at high levels of current block, but exhibited small efficacy on more severe repolarisation abnormalities. It also contributed to a decrease in APD and to a smaller extent in diastolic  $Ca^{2+}$  load, without any noticeable effect on  $Ca^{2+}$  transient amplitude. Our findings are hence in agreement

with the reported reduction of EADs in HCM cells and the partial reversal of the HCM phenotype under pharmacological  $I_{NaL}$  inhibition with ranolazine [3,42]. In addition, highly selective  $I_{NaL}$  blockers are currently being developed and may be available in the near future [43], which together with ongoing clinical trials to assess the effects of ranolazine in HCM patients, will allow for a systematic validation of our prospective *in silico* predictions.

In the HCM population, selective  $I_{NCX}$  block had an anti-arrhythmic effect, especially by reducing single EADs and shortening the AP. These results are in agreement with the reported potential of  $I_{NCX}$  block in suppressing EADs and preventing  $Ca^{2+}$  overload-induced triggered arrhythmias in canine and guinea pig studies [44–46]. To facilitate the interpretation of our results, Table 1 summarises the ionic currents investigated in our selective channel block simulation study, drugs targeting them, and their previously reported use in human HCM.

Our *in silico* predictions of combined multichannel  $I_{NCX}$  and either  $I_{NaL}$  or  $I_{CaL}$  blocks in HCM indicate an increase in the efficacy of suppression of repolarisation abnormalities and of the shortening of the AP with respect to the individual selective strategies. On the other hand, our results also show that  $I_{NCX}$  block may have a significant impact on  $Ca^{2+}$  handling, by increasing the  $Ca^{2+}$  transient amplitude and further contributing to diastolic  $Ca^{2+}$  overload. These findings are hence supported by those of Bourgonje et al. [28], showing a combined  $I_{CaL}$  and  $I_{NCX}$  block by SEA-0400 as an effective anti-arrhythmic strategy against dofetilide-induced arrhythmias, despite an increase in diastolic calcium content. Their results were however obtained in a canine chronic atrioventricular block model for compensated hypertrophy, not representative of the ionic remodelling of human HCM. Our study therefore constitutes the first investigation of a multichannel  $I_{NCX}$  strategy for the pharmacological management of human HCM, where the arrhythmic mechanisms may be different.



In HCM myocardium, alterations of  $\text{Ca}^{2+}$  handling (e.g. reduced SERCA function) may render the cardiomyocyte more dependent on  $I_{\text{NCX}}$  for  $\text{Ca}^{2+}$  transient decay. Additionally, the increased forward mode activity of the  $\text{Na}^+/\text{Ca}^{2+}$  exchanger appears to be crucial to maintain CaT amplitude in HCM. For these reasons, the effects of  $I_{\text{NCX}}$  blockers are likely to be more pronounced in HCM as compared with healthy myocardium. In the multichannel strategies, and under the premise of not significantly raising the diastolic  $\text{Ca}^{2+}$  levels observed in HCM,  $I_{\text{NCX}}$  block only partially ameliorates the drastic decrease of  $\text{Ca}^{2+}$  transient amplitude induced by  $I_{\text{CaL}}$  blockade, whereas when combined with  $I_{\text{NaL}}$  block it successfully reverts  $\text{Ca}^{2+}$  transient amplitude to the values exhibited by the non-diseased population. However, recent experimental findings using last-generation selective  $I_{\text{NCX}}$  blockers [44–46] have reported no significant alterations in the AP or  $\text{Ca}^{2+}$  transient, in spite of  $I_{\text{NCX}}$  being considered the main responsible of  $\text{Ca}^{2+}$  extrusion in cardiac myocytes. Future research will be devoted to confirm the potential anti-arrhythmic implications of the proposed multichannel strategies for the treatment of human HCM identified in our computational studies.

A different approach to counteract for repolarisation abnormalities in human HCM, not considered in this study, could be the increase of repolarising currents in HCM cardiomyocytes. One possibility could be the use of  $I_{\text{Kr}}$  agonists to selectively activate these channels, which *in vivo* guinea pig studies with pharmacologically induced QT prolongation successfully shortened the QTc interval [47]. However, additional studies showed that these  $I_{\text{Kr}}$  agonists also impair cardiac conduction, thus impeding their use as anti-arrhythmic drugs [47]. An alternative option might be the use of ATP-sensitive  $\text{K}^+$  channel openers, as already suggested for heart failure [48]. However, the increased energy requirements of myosin ATPase and the reduction in the phosphocreatine-to-ATP ratio (an established marker of cellular energy status) in HCM [6,8] may also compromise the availability of these channels for targeting the disease.

Finally, we did not explore the role of beta-adrenergic stimulation in modulating pro-arrhythmic abnormalities in human HCM, whose ionic characterisation still remains unknown in the disease. This may be an important factor of arrhythmic risk in HCM, due to the known effect of autonomic control in modulating ionic currents, and in particular L-type  $\text{Ca}^{2+}$  channels [49]. Increased intracellular  $\text{Na}^+$  and  $\text{Ca}^{2+}$  levels can also modulate mitochondrial activity linking ATP production to ATP demand, by activation of  $\text{Na}^+/\text{Ca}^{2+}$  exchange in the inner mitochondrial membrane, which keeps mitochondrial  $\text{Ca}^{2+}$  low preventing ATP supply meeting demand [50]. Computational models of the role of the beta-adrenergic cascade on ionic currents, as well as of mitochondrial ATP synthesis, have been recently proposed [49,51–53]. These constitute exciting and promising venues for future investigations.

## Disclosures

None.

## Acknowledgements

EP, AM, ABO and BR acknowledge the financial support of the Wellcome Trust Senior Research Fellowship in Basic Biomedical Sciences (to BR) (100246/Z/12/Z).

**Table 1**

Ionic currents investigated in this study, drugs targeting them, and their previously reported use in the pharmacological management of human HCM.

Compound	Ref	Dose ( $\mu\text{M}$ )	Main channel block	Other actions	Use in HCM
Ranolazine	[43]	17 $\mu\text{M}$	50% $I_{\text{NaL}}$	$I_{\text{Kr}}$ block	[3,42]
GS967	[43]	0.13 $\mu\text{M}$	50% $I_{\text{NaL}}$	–	–
Verapamil	[54]	0.33 $\mu\text{M}$	50% $I_{\text{CaL}}$	$I_{\text{Kr}}$ block	[5,41]
ORM-10,103	[44]	0.96 $\mu\text{M}$	50% $I_{\text{NCX}}$	–	–
SEA-0400	[55]	1 $\mu\text{M}$	66% $I_{\text{NCX}}$	$I_{\text{CaL}}$ block	–

## Appendix A. Supplementary data

Supplementary data to this article can be found online at <http://dx.doi.org/10.1016/j.jmcc.2015.09.003>.

## References

- [1] B.J. Maron, Sudden death in young athletes, *N. Engl. J. Med.* 349 (2003) 1064–1075, <http://dx.doi.org/10.1056/NEJMra022783>.
- [2] B.J. Maron, M.S. Maron, Hypertrophic cardiomyopathy, *Lancet* 381 (2013) 242–255, [http://dx.doi.org/10.1016/S0140-6736\(12\)60397-3](http://dx.doi.org/10.1016/S0140-6736(12)60397-3).
- [3] R. Coppini, C. Ferrantini, L. Yao, P. Fan, M. Del Lungo, F. Stillitano, et al., Late sodium current inhibition reverses electromechanical dysfunction in human hypertrophic cardiomyopathy, *Circulation* 128 (2013), e156, <http://dx.doi.org/10.1093/chemse/bjt087>.
- [4] I. Olivetto, F. Girolami, S. Nistri, A. Rossi, L. Rega, F. Garbini, et al., The many faces of hypertrophic cardiomyopathy: from developmental biology to clinical practice, *J. Cardiovasc. Transl. Res.* 2 (2009) 349–367, <http://dx.doi.org/10.1007/s12265-009-9137-2>.
- [5] R. Spoladore, M.S. Maron, R. D'Amato, P.G. Camici, I. Olivetto, Pharmacological treatment options for hypertrophic cardiomyopathy: high time for evidence, *Eur. Heart J.* 33 (2012) 1724–1733, <http://dx.doi.org/10.1093/eurheartj/ehs150>.
- [6] H. Ashrafian, W.J. McKenna, H. Watkins, Disease pathways and novel therapeutic targets in hypertrophic cardiomyopathy, *Circ. Res.* 109 (2011) 86–96, <http://dx.doi.org/10.1161/CIRCRESAHA.111.242974>.
- [7] J.C. Tardiff, L. Carrier, D.M. Bers, C. Poggesi, C. Ferrantini, R. Coppini, et al., Targets for therapy in sarcomeric cardiomyopathies, *Cardiovasc. Res.* (2015), <http://dx.doi.org/10.1093/cvr/cvv023>.
- [8] C. O'Mahony, P. Elliott, W. McKenna, Sudden cardiac death in hypertrophic cardiomyopathy, *Circ. Arrhythm. Electrophysiol.* 6 (2013) 443–451, <http://dx.doi.org/10.1161/CIRCEP.111.962043>.
- [9] B.J. Maron, W.K. Shen, M.S. Link, A.E. Epstein, A.K. Almquist, J.P. Daubert, et al., Efficacy of implantable cardioverter-defibrillators for the prevention of sudden death in patients with hypertrophic cardiomyopathy, *N. Engl. J. Med.* 342 (2000) 365–373, <http://dx.doi.org/10.1056/NEJM200002103420601>.
- [10] J. Mogensen, R.T. Murphy, T. Kubo, A. Bahl, J.C. Moon, I.C. Klausen, et al., Frequency and clinical expression of cardiac troponin I mutations in 748 consecutive families with hypertrophic cardiomyopathy, *J. Am. Coll. Cardiol.* 44 (2004) 2315–2325, <http://dx.doi.org/10.1016/j.jacc.2004.05.088>.
- [11] R. Coppini, C. Ferrantini, L. Mazzoni, L. Sartiani, I. Olivetto, C. Poggesi, et al., Regulation of intracellular  $\text{Na}^+$  in health and disease: pathophysiological mechanisms and implications for treatment, *Glob. Cardiol. Sci. Pract.* 2013 (2013) 222–242, <http://dx.doi.org/10.5339/gcsp.2013.30>.
- [12] W.T. Clusin, Calcium and cardiac arrhythmias: DADs, EADs, and alternans, *Crit. Rev. Clin. Lab. Sci.* 40 (2003) 337–375, <http://dx.doi.org/10.1080/713609356>.
- [13] J.N. Weiss, A. Garfinkel, H.S. Karagueuzian, P.-S. Chen, Z. Qu, Early afterdepolarizations and cardiac arrhythmias, *Heart Rhythm.* 7 (2010) 1891–1899, <http://dx.doi.org/10.1016/j.hrthm.2010.09.017>.
- [14] R.F. Gilmour, Early afterdepolarization-induced triggered activity: initiation and reinitiation of reentrant arrhythmias, *Heart Rhythm.* 1 (2004) 449–450, <http://dx.doi.org/10.1016/j.hrthm.2004.07.005>.
- [15] P. Volders, Progress in the understanding of cardiac early afterdepolarizations and torsades de pointes: time to revise current concepts, *Cardiovasc. Res.* 46 (2000) 376–392, [http://dx.doi.org/10.1016/S0008-6363\(00\)00022-5](http://dx.doi.org/10.1016/S0008-6363(00)00022-5).
- [16] B. Szabo, T. Kovacs, R. Lazzara, Role of calcium loading in early afterdepolarizations generated by  $\text{Cs}^+$  in canine and guinea pig purkinje fibers, *J. Cardiovasc. Electrophysiol.* 6 (1995) 796–812, <http://dx.doi.org/10.1111/j.1540-8167.1995.tb00356.x>.
- [17] O.J. Britton, A. Bueno-Orovio, K. Van Ammel, H.R. Lu, R. Toward, D.J. Gallacher, et al., Experimentally calibrated population of models predicts and explains intersubject variability in cardiac cellular electrophysiology, *Proc. Natl. Acad. Sci. U. S. A.* 110 (2013) E2098–E2105, <http://dx.doi.org/10.1073/pnas.1304382110>.
- [18] T. O'Hara, L. Virág, A. Varró, Y. Rudy, Simulation of the undiseased human cardiac ventricular action potential: model formulation and experimental validation, *PLoS Comput. Biol.* 7 (2011), e1002061, <http://dx.doi.org/10.1371/journal.pcbi.1002061>.
- [19] W. Groenendaal, F. Ortega, A., Kherlopian AR, Zygmunt AC, Krogh-Madsen T, Christini Dj, Cell-specific cardiac electrophysiology models, *PLoS Comput. Biol.* 11 (2015), e1004242, <http://dx.doi.org/10.1371/journal.pcbi.1004242>.
- [20] M.D. McKay, R.J. Beckman, W.J. Conover, A comparison of three methods for selecting values of input variables in the analysis of output from a computer code, *Technometrics* 21 (1979) 239, <http://dx.doi.org/10.2307/1268522>.
- [21] M.D. Bootman, K. Rietdorf, T. Collins, S. Walker, M. Sanderson,  $\text{Ca}^{2+}$ -sensitive fluorescent dyes and intracellular  $\text{Ca}^{2+}$  imaging, *Cold Spring Harb. Protoc.* 2013 (2013) 83–99, <http://dx.doi.org/10.1101/pdb.top066050>.
- [22] L. Leatherbury, Q. Yu, B. Chatterjee, D.L. Walker, Z. Yu, X. Tian, et al., A novel mouse model of X-linked cardiac hypertrophy, *Am. J. Physiol. Heart Circ. Physiol.* 294 (2008) H2701–H2711, <http://dx.doi.org/10.1152/ajpheart.00160.2007>.
- [23] P. Robinson, P.J. Griffiths, H. Watkins, C.S. Redwood, Dilated and hypertrophic cardiomyopathy mutations in troponin and alpha-tropomyosin have opposing effects on the calcium affinity of cardiac thin filaments, *Circ. Res.* 101 (2007) 1266–1273, <http://dx.doi.org/10.1161/CIRCRESAHA.107.156380>.
- [24] H. Watkins, H. Ashrafian, C. Redwood, Inherited cardiomyopathies, *N. Engl. J. Med.* 364 (2011) 1643–1656, <http://dx.doi.org/10.1093/bja/aes327>.
- [25] H. Ashrafian, C. Redwood, E. Blair, H. Watkins, Hypertrophic cardiomyopathy: a paradigm for myocardial energy depletion, *Trends Genet.* 19 (2003) 263–268, [http://dx.doi.org/10.1016/S0168-9525\(03\)00081-7](http://dx.doi.org/10.1016/S0168-9525(03)00081-7).

- [26] A. Bueno-Orovio, C. Sánchez, E. Pueyo, B. Rodriguez, Na/K pump regulation of cardiac repolarization: insights from a systems biology approach, *Pflügers Arch.* 466 (2014) 183–193, <http://dx.doi.org/10.1007/s00424-013-1293-1>.
- [27] S. Morotti, A.G. Edwards, A.D. McCulloch, D.M. Bers, E. Grandi, A novel computational model of mouse myocyte electrophysiology to assess the synergy between Na<sup>+</sup> loading and CaMKII, *J. Physiol.* 592 (2014) 1181–1197, <http://dx.doi.org/10.1113/jphysiol.2013.266676>.
- [28] G.-X. Liu, B.-R. Choi, O. Ziv, W. Li, E. de Lange, Z. Qu, et al., Differential conditions for early after-depolarizations and triggered activity in cardiomyocytes derived from transgenic LQT1 and LQT2 rabbits, *J. Physiol.* 590 (2012) 1171–1180, <http://dx.doi.org/10.1113/jphysiol.2011.218164>.
- [29] J. Pitt-Francis, P. Pathmanathan, M.O. Bernabeu, R. Bordas, J. Cooper, A.G. Fletcher, et al., Chaste: a test-driven approach to software development for biological modelling, *Comput. Phys. Commun.* 180 (2009) 2452–2471, <http://dx.doi.org/10.1016/j.cpc.2009.07.019>.
- [30] D.M. Roden, Repolarization reserve: a moving target, *Circulation* 118 (2008) 981–982, <http://dx.doi.org/10.1161/CIRCULATIONAHA.108.798918>.
- [31] P. Gemmell, K. Burrage, B. Rodriguez, T.A. Quinn, Population of computational rabbit-specific ventricular action potential models for investigating sources of variability in cellular repolarisation, *PLoS One* 9 (2014), e90112, <http://dx.doi.org/10.1371/journal.pone.0090112>.
- [32] C. Sánchez, A. Bueno-Orovio, E. Wettwer, S. Loose, J. Simon, U. Ravens, et al., Inter-subject variability in human atrial action potential in sinus rhythm versus chronic atrial fibrillation, *PLoS One* 9 (2014), e105897, <http://dx.doi.org/10.1371/journal.pone.0105897>.
- [33] J. Walmsley, J.F. Rodriguez, G.R. Mirams, K. Burrage, I.R. Efimov, B. Rodriguez, MRNA expression levels in failing human hearts predict cellular electrophysiological remodeling: a population-based simulation study, *PLoS One* 8 (2013), e56359, <http://dx.doi.org/10.1371/journal.pone.0056359>.
- [34] A.X. Sarkar, E.A. Sobie, Quantification of repolarization reserve to understand interpatient variability in the response to proarrhythmic drugs: a computational analysis, *Heart Rhythm* 8 (2011) 1749–1755, <http://dx.doi.org/10.1016/j.hrthm.2011.05.023>.
- [35] A.X. Sarkar, D.J. Christini, E.A. Sobie, Exploiting mathematical models to illuminate electrophysiological variability between individuals, *J. Physiol.* 590 (2012) 2555–2567, <http://dx.doi.org/10.1113/jphysiol.2011.223313>.
- [36] A.X. Sarkar, E.A. Sobie, Regression analysis for constraining free parameters in electrophysiological models of cardiac cells, *PLoS Comput. Biol.* 6 (2010), e1000914, <http://dx.doi.org/10.1371/journal.pcbi.1000914>.
- [37] J.T. Vermeulen, M.A. McGuire, T. Opthof, R. Coronel, J.M. de Bakker, C. Klöpping, et al., Triggered activity and automaticity in ventricular trabeculae of failing human and rabbit hearts, *Cardiovasc. Res.* 28 (1994) 1547–1554.
- [38] C.T. January, J.M. Riddle, Early afterdepolarizations: mechanism of induction and block. A role for L-type Ca<sup>2+</sup> current, *Circ. Res.* 64 (1989) 977–990.
- [39] J. Zeng, Y. Rudy, Early afterdepolarizations in cardiac myocytes: mechanism and rate dependence, *Biophys. J.* 68 (1995) 949–964, [http://dx.doi.org/10.1016/S0006-3495\(95\)80271-7](http://dx.doi.org/10.1016/S0006-3495(95)80271-7).
- [40] A.G. Edwards, E. Grandi, J.E. Hake, S. Patel, P. Li, S. Miyamoto, et al., Non-equilibrium reactivation of Na<sup>+</sup> current drives early afterdepolarizations in mouse ventricle, *Circ. Arrhythm. Electrophysiol.* (2014), <http://dx.doi.org/10.1161/CIRCEP.113.001666>.
- [41] B.J. Gersh, B.J. Maron, R.O. Bonow, J.A. Dearani, M.A. Fifer, M.S. Link, et al., 2011 ACCF/AHA guideline for the diagnosis and treatment of hypertrophic cardiomyopathy: a report of the American College Of Cardiology Foundation/American Heart Association Task Force On Practice Guidelines, *J. Thorac. Cardiovasc. Surg.* 142 (2011) e153–e203, <http://dx.doi.org/10.1016/j.jtcvs.2011.10.020>.
- [42] H. Barajas-Martínez, D. Hu, R.J. Goodrow, F. Joyce, C. Antzelevitch, Electrophysiologic characteristics and pharmacologic response of human cardiomyocytes isolated from a patient with hypertrophic cardiomyopathy, *Pacing Clin. Electrophysiol.* 36 (2013) 1512–1515, <http://dx.doi.org/10.1111/pace.12227>.
- [43] L. Belardinelli, G. Liu, C. Smith-Maxwell, W.-Q. Wang, N. El-Bizri, R. Hirakawa, et al., A novel, potent, and selective inhibitor of cardiac late sodium current suppresses experimental arrhythmias, *J. Pharmacol. Exp. Ther.* 344 (2013) 23–32, <http://dx.doi.org/10.1124/jpet.112.198887>.
- [44] N. Jost, N. Nagy, C. Corici, Z. Kohajda, A. Horváth, K. Acsai, et al., ORM-10103, a novel specific inhibitor of the Na<sup>+</sup>/Ca<sup>2+</sup> exchanger, decreases early and delayed afterdepolarizations in the canine heart, *Br. J. Pharmacol.* 170 (2013) 768–778, <http://dx.doi.org/10.1111/bph.12228>.
- [45] Z.A. Nagy, L. Virág, A. Tóth, P. Biliczki, K. Acsai, T. Bányász, et al., Selective inhibition of sodium-calcium exchanger by SEA-0400 decreases early and delayed afterdepolarization in canine heart, *Br. J. Pharmacol.* 143 (2004) 827–831, <http://dx.doi.org/10.1038/sj.bjp.0706026>.
- [46] N. Nagy, A. Kormos, Z. Kohajda, A. Szebeni, J. Szepesi, P. Pollesello, et al., Selective Na(+)/Ca(2+) exchanger inhibition prevents Ca(2+) overload-induced triggered arrhythmias, *Br. J. Pharmacol.* 171 (2014) 5665–5681, <http://dx.doi.org/10.1111/bph.12867>.
- [47] R.S. Hansen, S.-P. Olesen, L.C.B. Rønne, M. Grønnet, In vivo effects of the IKr agonist NS3623 on cardiac electrophysiology of the guinea pig, *J. Cardiovasc. Pharmacol.* 52 (2008) 35–41, <http://dx.doi.org/10.1097/FJC.0b013e31817dd013>.
- [48] P. Pollesello, A. Mebazaa, ATP-dependent potassium channels as a key target for the treatment of myocardial and vascular dysfunction, *Curr. Opin. Crit. Care* 10 (2004) 436–441.
- [49] J. Heijman, P.G.a. Volders, R.L. Westra, Y. Rudy, Local control of β-adrenergic stimulation: effects on ventricular myocyte electrophysiology and Ca<sup>2+</sup>-transient, *J. Mol. Cell. Cardiol.* 50 (2011) 863–871, <http://dx.doi.org/10.1016/j.yjmcc.2011.02.007>.
- [50] J. Howie, L.B. Tulloch, M.J. Shattock, W. Fuller, Regulation of the cardiac Na(+) pump by palmitoylation of its catalytic and regulatory subunits, *Biochem. Soc. Trans.* 41 (2013) 95–100, <http://dx.doi.org/10.1042/BST20120269>.
- [51] M. Kuzumoto, A. Takeuchi, H. Nakai, C. Oka, A. Noma, S. Matsuoka, Simulation analysis of intracellular Na<sup>+</sup> and Cl<sup>-</sup> homeostasis during β1-adrenergic stimulation of cardiac myocyte, *Prog. Biophys. Mol. Biol.* 96 (2008) 171–186, <http://dx.doi.org/10.1016/j.pbiomolbio.2007.07.005>.
- [52] P.M. Kekenus-Huskey, T. Liao, A.K. Gillette, J.E. Hake, Y. Zhang, A.P. Michailova, et al., Molecular and subcellular-scale modeling of nucleotide diffusion in the cardiac myofibrillar lattice, *Biophys. J.* 105 (2013) 2130–2140, <http://dx.doi.org/10.1016/j.bpj.2013.09.020>.
- [53] E.B. Shim, H.M. Jun, C.H. Leem, S. Matsuoka, A. Noma, A new integrated method for analyzing heart mechanics using a cell-hemodynamics-autonomic nerve control coupled model of the cardiovascular system, *Prog. Biophys. Mol. Biol.* 96 (2008) 44–59, <http://dx.doi.org/10.1016/j.pbiomolbio.2007.07.015>.
- [54] J. Okada, T. Yoshinaga, J. Kurokawa, T. Washio, T. Furukawa, K. Sawada, et al., Screening system for drug-induced arrhythmogenic risk combining a patch clamp and heart simulator, *Sci. Adv.* 1 (2015) 1–7, <http://dx.doi.org/10.1126/sciadv.140014>.
- [55] V.J.A. Bourgonje, Vos M a., Ozdemir S, Doisne N, Acsai K, Varro A, et al. Combined Na<sup>+</sup>/Ca<sup>2+</sup> exchanger and I-type calcium channel block as a potential strategy to suppress arrhythmias and maintain ventricular function, *Circ. Arrhythm. Electrophysiol.* 6 (2013) 371–379, <http://dx.doi.org/10.1161/CIRCEP.113.000322>.