Review

ENVIRONMENTAL CONTAMINANT EXPOSURES DURING PREGNANCY INFLUENCE PRENATAL AND EARLY-LIFE: A COMPREHENSIVE REVIEW

María Dolores Gómez-Roig^{1,2}, Rosalia Pascal^{1,2*}, Marc Josep Cahuana¹, Óscar García Algar^{1,3}, Giorgia Sebastiani^{1,3}, Vicente Andreu-Fernández³, Leopoldo Martínez⁴, Gerardo Rodríguez

Martínez⁵, Iris Iglesia⁵, Olimpia Ortiz⁶, María Dolores Mesa^{6,7}, María Jesús Cabero Pérez⁸, Lorenzo

Guerra⁸, Elisa Llurba⁹, Carla Domínguez⁹, Julia Zaniani⁹, María Foraster¹⁰, Elvira Larqué¹¹,

Fernando Cabañas^{12,13}, Manuela López-Azorin¹², Carmen Rosa Pallás-Alonso¹⁴, Máximo Vento¹⁵

¹ BCNatal, Barcelona Center for Maternal Fetal and Neonatal Medicine, Hospital Sant Joan de Déu and Hospital Clínic. Universitat de Barcelona, Spain. (MD.G, RP)

² Institut de Recerca Sant Joan de Déu (IR-SJD), 08028 Barcelona, Spain.

³ Neonatology Unit, Hospital Clinic-Maternitat, ICGON, Universitat de Barcelona, Barcelona, Spain.

⁴ Servicio de Cirugía Pediátrica, Hospital la Paz. Instituto de Investigación la Paz (IdiPAZ), 28046 Madrid, Spain.

⁵ Growth, Exercise, Nutrition and Development (GENUD) Research Group, Universidad de Zaragoza, Spain. Instituto de Investigación Sanitaria Aragón (IIS Aragón), Spain

⁶ Department of Biochemistry and Molecular Biology II, Institute of Nutrition and Food Technology "José Mataix", Biomedical Research Center, University of Granada, Parque Tecnológico de la Salud, Granada, Spain.

⁷ Instituto de Investigación Biosanitaria, Complejo Hospitalario Universitario de Granada, Granada, Spain

⁸ Hospital Universitario Marqués de Valdecilla, Santander, Spain.

⁹ Obstetrics and Gynaecology Department, High Risk Unit, Sant Pau University Hospital. Barcelona, Spain. Women and Perinatal Health Research Group. Biomedical Research Institute Sant Pau (IIB-SantPau), Sant Pau University Hospital. Barcelona, Spain. School of Medicine, Universitat Autònoma de Barcelona, Barcelona, Spain.

¹⁰ ISGlobal,08003 Barcelona. Universitat Pompeu Fabra (UPF),08002 Barcelona. CIBER Epidemiología y Salud Pública (CIBERESO),28029 Madrid. Spain.

¹¹ Department of Physiology, Biomedical Institute of Research of Murcia Region (IMIB), Murcia University, Murcia, Spain

¹² Department of Neonatology, Quironsalud, Madrid University Hospital, Madrid, Spain

¹³ Biomedical Research Foundation, La Paz, Madrid University Hospital, Madrid, Spain

¹⁴ Donated Milk Bank, Health Research Institute i + 12, University Hospital 12 de Octubre, Universidad Complutense, 28040 Madrid, Spain.

¹⁵ Neonatal Research Group, Health Research Institute La Fe, University and Polytechnic Hospital La Fe, 46026 Valencia, Spain.

Short Title: Influence of environmental contaminants exposure in pregnancy

*Corresponding Author

Rosalia Pascal Capdevila

Barcelona Center for Maternal Fetal and Neonatal Medicine, Hospital Sant Joan de Déu and Hospital Clínic, Universitat de Barcelona.

PasseigSant Joan de Déu, 2, 08950 Esplugues de Llobregat, Barcelona, Spain

Tel: 34 932 53 21 00 - 71390

rpascal@sjdhospitalbarcelona.org

Keywords: Environmental exposure, endocrine disruptor compounds, heavy metals, noise, air pollution, pregnancy exposition.

1 Abstract

Preconceptional and prenatal exposure to environmental toxics may have an effect on an individual
future health, being pregnancy and early life critical and sensitive windows of susceptibility.

4 The aim of this review is to summarize the current evidence on the toxic effects of environment exposure5 during pregnancy, neonatal and childhood.

- 6 Alcohol use is related to Fetal Alcohol Spectrum Disorders (FASD) being Fetal Alcohol Syndrome (FAS)
- 7 its most extreme form. Smoking is associated with placental abnormalities, preterm birth, increased risk
- 8 of abortion, stillbirth or impaired growth and development, as well as to intellectual impairment, obesity
- 9 and cardiovascular diseases later in life. Negative birth outcomes have been linked to drugs of abuse.
- Pregnant and lactating women should be aware of the risks of chemicals acting as Endocrine Disruptor Compounds (EDCs) and heavy metals vehiculized by food intake and with deleterious effects on pregnancy and development. EDCs can work by altering body hormones and function, and its major
- 13 evidence of effects on prenatal exposure has been found for preeclampsia and intrauterine growth
- 14 restriction, preterm birth and thyroid function. Metals can accumulate in the placenta causing fetal growth
- 15 restriction.
- 16 Evidence of air pollution effects over pregnancy is constantly growing. It has been related to preterm
- 17 birth, with worrying evidence that synergies between its components enhance their adverse effects; fetal
- 18 growth restriction; increased uterine vascular resistance and impaired vascularization of the placenta;
- 19 increased gestational diabetes and reduced telomeres length.
- Initial studies suggest association between preeclampsia and environmental noise, particularly earlyonset preeclampsia.
- EDCs, heavy metals and air pollution are believed to have negative effects on the placenta, with consequential reduction in fetal growth, increased preterm birth, thyroid function disorders and neural tube defects.
- Physical activity during pregnancy is believed to have psychological benefits and has also been associated with shorter and less complicated labour, lower incidence of gestational diabetes, preterm birth, large for gestational age new-borns and hypertensive disorders.
- 28 The advantages of breast-feeding outweigh any risks from contaminants. However, it is important to
- 29 assess health outcomes of toxic exposures via breastfeeding that could have deleterious consequences
- 30 for new-born infants.
- In conclusion, there is rising evidence of the negative effects of environmental exposure during
 pregnancy and breastfeeding and should be considered a major public health issue in the early future.

33 1. Introduction

34

Environment, lifestyle and personal factors are considered to be health determinants with the capacityto influence disease, quality of life and mortality.

37 Environmental contaminants include those apparently under control, like food or abuse substances such

38 as alcohol and tobacco. However, we must not forget about air pollution, chemicals, water contamination

39 and radiation, often depending on governmental and industry policies.

40 We live surrounded by pollutants and objects with chemical components, but we don't always have

- 41 information about security exposure limits or the synergic effects of their combinations, thus representing
 42 a major public health concern(1,2).
- 43 Scientific evidence over the past years has raised concern that preconceptional and prenatal exposure

44 to toxic environmental agents may have a critical and lasting effect on future health and susceptibility to

45 disease(2-4). Given that development continues after birth, critical and sensitive windows occur

46 periconceptually (before, during and shortly after fertilization of the egg) and during pregnancy, but also

- 47 during infancy, childhood and puberty(4) [Fig.1].
- 48 We must not forget that most classes of environmental pollutants can cross into the fetal environment.

49 Some of them are xenobiotic and can accumulate in the placenta and foetus resulting in an even higher

- 50 fetal than maternal exposure and damage.
- 51 The World Health Organization (WHO) warns that an estimated 12.6 million people die every year as a

52 consequence of an unhealthy environment(5). Scientific societies such as the International Federation

53 of Gynaecology and Obstetrics (FIGO) work to raise awareness of this fact and prevent exposure to

54 toxins that negatively influence the health of mothers and their new-borns(6)

WINDOWS OF SUSCEPTIBILITY



55

- Figure 1: Human organogenesis and windows of susceptibility: prenatal and postnatal exposure. Modified from WHO State of the
 Science of EDCs 2012. Summary for Decision-Makers(7)
- 58

2. Environmental exposures in pregnancy

60

61 Pregnancy exposure to toxic environmental agents has an influence on perinatal outcomes but also on

62 infancy and childhood health [Table 1].

- 63
- 64

CONTAMINANT	CHEMICALS COMPOUNDS	ADVERSE HEALTH OUTCOMES	
Alcohol		Fetal Alcohol Spectrum Disorders (FASD), growth restriction, behavioural problems	
Cannabis, cocaine, heroin and methamphetamine		Fetal loss, preterm birth, small-for-gestational age, birth defects, behavioral problems	
Air pollutants	CO, NO, NO2 , SO2 , O3	Preterm Birth and small-for-gestational-age infants Autism spectrum disorder Increased risk of sudden infant death blood pressure Type 1 diabetes	
Polycyclic aromatic hydrocarbons (PAHS)	Coal or fossil fuel, forest fires, waste incineration	Preterm Birth and Small-for-gestational-age infants Asthma and allergic disease	
Particulate matters	Toxics with aerodynamic diameter (PM10-PM2.5)	Preterm Birth and lower birth weight. Asthma	
Persistent organic compounds	Organochlorine Compounds Polychlorobiphenyls (PCBs) Perfluoroalkylaed substances	Lower birth weight. Small-for-gestational-age infants Adverse neurodevelopmental outcome Attention deficit hyperactivity disorder Autism spectrum disorder Congenital anomalies Asthma and allergic disease Increased risk of sudden infant death blood pressure Leukemia	
Not persistent organic compounds	Phthalates, Phenols, and Parabens	Behavioral problems Attention deficit hyperactivity disorder Congenital anomalies	
Tobacco smoke	Nicotine, CO, aniline, methanol, hydrogen sulfide, arsenic, lead, cadmium.	Preterm Birth Small-for-gestational-age infants Adverse neurodevelopmental outcome Attention deficit hyperactivity disorder Asthma and allergic disease Increased risk of sudden infant death Congenital anomalies Leukemia	
Toxic Metals	Lead, cadmium, mercury, arsenic	Small-for-gestational-age infants Adverse neurodevelopmental outcome	

65

66 Table 1: Environmental contaminant exposure during pregnancy and adverse health outcomes

67 2.1. Toxic effect of prenatal exposure to substance use

68

Different toxic substances have been studied to assess their effects on pregnancy, neonatal and earlylife.

71 **2.1.1.** Alcohol

72

In our society, there is a high prevalence of alcohol consumption during pregnancy. The RCOG (Royal
College of Obstetricians and Gynaecologists) has reported that 29% of British pregnant women drinks
alcohol(8). Studies in Barcelona reveal, through the use of biological matrices, a 45% mild-moderate
social consumption(9,10).

Alcohol consumption during pregnancy may lead to adverse effects on fetal development described within the Fetal Alcohol Spectrum Disorders (FASD)(11). Currently, there is no definition of a safe amount and consumption period during pregnancy, implying it should be avoided. FASD affect up to 1% of world's paediatric population and its most extreme form is defined as Fetal Alcohol Syndrome (FAS)(11–13). Its clinical features can be broadly divided into: morphological malformations, especially craniofacial features (midfacial hypoplasia, wide spaced eyes and a smooth philtrum); growth restriction
and central nervous system impairment, resulting in motor, cognitive, learning and behavioural
disorders(13,14).

During fetal development, alcohol affects multiple metabolic pathways, partly through the alteration of DNA methyltransferase activities, which shapes the global epigenetic pattern of the developing foetus(15,16). Consequently, the expression of key genes is deregulated(17–20), affecting organogenesis especially of the fetal brain(21). Moreover, ethanol metabolism generates high amounts of reactive oxygen species (ROS), promoting oxidative stress and inhibiting endogenous antioxidant mechanisms(22). The increase of ROS alters both protein structures and mitochondrial respiration: which finally induces cellular apoptosis(23).

92 Prenatal Ethanol Exposed (PEE) detection is focused on the use of alcohol consumption questionnaires 93 and biomarkers in biological matrices. Determination of fatty acid ethyl esters (FAEEs) or ethyl-94 glucuronide (EtG) in meconium and maternal hair is the best procedure to identify PEE new-95 borns(10,24,25).

96 **2.1.2.** *Tobacco*

97

98 The global prevalence of smoking during pregnancy is high: according to a national survey conducted
99 in the United States, in 2012, 15,9% of pregnant women smoke cigarettes. Similar patterns of use have
100 been observed in Europe(26).

101 Smoking during gestation is associated with pregnancy complications such as severe preeclampsia, 102 placental abruption, placenta previa, preterm birth, increased risk of abortion, stillbirth or impaired growth 103 and development among many others(27-30), and with long term consequences such as intellectual 104 impairment later in life, leukaemia or asthma and allergic disease(29,31-36). Lean body mass of babies 105 from non-smoker mothers seems to be more affected than fat mass(37), and during the first years of 106 life, children from smoking mothers show complete catch-up growth(38). Based on the programming 107 effect(39), maternal smoking during pregnancy might determine children's weight status, blood pressure 108 or cardiovascular diseases in the medium-long term future. Although underlying mechanisms are not 109 clear, longitudinal studies sustain that children from smoker-mothers have a higher risk of developing 110 obesity over time(40.41). There is also a causal association between maternal exposure to cigarette 111 smoke and the risk of orofacial clefts, congenital heart disease, neural tube defects and gastrointestinal 112 malformations(42-44).

Thus, the deleterious effect of tobacco during pregnancy is well defined. However, due to its thousands of biologically active and toxic compounds, it is difficult to determine the causative agent of these adverse events. The anorexigenic effect of nicotine and its blood flow restriction to the placenta, the carbon monoxide exposure involving tissue hypoxia and the effect on DNA methylation are some of the most studied mechanisms(28,45).

118 2.1.3. Drugs of abuse

119

120 Prenatal substance abuse has increased noticeably among pregnant women, but the prevalence is still 121 underestimated. In 2017, the American National Survey on Drug Use and Health (NSDUH) assessed 122 that 194,000 pregnant women, aged from 15 to 44 years, had used illegal drugs in the past 123 month(46,47). Hair testing is the most sensitive and specific analysis to detect the concentration of 124 chronic drug exposure(48).

125 Negative birth outcomes have been linked to drugs of abuse, although the clear influence of each 126 substance is unknown because of the confounding effects of coexisting substances. Moreover, addicted 127 women often experience inadequate prenatal care, malnutrition, chronic illness and poverty, which 128 exacerbate the impairment of fetal development(26).

129 The principal consequences of opioid exposure in pregnancy are postnatal growth delay, microcephaly, 130 neurobehavioral disabilities and sudden infant death syndrome(49,50). Maternal opiate use increases 131 the risk of neonatal abstinence syndrome (NAS)(50), which comprises a wide range of symptoms, 132 including irritability, poor feeding, tremors, hypertonia, vomiting, loose stools, seizures, and respiratory

133 distress.

134 Cannabis, cocaine, heroin and methamphetamine are the most consumed substances and can cause 135 fetal loss, preterm birth, small-for-gestational age, birth defects and admission to the neonatal intensive 136 care unit(51). Cocaine and methamphetamine have been linked to premature rupture of membranes 137 and placental abruption, preeclampsia and gestational hypertension(52-55). In addition, all types of 138 drugs induce epigenetic changes in brain morphology, synaptic plasticity and behaviour(56). Prenatal 139 drugs use has been associated with microcephaly and adverse consequences for the growth of fetal 140 and adolescent brains(57), leading to lack of attention, reduced executive functioning skills and 141 disabilities in learning and memory, with consequent poorer academic attainment and more behavioural 142 problems(58-60).

143 2.2. The effect of maternal food intake

144

145 Pregnant and lactating women should be aware of the risks of heavy metals and other food toxic 146 compounds(61). These chemicals act as Endocrine Disruptor Compounds (EDCs) with deleterious 147 effects on pregnancy and development commented later on.

148 Highly toxic chemicals such as mercury, lead, arsenic, cadmium and chromium are elements that can 149 be vehiculized in foods and accumulated in the body(62) [Table 2]. They can be found in the environment 150 by means of voluntary application (plaguicides) and involuntary migration (from food containers and 151 plastic utensils), and then introduced in the food chain. In fact, food and specially those aliments from 152 animal origin with a high fat content are considered to be the main source of exposure to many pollutants 153 for the majority of the population(63).

154 Other compounds that can also be present in foods are organophosphate pesticides (OPPs), 155 polychlorinated biphenyl ethers (PBDEs)(64), acrylamide(65), perfluoroalkyl(66), as well as some 156 mycotoxins(67) and bacteria-derived toxics(68).

157

TOXIC COMPOUND	FOOD PRESENCE	TOXIC COMPOUND	FOOD PRESENCE
Mercury (59)	Fish/seafood (swordfish, sharks) Wild mushrooms Dietary supplements Non-alcoholic beverages	Cadmium (60,61)	Cereals/grains (rice, wheat) Vegetables (roots) Meat/poultry Seafood (bivalve molluscs)
Methylmercury	Tuna, swordfish, cod, whiting and pike.	Hexavalent Chromium (62)	Drinking water Special nutritional use products, Herbs, spices, condiments Sugar
Lead (63)	Bread and rolls Tea Tap water Potatoes Fermented milk Beer-like beverages	Aluminium (64)	Cereals Vegetables Beverages Infant formulae
Arsenic (65)	Fish/seafood Algae (hijiki) Cereals (rice grains)		

158

159 Table 2: Toxic chemicals and main dietary sources

160 **2.3.** The effect of maternal physical activity

161

162 Until a few decades ago, pregnant women were discouraged from exercise. However, this was mainly 163 due to social and cultural biases and unfounded concerns about safety for the foetus, rather than based 164 on scientific investigation. In recent years, there has been a growing interest in the effects of physical 165 exercise during pregnancy, so that the beneficial effects of regular physical exercise, both for the mother 166 and the foetus, are well-established, based mostly on systematic reviews and randomized 167 metanalysis(69-71). These physical benefits include maternal fitness and the prevention of excessive 168 weight gain, as well as psychological benefits. Regular exercise during pregnancy has also been 169 associated with shorter and less complicated labour, as well as the prevention of maternal-fetal diseases 170 such as gestational diabetes, preterm birth, being born large for gestational age and a lower incidence 171 of hypertensive disorders(72-74). Evenson et al identified, summarized and contrasted 11 clinical or 172 public health guidelines for physical activity during pregnancy from nine countries around the world 173 (Australia, Canada, Denmark, France, Japan, Norway, Spain, United Kingdom, United States). These 174 clinical guidelines mostly indicated the recommendation for physical activity during pregnancy, its 175 intensity and duration/time, as well as absolute and relative contraindications and indications for 176 discontinuing exercise during pregnancy(75).

Pregnancy may be one of the most important times to adopt a routine of regular exercise given thatlifestyle during pregnancy imprints the future health of the child.

179 2.4. Prenatal exposure to air pollution as a potential risk factor

Air pollution has a heterogeneous composition: particulate matter (PM), ozone pollution (O₃), carbon monoxide (CO), nitrogen oxides (NO₂, NOX) and sulfur dioxide (SO₂)(76,77). PM is a mixture of suspended particles with different chemical compositions usually classified by its size (PM10, PM2.5)(77). It has been widely studied due to its ability to trigger oxidative stress and inflammation in the lung's alveoli(78–80) and to cross the alveolar epithelium into the systemic circulation(81).

- 185 Evidence of pollution effects over pregnancy is constantly growing, and its relation with adverse perinatal 186 outcomes such as low birth weight (<2500 g) or pregnancy-induced hypertensive disorders is being well 187 established(78,82,83). Olsson et al. observed a positive association between NOx levels and an 188 increased risk for pregnancy-induced hypertensive disorders (OR 1.12, 95% CI 1.06 to 1.18 per 189 10µg/m3 increase in the NOx level)(84). A systematic review conducted by Pedersen et al.(85), 190 concludes that pregnancy-induced hypertensive disorders were associated with PM2.5 (OR=1.57; 95% 191 CI, 1.26–1.96 per a 5µg/m3 increment), NO₂ (OR=1.20; 95% CI, 1.00–1.44 per a 10-µg/ m3 increase) 192 and PM10, (OR=1.13; 95% CI, 1.02-1.26, per a 10-µg/m3 increase). As for fetal growth, PM2,5 193 exposure was negatively associated with reduced head circumference at birth and birth weight(86) while 194 NO₂ was significantly linked to a shorter length at birth(86-88). NO_x has been related to a decrease of 195 abdominal circumference and femoral length and a reduce of birth weight(89).
- Exposure to PM and O₃ has been associated to a higher risk of preterm birth (27,87,88,90). Moreover,
 synergies between PM2.5 and O₃ showed more risk (RR 3,63) than their independent effects (RR 0,99
 and 1,34, respectively)(91).

PM10, NO and O₃ have been associated to macrosomia(92) and PM2,5 has been related in animal
 studies to profound metabolic effects (like glucose intolerance, decrease insulin sensitivity and altered
 hepatic glucose and lipid metabolism) through oxidative stress(93).

In a multicentric European birth cohort of 1396 subjects, the Helix Project, exposure levels of NO₂ and
 PM 2,5 were inversely associated with telomer length(94)

204 Regarding the fetal nervous system, in utero exposure to PM2.5 during the first trimester was found to 205 decrease placental transcription of brain-derived neurotrophic factor (BDNF). This factor plays an 206 important role in fetal neurodevelopment(95). Furthermore, neuropathological changes (microglial 207 activation, ventriculomegaly, increased size of the Corpus Callosum, reduction in hippocampal size) 208 were found to be induced by prenatal exposure to ultrafine particles in mice(96). In addition, Guxens et 209 al. showed cerebral cortex alterations and impairment of inhibitory control function in children exposed 210 to fine particles during gestation. This impaired function is related to attention-deficit or hyperactivity 211 disorder(97). According to Danish investigators, gestational exposure to air pollution may also increase 212 the risk of autism spectrum disorder(98).

- 213 2.5. Prenatal exposure to endocrine disruptors and toxic metals
- 214

215 EDCs are exogenous chemicals (phenols, phthalates, parabens, flame retardants and heavy metals)

that can alter the hormonal and homeostatic systems of the organism(99). Exposure to EDCs may occur in pregnancy by way of personal hygiene products, cosmetics, cleaning products, electronic devices

and consumption of animal, plant or processed foods(2).

EDCs can work by altering normal hormonal production and levels and mimicking their function. Its main effects on prenatal exposure have been studied with major evidence on preeclampsia and intrauterine growth restriction, preterm birth and thyroid dysfunction(100,101).

When talking about hypertensive disorders of pregnancy, evidence is strongest for links between persistent chemicals (lead, cadmium, organochlorine pesticides and polycyclic biphenyls) and preeclampsia, although low-exposure levels associations are not always detectable. However, results have been inconclusive for bisphenols, phthalates and organophosphates(101).

226 Metals and metalloids accumulate in the placenta, causing a decrease in uterine blood flow and having 227 a negative impact on fetal growth(100,102). It has also been described that plasticizers, like 228 diethylhexylphthalate (DEHP) and its active metabolites, and bisphenols A (BPA) induce preeclampsia 229 and growth restriction(103-105). The exposure to pesticides such as dichlorodiphenyltrichloroethane 230 (DDT) and its metabolites have also been suggested to have a detrimental effect on fetal growth(106). 231 Organochlorine pesticides may lead to preterm birth through disturbance of normal estrogen-232 progesterone ratio(107), might increase the risk of autism spectrum disorder(108) and with also 233 evidence of thyroid disrupting properties(109).

Flame-retardants such as PBDE and tetrabromopisphenol A (TBBPA) have been linked to growth restriction and preterm birth, as well as impairment of the thyroid hormone function(110,111).

A growing number of studies suggest a link between congenital anomalies and maternal exposure to organic solvents, pesticides and dioxins (cleft lip and palate, neural tube defects, and congenital heart disease)(112,113). Toluene embryopathy has been described after maternal inhalation of paint or glue(114). Phthalates have antiandrogenic-like properties and have a great role in hypospadias and cryptorchidie(115). Pesticides are considered to be a risk factor for childhood leukaemia(116). Finally, maternal exposure to BPA increases rates of depression, behavioural problems and alterations in white

242 matter in preschool aged children(117,118).

243 2.6. Prenatal noise stress

244

Noise pollution is a major environmental health concern. It is estimated that 113 million people in Europe are exposed to excessive environmental noise levels according to the European Environmental Noise Directive (2002/49/EC), majorly from road traffic noise. The implication of environmental noise on several health disorders is already recognized(119,120). 249 Exposure to noise has been associated with cardiovascular effects, like hypertension, stroke and 250 myocardial infarction in many studies(120), with high-quality evidence for the association between road 251 traffic noise and incidence of ischemic heart disease(121). The suggested biological pathways indicate 252 that repeated exposure to noise causes stress responses, as well as sleep disturbance, leading to 253 endocrine and sympathetic responses, which increase blood pressure, heart rate, and cardiac output 254 through the release of catecholamines(119) and corticosteroids(122), and to oxidative-stress and 255 immunological responses(123). These reactions persist even while asleep and can lead to chronic 256 physiological deregulations(120). However, few studies have investigated the effect of exposure to noise 257 in pregnant women, being preeclampsia of special interest. A recent study of 269.263 deliveries in 258 Quebec, Canada(124), showed that women exposed to > 65 dB(A) had 1.29 times the odds of severe 259 (95%CI: 1.09-1.54) and 1.71 times the odds of early onset (95%CI: 1.20-2.43) preeclampsia compared 260 to those exposed to <50 dB(A)(124).

A Danish prospective cohort study with 72,745 women showed that a 10-dB increase in road traffic noise was associated with a 10% increase in the risk of preeclampsia(125). These associations were stronger for the mild subtypes of preeclampsia and early preeclampsia and not evident for severe preeclampsia. They concluded that the effects of air pollution and noise were generally difficult to separate.

In conclusion, these initial studies suggest that exposure to environmental noise is associated withpreeclampsia, particularly early onset preeclampsia. However, more studies are needed.

268 **3. Environmental effects on placenta**

269

The placenta is a highly sensitive organ to environmental contaminants with estrogenic activity as it expresses the oestrogen receptors ER α and ER β (126). Although there are many reports in the literature of the in vitro action of different EDCs in human placenta, some controversies remain regarding the timing, dose and duration of exposure(127). It is important to emphasize that the effects of EDCs in human trophoblasts are dose-dependent with low doses being more effective than high doses(115): This is concerning because the efficacious low doses correspond to the levels detected in the human population.

Fergusson et al.(128) found a positive association between BPA levels and an increase of plasma soluble vascular endothelial growth factor receptor 1 (sFIt-1) as well as an increase in the ratio of sFIt-1 to placental growth factor (PIGF), suggesting an altered placentation and trophoblast function related to preeclampsia and hypertensive disorders(129).

In vitro studies showed that para-Nonylphenol (p-NP) substances, used as plasticizer and surfactant in the manufacturing industry, could increase β -hCG secretion, cell apoptosis and reduce trophoblasts migration and invasion. Exposure to BPA and p-NP down-regulated expression of some placental carriers like ABCG2, a key transporter for xenobiotics(130).

- In addition, PBDEs mixtures enhanced the placental proinflammatory response to infection. This may increase the risk of infection-mediated preterm birth by lowering the threshold for bacteria to stimulate
- a proinflammatory response(131). Rats exposed to PBDEs during gestation showed effects on placenta
- and foetus that varied by foetal sex. mRNA expression in the placenta also significantly varied by foetal
- sex and dose. Thus, PBDEs are impacting thyroid hormones regulation in a sex-specific manner during
- this critical window of development(132).
- Higher concentrations of polycyclic aromatic hydrocarbons (PAHs) such as benzo[a]pyene (BP), benzo[b]fluorene (BbF) and dibenz[a,h]anthracene (DBA) were found in placenta from preterm deliveries compared with term deliveries(133).
- Related to heavy metals, in the New Hampshire Birth Cohort Study (N = 1159), with every ng/g increase in the Cadmiun concentration of placenta there was lower placental weight (-7.81 g; 95% CI: -15.42, -2.48). For placentae with below median Zinc and Selenium concentrations, decrements in placental weight were - 8.81 g (95% CI: -16.85, -0.76) and - 13.20 g (95% CI: -20.70, -5.70), respectively. However, no appreciable differences were observed with other elements (arsenic, mercury and lead)(134).
- 300 As far as air pollution is concerned, circulating proinflammatory cytokines induced by PM may disrupt 301 trophoblastic invasion during placenta formation(135,136). Likewise, PM could enter in uteroplacental 302 circulation resulting in placenta pathological changes(137). Placenta chorioamnionitis and thrombosis 303 of placental capillaries have been demonstrated by LiuY in a rat model after PM2.5 exposure. These 304 changes in placenta tissue lead to reduced maternofoetal exchange surface and to placental 305 dysfunction(78). Neven et al. analysed placental DNA and found an association between elevated 306 placental mutation rate and prenatal exposure to PM2.5 and black carbon. They postulated that this 307 placental mutations could represent some of the earliest effects to air pollutants exposure in the process 308 of carcinogenesis(137).

309 4. Breastfeeding: Environmental toxins in human milk and early-life consequences310

- Breast feeding is the gold standard of new-born and child nutrition during at least the first 6 months of life(138). Bottle-feeding is associated to the transfer of toxic substances from recipients to milk. However, milk transfer of toxic substances to which the mother has previously been exposed, may also occur during breastfeeding. Several comprehensive reviews conclude that breastfeeding is generally contraindicated in mothers who use illegal drugs(139), although pharmacokinetic data are sparse in lactating woman(140).
- Smoking and alcohol consumption should be avoided during the breastfeeding period. Alcohol interferes with the milk ejection reflex, which may reduce milk production. Human milk alcohol levels generally parallel maternal blood alcohol levels. Studies evaluating infant effects of maternal alcohol consumption have been mixed, with some mild effects seen in infant sleep patterns, amount of milk consumed during

- 321 breastfeeding sessions and early psychomotor development. Some authors suggest limiting alcohol
- 322 intake to the equivalent of 8 ounces of wine or two beer is recommended(139). However, others state
- 323 that alcohol consumption during both pregnancy and breastfeeding should be totally avoided since there
- is not a proven safe consumption dose(10,47). Nicotine and other compounds are known to be milk transferred to the infant causing increases in the incidence of respiratory allergy in infants and in Sudden
- 323 tansiened to the infant causing increases in the incidence of respiratory allergy in finants and in Sudden
- 326 Infant Death Syndrome (SIDS) risks(141).

327 Infants are exposed trough breastfeeding to a mixture of environmental chemicals. Lactating mothers

- 328 are among the high-risk population to mercury exposure because they may suffer the consequences of
- 329 mercury themselves, but also they may transfer significant quantities of mercury to their babies(142).
- 330 BPA has also been widely studied: The temporary tolerability daily intake (t-TDI) of 4 µg/kg. bw⁻¹ day⁻¹ 331 for oral exposure to BPA has been established (143). In lactating mothers, BPA is rapidly introduced into 332 the breast causing an elevation of BPA content in the milk within hours. Only the unconjugated BPA 333 present in the milk is active, consequently its determination is more suitable for the assessment of BPA 334 risk in breastfed infants(144). Interestingly, while BPA content in mature milk reflects recent ingestion, 335 its content in colostrum reflects ingestion in the second half of pregnancy(145). The place of residence 336 of the mother and the use of personal care products showed significant association with BPA 337 concentration(146).
- 338 Human milk contains conjugated and un-conjugated parabens and provides the exposure of the mother, 339 the foetus, and the neonate in a period of high vulnerability to the endocrine disruptors(147). In a 340 Spanish study, the detection frequency ranges of parabens in breast milk were 41-60% and 61-89% for 341 unconjugated and total (unconjugated + conjugated) parabens, respectively. The frequency of use of 342 some cosmetic products and human milk protein levels were the main predictors of parabens in milk. 343 The new-borns estimated daily intake of parabens through human milk (median= 0.014 μ g/kg bw-day) 344 was several orders of magnitude lower than the 1-10 mg/kg bw-day acceptable daily intake as 345 established by European Food Safety Authority (EFSA)(143,148). In a recent study, Sanchis et al found 346 high urinary concentrations of Methylparaben (MP), Ethylparaben (EP) and BPA in lactating mothers 347 although estimated exposures was lower than the reference values for risk assessment. The use of 348 personal care products was associated with higher urinary levels of MP and Propylparaben (PP). MP 349 was also associated with the consumption of packaged and bakery products(149).
- All these chemicals may influence infant gut microbial function(150), increase risk of hyperkinetic disorder(151), toxicity to the liver and kidney, cancer, reproductive and respiratory disorders (143,152) or changes in thyroid and growth hormones that may have effects on neurodevelopment(143,153).
- 353 When mother's milk is not available or is insufficient, pasteurized donor milk is recommended. The use
- of illegal drugs, alcohol and tobacco is an exclusion criterion for accepting a nursing mother as a milk
- donor. Escuder et al. found that donors do not use illegal drugs during either the donation period or the

- months leading up to it, they are occasionally exposed to tobacco smoke and almost all of them consumecaffeine(154).
- 358 Although most scientific evidence indicate that the advantages of breast-feeding outweigh any risks from
- 359 contaminants exposure to these toxics can have deleterious consequences especially for a vulnerable
- 360 population such as lactating women and breastfed new-born infants. Special caution with preterm infants
- 361 should be posed.

362 **5. Conclusions**

363

Environment exposure is considered to be a health determinant with the capacity to influence disease, quality of life and mortality. Although this exposure can be deleterious for any person, pregnancy and early life exposure have been demonstrated to be critical windows of susceptibility, with a lasting effect on future health and susceptibility to disease(2–4).

368 The use of alcohol, tobacco and drugs of abuse has been linked to a serious of deleterious effects in

the new-born and later in life, including FASD and other negative pregnancy and birth outcomes. EDCsand heavy metals vehiculized by food intake or present in the environment are related to preeclampsia,

- 371 foetal growth restriction, preterm birth and thyroid misfunction. Air pollution has been linked to preterm
- birth, foetal growth restriction, effects on pregnancy vascularization, increased gestational diabetes and
- 373 reduced telomeres length. Association between preeclampsia and environmental noise is rising. On the
- 374 contrary, physical activity during pregnancy is believed to have remarkable benefits and therefore should
- be recommended during pregnancy. Breastfeeding should be recommended; however, mothers should
- be aware of toxic exposures via breastfeeding that could have consequences for new-born infants.

Therefore, doctors should have knowledge of harmful exposures to be able to counsel patients on the risk and advise them with precautions to minimize exposure, especially during pregnancy and breastfeeding. Governmental protection should be strengthened, by limiting environmental exposure to substances with evidence of a deleterious effect. However, only with a global public health policy in the early future could all this evidence be translated into action.

382

383 **Disclosure statement:** The authors have no conflicts of interest to declare.

Funding Sources: This research was funded by the PN I+D+I 2008–2011 (Spain), ISCIII- Sub Directorate General for Research Assessment and Promotion and the European Regional Development
 Fund (ERDF), RETICS Maternal and Child Health and Development Network, SAMID Network, Ref.
 RD16/0022/0015

Author Contributions: All authors have contributed to the writing of the work. MD.G. and RP had primary responsibility for the final content. All authors have read and agreed to the published version of the manuscript.

6. References

- 1. Trasande L, Zoeller RT, Hass U, Kortenkamp A, Grandjean P, Myers JP, et al. Burden of disease and costs of exposure to endocrine disrupting chemicals in the European Union: an updated analysis. Andrology [Internet]. 2016 Jul 1 [cited 2020 Mar 15];4(4):565–72. Available from: http://www.ncbi.nlm.nih.gov/pubmed/27003928
- Kelley AS, Banker M, Goodrich JM, Dolinoy DC, Burant C, Domino SE, et al. Early pregnancy exposure to endocrine disrupting chemical mixtures are associated with inflammatory changes in maternal and neonatal circulation. Sci Rep [Internet]. 2019 Apr 1 [cited 2020 Mar 15];9(1):5422. Available from: http://www.ncbi.nlm.nih.gov/pubmed/30931951
- 3. Bommarito PA, Martin E, Fry RC. Effects of prenatal exposure to endocrine disruptors and toxic metals on the fetal epigenome. Epigenomics [Internet]. 2017 Mar 1 [cited 2020 Mar 15];9(3):333–50. Available from: http://www.ncbi.nlm.nih.gov/pubmed/28234024
- Woodruff TJ, Carlson A, Schwartz JM, Giudice LC. Proceedings of the Summit on Environmental Challenges to Reproductive Health and Fertility: executive summary. Fertil Steril [Internet]. 2008 Feb [cited 2020 Mar 15];89(2):281–300. Available from: http://www.ncbi.nlm.nih.gov/pubmed/18275883
- 5. An estimated 12.6 million deaths each year are attributable to unhealthy environments [Internet]. [cited 2020 Mar 15]. Available from: https://www.who.int/news-room/detail/15-03-2016-an-estimated-12-6-million-deaths-each-year-are-attributable-to-unhealthy-environments
- 6. Reproductive and Developmental Environmental Health | FIGO [Internet]. [cited 2020 Mar 15]. Available from: https://www.figo.org/working-group-reproductive-anddevelopmental-environmental-health
- 7. WHO | State of the science of endocrine disrupting chemicals 2012 [Internet]. [cited 2020 Apr 13]. Available from: https://www.who.int/ceh/publications/endocrine/en/
- 8. BJOG release: Pre-pregnancy alcohol consumption affects alcohol use among pregnant women, suggests new study [Internet]. [cited 2020 Mar 15]. Available from: https://www.rcog.org.uk/en/news/bjog-release-pre-pregnancy-alcohol-consumption-affects-alcohol-use-among-pregnant-women-suggests-new-study/
- Garcia-Algar O, Kulaga V, Gareri J, Koren G, Vall O, Zuccaro P, et al. Alarming prevalence of fetal alcohol exposure in a Mediterranean city. Ther Drug Monit [Internet]. 2008 Apr [cited 2020 Mar 15];30(2):249–54. Available from: http://www.ncbi.nlm.nih.gov/pubmed/18367990
- 10. Gomez-Roig MD, Marchei E, Sabra S, Busardò FP, Mastrobattista L, Pichini S, et al. Maternal hair testing to disclose self-misreporting in drinking and smoking behavior during pregnancy. Alcohol [Internet]. 2018 Mar 1 [cited 2020 Mar 15];67:1–6. Available from: http://www.ncbi.nlm.nih.gov/pubmed/29289821
- Hoyme HE, Kalberg WO, Elliott AJ, Blankenship J, Buckley D, Marais AS, et al. Updated clinical guidelines for diagnosing fetal alcohol spectrum disorders. Pediatrics [Internet]. 2016 Aug 1 [cited 2020 Mar 15];138(2). Available from: http://www.ncbi.nlm.nih.gov/pubmed/27464676
- 12. Lange S, Probst C, Gmel G, Rehm J, Burd L, Popova S. Global prevalence of fetal alcohol spectrum disorder among children and youth: A systematic review and meta-analysis. JAMA Pediatr. 2017 Oct 1;171(10):948–56.
- 13. Sokol RJ, Delaney-Black V, Nordstrom B. Fetal Alcohol Spectrum Disorder [Internet].

Vol. 290, Journal of the American Medical Association. 2003 [cited 2020 Mar 15]. p. 2996–9. Available from: http://www.ncbi.nlm.nih.gov/pubmed/14665662

- 14. Sampson PD, Streissguth AP, Bookstein FL, Little RE, Clarren SK, Dehaene P, et al. Incidence of fetal alcohol syndrome and prevalence of alcohol-related neurodevelopmental disorder. Teratology [Internet]. 1997 Nov [cited 2020 Mar 15];56(5):317–26. Available from: http://www.ncbi.nlm.nih.gov/pubmed/9451756
- Pruett D, Waterman EH, Caughey AB. Fetal alcohol exposure: Consequences, diagnosis, and treatment [Internet]. Vol. 68, Obstetrical and Gynecological Survey. 2013 [cited 2020 Mar 15]. p. 62–9. Available from: http://www.ncbi.nlm.nih.gov/pubmed/23322082
- Legault LM, Bertrand-Lehouillier V, McGraw S. Pre-implantation alcohol exposure and developmental programming of FASD: An epigenetic perspective [Internet]. Vol. 96, Biochemistry and Cell Biology. Canadian Science Publishing; 2018 [cited 2020 Mar 15].
 p. 117–30. Available from: http://www.ncbi.nlm.nih.gov/pubmed/29088550
- 17. Miller MW. Limited ethanol exposure selectively alters the proliferation of precursor cells in the cerebral cortex. Alcohol Clin Exp Res [Internet]. 1996 Feb [cited 2020 Mar 15];20(1):139–43. Available from: http://www.ncbi.nlm.nih.gov/pubmed/8651443
- Medina AE, Krahe TE. Neocortical plasticity deficits in fetal alcohol spectrum disorders: Lessons from barrel and visual cortex [Internet]. Vol. 86, Journal of Neuroscience Research. 2008 [cited 2020 Mar 15]. p. 256–63. Available from: http://www.ncbi.nlm.nih.gov/pubmed/17671993
- Nithianantharajah J, Hannan AJ. Enriched environments, experience-dependent plasticity and disorders of the nervous system. Vol. 7, Nature Reviews Neuroscience. 2006. p. 697–709.
- 20. Fernandez-Lizarbe S, Pascual M, Guerri C. Critical Role of TLR4 Response in the Activation of Microglia Induced by Ethanol. J Immunol [Internet]. 2009 Oct 1 [cited 2020 Mar 15];183(7):4733–44. Available from: http://www.ncbi.nlm.nih.gov/pubmed/19752239
- 21. Guerri C, Bazinet A, Riley EP. Foetal Alcohol Spectrum Disorders and alterations in brain and behaviour. Alcohol Alcohol [Internet]. [cited 2020 Mar 15];44(2):108–14. Available from: http://www.ncbi.nlm.nih.gov/pubmed/19147799
- Wu D, Cederbaum AI. Alcohol, oxidative stress, and free radical damage. Alcohol Res Health [Internet]. 2003 [cited 2020 Mar 15];27(4):277–84. Available from: http://www.ncbi.nlm.nih.gov/pubmed/15540798
- Liesi P. Ethanol-exposed central neurons fail to migrate and undergo apoptosis. J Neurosci Res [Internet]. 1997 Jun 1 [cited 2020 Mar 15];48(5):439–48. Available from: http://www.ncbi.nlm.nih.gov/pubmed/9185667
- Pichini S, García-Algar O, Alvarez A-T, Mercadal M, Mortali C, Gottardi M, et al. Pediatric exposure to drugs of abuse by hair testing: monitoring 15 years of evolution in Spain. Int J Environ Res Public Health [Internet]. 2014 Aug 14 [cited 2020 Mar 15];11(8):8267–75. Available from: http://www.ncbi.nlm.nih.gov/pubmed/25153461
- Joya X, Marchei E, Salat-Batlle J, García-Algar O, Calvaresi V, Pacifici R, et al. Fetal exposure to ethanol: Relationship between ethyl glucuronide in maternal hair during pregnancy and ethyl glucuronide in neonatal meconium. Clin Chem Lab Med [Internet].
 2016 Mar 1 [cited 2020 Mar 15];54(3):427–35. Available from: http://www.ncbi.nlm.nih.gov/pubmed/26351940
- 26. Forray A. Substance use during pregnancy [version 1; referees: 2 approved]. Vol. 5, F1000Research. Faculty of 1000 Ltd; 2016.

- 27. Porpora MG, Piacenti I, Scaramuzzino S, Masciullo L, Rech F, Panici PB. Environmental contaminants exposure and preterm birth: A systematic review. Vol. 7, Toxics. MDPI AG; 2019.
- Wiklund P, Karhunen V, Richmond RC, Parmar P, Rodriguez A, De Silva M, et al. DNA methylation links prenatal smoking exposure to later life health outcomes in offspring. Clin Epigenetics [Internet]. 2019 Jul 1 [cited 2020 Mar 15];11(1):97. Available from: http://www.ncbi.nlm.nih.gov/pubmed/31262328
- Lange S, Probst C, Rehm J, Popova S. National, regional, and global prevalence of smoking during pregnancy in the general population: a systematic review and metaanalysis. Lancet Glob Heal [Internet]. 2018 Jul 1 [cited 2020 Mar 15];6(7):e769–76. Available from: http://www.ncbi.nlm.nih.gov/pubmed/29859815
- 30. Quelhas D, Kompala C, Wittenbrink B, Han Z, Parker M, Shapiro M, et al. The association between active tobacco use during pregnancy and growth outcomes of children under five years of age: A systematic review and meta-analysis. Vol. 18, BMC Public Health. BioMed Central Ltd.; 2018.
- Wu CC, Hsu TY, Chang JC, Ou CY, Kuo HC, Liu CA, et al. Paternal tobacco smoke correlated to offspring asthma and prenatal epigenetic programming. Front Genet. 2019;10(MAY).
- Mishra A, Mishra K, Marks AJ, Mishra NC. Cronicon EC PULMONOLOGY AND RESPIRATORY MEDICINE Editorial Prenatal Cigarette Smoke Exposure and Development of Asthma in Progeny.
- Burke H, Leonardi-Bee J, Hashim A, Pine-Abata H, Chen Y, Cook DG, et al. Prenatal and passive smoke exposure and incidence of asthma and wheeze: Systematic review and meta-analysis. Vol. 129, Pediatrics. American Academy of Pediatrics; 2012. p. 735– 44.
- Rosas-Salazar C, Hartert T V. Prenatal exposures and the development of childhood wheezing illnesses. Vol. 17, Current Opinion in Allergy and Clinical Immunology. Lippincott Williams and Wilkins; 2017. p. 110–5.
- 35. Frederiksen LE, Erdmann F, Wesseling C, Winther JF, Mora AM. Parental tobacco smoking and risk of childhood leukemia in Costa Rica: A population-based case-control study. Environ Res. 2020 Jan 1;180.
- 36. De Smith AJ, Kaur M, Gonseth S, Endicott A, Selvin S, Zhang L, et al. Correlates of prenatal and early-life tobacco smoke exposure and frequency of common gene deletions in childhood acute lymphoblastic leukemia. Cancer Res. 2017;77(7):1674–83.
- 37. Samper MP, Jiménez-Muro A, Nerín I, Marqueta A, Ventura P, Rodríguez G. Maternal active smoking and newborn body composition. Early Hum Dev [Internet]. 2012 Mar [cited 2020 Mar 15];88(3):141–5. Available from: http://www.ncbi.nlm.nih.gov/pubmed/21821370
- 38. Fenercioglu AK, Tamer I, Karatekin G, Nuhoglu A. Impaired postnatal growth of infants prenatally exposed to cigarette smoking. Tohoku J Exp Med [Internet]. 2009 Jul [cited 2020 Mar 15];218(3):221–8. Available from: http://www.ncbi.nlm.nih.gov/pubmed/19561393
- Somm E, Schwitzgebel VM, Vauthay DM, Aubert ML, Hüppi PS. Prenatal nicotine exposure and the programming of metabolic and cardiovascular disorders [Internet]. Vol. 304, Molecular and Cellular Endocrinology. 2009 [cited 2020 Mar 15]. p. 69–77. Available from: http://www.ncbi.nlm.nih.gov/pubmed/19433250
- 40. Magriplis E, Farajian P, Panagiotakos DB, Risvas G, Zampelas A. Maternal smoking and risk of obesity in school children: Investigating early life theory from the GRECO study.

Prev Med reports [Internet]. 2017 Dec 1 [cited 2020 Mar 15];8:177–82. Available from: http://www.ncbi.nlm.nih.gov/pubmed/29071203

- 41. Iguacel I, Escartín L, Fernández-Alvira JM, Iglesia I, Labayen I, Moreno LA, et al. Early life risk factors and their cumulative effects as predictors of overweight in Spanish children. Int J Public Health [Internet]. 2018 May 1 [cited 2020 Mar 15];63(4):501–12. Available from: http://www.ncbi.nlm.nih.gov/pubmed/29549397
- 42. Gunnerbeck A, Bonamy AKE, Wikström AK, Granath F, Wickström R, Cnattingius S. Maternal snuff use and smoking and the risk of oral cleft malformations A population-based cohort study. PLoS One [Internet]. 2014 Jan 15 [cited 2020 Mar 31];9(1):e84715. Available from: http://www.ncbi.nlm.nih.gov/pubmed/24454740
- 43. Leite M, Albieri V, Kjaer SK, Jensen A. Maternal smoking in pregnancy and risk for congenital malformations: Results of a Danish register-based cohort study. Acta Obstet Gynecol Scand [Internet]. 2014 Aug [cited 2020 Mar 31];93(8):825–34. Available from: http://www.ncbi.nlm.nih.gov/pubmed/24861914
- Sabbagh HJ, Hassan MHA, Innes NPT, Elkodary HM, Little J, Mossey PA. Passive smoking in the etiology of non-syndromic orofacial clefts: A systematic review and metaanalysis [Internet]. Vol. 10, PLoS ONE. Public Library of Science; 2015 [cited 2020 Mar 31]. p. e0116963. Available from: http://www.ncbi.nlm.nih.gov/pubmed/25760440
- 45. Wickstrom R. Effects of Nicotine During Pregnancy: Human and Experimental Evidence. Curr Neuropharmacol. 2007 Aug 30;5(3):213–22.
- 46. Center for Behavioral Health Statistics S. RESULTS FROM THE 2017 NATIONAL SURVEY ON DRUG USE AND HEALTH: DETAILED TABLES.
- Cortes L, Almeida L, Sabra S, Muniesa M, Busardo FP, Garcia-Algar O, et al. Maternal Hair and Neonatal Meconium to Assess Gestational Consumption and Prenatal Exposure to Drugs of Abuse and Psychoactive Drugs. Curr Pharm Biotechnol. 2018 Apr 6;19(2):136–43.
- 48. Falcon M, Pichini S, Joya J, Pujadas M, Sanchez A, Vall O, et al. Maternal hair testing for the assessment of fetal exposure to drug of abuse during early pregnancy: Comparison with testing in placental and fetal remains. Forensic Sci Int [Internet]. 2012 May 10 [cited 2020 Mar 15];218(1–3):92–6. Available from: http://www.ncbi.nlm.nih.gov/pubmed/22036306
- Minozzi S, Amato L, Bellisario C, Ferri M, Davoli M. Maintenance agonist treatments for opiate-dependent pregnant women. Vol. 2013, Cochrane Database of Systematic Reviews. John Wiley and Sons Ltd; 2013.
- 50. Hudak ML, Tan RC, Frattarelli DAC, Galinkin JL, Green TP, Neville KA, et al. Neonatal drug withdrawal. Pediatrics. 2012 Feb;129(2).
- 51. Jaques SC, Kingsbury A, Henshcke P, Chomchai C, Clews S, Falconer J, et al. Cannabis, the pregnant woman and her child: weeding out the myths. J Perinatol [Internet]. 2014 Jun [cited 2020 Mar 15];34(6):417–24. Available from: http://www.ncbi.nlm.nih.gov/pubmed/24457255
- 52. Addis A, Moretti ME, Ahmed Syed F, Einarson TR, Koren G. Fetal effects of cocaine: an updated meta-analysis. Reprod Toxicol [Internet]. [cited 2020 Mar 15];15(4):341–69. Available from: http://www.ncbi.nlm.nih.gov/pubmed/11489591
- 53. Gouin K, Murphy K, Shah PS. Effects of cocaine use during pregnancy on low birthweight and preterm birth: Systematic review and metaanalyses. Am J Obstet Gynecol [Internet]. 2011 Apr [cited 2020 Mar 15];204(4):340.e1-340.e12. Available from: http://www.ncbi.nlm.nih.gov/pubmed/21257143

- 54. Wright TE, Schuetter R, Tellei J, Sauvage L. Methamphetamines and pregnancy outcomes. J Addict Med. 2015 Dec 1;9(2):111–7.
- 55. Brecht ML, Herbeck DM. Pregnancy and fetal loss reported by methamphetamine-using women. Subst Abus Res Treat [Internet]. 2014 Apr 28 [cited 2020 Mar 15];8:25–33. Available from: http://www.ncbi.nlm.nih.gov/pubmed/24855369
- 56. Mason B, Donaldson ST, Hunter RG. The developmental neuroepigenetics of substance abuse. J Drug Alcohol Res. 2018;7(March):1–27.
- 57. van Dyk J, Ramanjam V, Church P, Koren G, Donald K. Maternal methamphetamine use in pregnancy and long-term neurodevelopmental and behavioral deficits in children. J Popul Ther Clin Pharmacol. 2014 May 20;21(2).
- Hayatbakhsh MR, Flenady VJ, Gibbons KS, Kingsbury AM, Hurrion E, Mamun AA, et al. Birth outcomes associated with cannabis use before and during pregnancy. Pediatr Res. 2012 Feb;71(2):215–9.
- Warner TD, Roussos-Ross D, Behnke M. It's not your mother's marijuana: Effects on maternal-fetal health and the developing child. Vol. 41, Clinics in Perinatology. W.B. Saunders; 2014. p. 877–94.
- Strathearn L, Mayes LC. Cocaine addiction in mothers: Potential effects on maternal care and infant development. Vol. 1187, Annals of the New York Academy of Sciences. Blackwell Publishing Inc.; 2010. p. 172–83.
- Freire C, Amaya E, Gil F, Murcia M, LLop S, Casas M, et al. Placental metal concentrations and birth outcomes: The Environment and Childhood (INMA) project. Int J Hyg Environ Health [Internet]. 2019 Apr 1 [cited 2020 Mar 15];222(3):468–78. Available from: http://www.ncbi.nlm.nih.gov/pubmed/30638867
- 62. Bank-Nielsen PI, Long M, Bonefeld-Jørgensen EC. Pregnant inuit women's exposure to metals and association with fetal growth outcomes: ACCEPT 2010–2015. Int J Environ Res Public Health [Internet]. 2019 Apr 1 [cited 2020 Mar 15];16(7). Available from: http://www.ncbi.nlm.nih.gov/pubmed/30939809
- 63. Contaminants químics ambientals presents en els aliments.
- van de Bor M. Fetal toxicology. In: Handbook of Clinical Neurology [Internet]. Elsevier B.V.; 2019 [cited 2020 Mar 15]. p. 31–55. Available from: http://www.ncbi.nlm.nih.gov/pubmed/31324317
- 65. Kadawathagedara M, Botton J, de Lauzon-Guillain B, Meltzer HM, Alexander J, Brantsaeter AL, et al. Dietary acrylamide intake during pregnancy and postnatal growth and obesity: Results from the Norwegian Mother and Child Cohort Study (MoBa). Environ Int [Internet]. 2018 Apr 1 [cited 2020 Mar 15];113:325–34. Available from: http://www.ncbi.nlm.nih.gov/pubmed/29398013
- 66. Zhang M, Wang P, Lu Y, Lu X, Zhang A, Liu Z, et al. Bioaccumulation and human exposure of perfluoroalkyl acids (PFAAs) in vegetables from the largest vegetable production base of China. Environ Int [Internet]. 2020 Feb 1 [cited 2020 Mar 15];135:105347. Available from: http://www.ncbi.nlm.nih.gov/pubmed/31794940
- 67. Smith LE, Prendergast AJ, Turner PC, Humphrey JH, Stoltzfus RJ. Aflatoxin exposure during pregnancy, maternal anemia, and adverse birth outcomes [Internet]. Vol. 96, American Journal of Tropical Medicine and Hygiene. American Society of Tropical Medicine and Hygiene; 2017 [cited 2020 Mar 15]. p. 770–6. Available from: http://www.ncbi.nlm.nih.gov/pubmed/28500823
- Couper R, Byard R, Ford A. Maternal diarrhea and ischemic enteritis in newborn twins: Possible association with an hemolytic uremic syndrome outbreak [1] [Internet]. Vol. 25,

Journal of Pediatric Gastroenterology and Nutrition. 1997 [cited 2020 Mar 15]. p. 366. Available from: http://www.ncbi.nlm.nih.gov/pubmed/9285394

- 69. Hosp N, José M, Cordero A, Rodríguez-Blanque R, Carlos Sánchez-García J, Manuel Sánchez-López A, et al. Nutrición Hospitalaria Trabajo Original Pediatría Correspondencia: Infl uencia del ejercicio físico durante el embarazo sobre el peso del recién nacido: un ensayo clínico aleatorizado Infl uence of physical exercise during pregnancy on newborn weight: a r. Nutr Hosp [Internet]. 2017;34(4):834–40. Available from: http://dx.doi.org/10.20960/nh.1095
- 70. Kramer MS, McDonald SW. Aerobic exercise for women during pregnancy. Cochrane Database of Systematic Reviews. John Wiley and Sons Ltd; 2009.
- 71. Schlüssel MM, De Souza EB, Reichenheim ME, Kac G. Physical activity during pregnancy and maternal-child health outcomes: A systematic literature review. Vol. 24, Cadernos de Saude Publica. 2008.
- 72. da Silva SG, Ricardo LI, Evenson KR, Hallal PC. Leisure-Time Physical Activity in Pregnancy and Maternal-Child Health: A Systematic Review and Meta-Analysis of Randomized Controlled Trials and Cohort Studies. Vol. 47, Sports Medicine. Springer International Publishing; 2017. p. 295–317.
- 73. Di Mascio D, Magro-Malosso ER, Saccone G, Marhefka GD, Berghella V. Exercise during pregnancy in normal-weight women and risk of preterm birth: a systematic review and meta-analysis of randomized controlled trials [Internet]. Vol. 215, American Journal of Obstetrics and Gynecology. Mosby Inc.; 2016 [cited 2020 Mar 16]. p. 561–71. Available from: http://www.ncbi.nlm.nih.gov/pubmed/27319364
- 74. Sanabria-Martínez G, Poyatos-León R, Notario-Pacheco B, Álvarez-Bueno C, Cavero-Redondo I, Martinez-Vizcaino V. Effects of physical exercise during pregnancy on mothers' and neonates' health: A protocol for an umbrella review of systematic reviews and meta-analysis of randomised controlled trials. BMJ Open. 2019 Sep 1;9(9).
- 75. Evenson KR, Barakat R, Brown WJ, Dargent-Molina P, Haruna M, Mikkelsen EM, et al. Guidelines for Physical Activity During Pregnancy: Comparisons From Around the World. Vol. 8, American Journal of Lifestyle Medicine. SAGE Publications Inc.; 2014. p. 102–21.
- 76. USEPA USEPA. Air Quality Criteria for Particulate Matter October 2004, Volume 2. Air Qual Criteria Part Matter [Internet]. 2004;II(October):1148. Available from: file:///C:/Users/Laíssa/Downloads/VOL_II_FINAL_PM_AQCD_OCT2004.PDF
- 77. Brook RD, Rajagopalan S, Pope CA, Brook JR, Bhatnagar A, Diez-Roux A V., et al. Particulate matter air pollution and cardiovascular disease: An update to the scientific statement from the american heart association. Vol. 121, Circulation. 2010. p. 2331–78.
- 78. Liu Y, Wang L, Wang F, Li C. Effect of Fine Particulate Matter (PM2.5) on Rat Placenta Pathology and Perinatal Outcomes. Med Sci Monit [Internet]. 2016 Sep 15 [cited 2020 Mar 16];22:3274–80. Available from: http://www.ncbi.nlm.nih.gov/pubmed/27629830
- Donaldson K, Stone V, Seaton A, MacNee W. Ambient particle inhalation and the cardiovascular system: potential mechanisms. Environ Health Perspect [Internet]. 2001 Aug [cited 2020 Mar 16];109(suppl 4):523–7. Available from: https://ehp.niehs.nih.gov/doi/10.1289/ehp.01109s4523
- 80. Editor S. Molecular and Integrative Toxicology.
- Nemmar A, Vanbilloen H, Hoylaerts MF, Hoet PHM, Verbruggen A, Nemery B. Passage of intratracheally instilled ultrafine particles from the lung into the systemic circulation in hamster. Am J Respir Crit Care Med [Internet]. 2001 Nov 1 [cited 2020 Mar 16];164(9):1665–8. Available from: http://www.ncbi.nlm.nih.gov/pubmed/11719307

- 82. Rudra CB, Williams MA, Sheppard L, Koenig JQ, Schiff MA. Ambient Carbon Monoxide and Fine Particulate Matter in Relation to Preeclampsia and Preterm Delivery in Western Washington State. Environ Health Perspect [Internet]. 2011 Jun [cited 2020 Mar 16];119(6):886–92. Available from: https://ehp.niehs.nih.gov/doi/10.1289/ehp.1002947
- 83. Van Den Hooven EH, De Kluizenaar Y, Pierik FH, Hofman A, Van Ratingen SW, Zandveld PYJ, et al. Air pollution, blood pressure, and the risk of hypertensive complications during pregnancy: The generation r study. Hypertension [Internet]. 2011 Mar [cited 2020 Mar 16];57(3):406–12. Available from: http://www.ncbi.nlm.nih.gov/pubmed/21220700
- 84. Olsson D, Mogren I, Eneroth K, Forsberg B. Traffic pollution at the home address and pregnancy outcomes in Stockholm, Sweden. BMJ Open. 2015;5(8):1–8.
- 85. Pedersen M, Stayner L, Slama R, Sørensen M, Figueras F, Nieuwenhuijsen MJ, et al. Ambient air pollution and pregnancy-induced hypertensive disorders: A systematic review and meta-analysis. Hypertension [Internet]. 2014 Sep [cited 2020 Mar 16];64(3):494–500. Available from: http://www.ncbi.nlm.nih.gov/pubmed/24935943
- Fu L, Chen Y, Yang X, Yang Z, Liu S, Pei L, et al. The associations of air pollution exposure during pregnancy with fetal growth and anthropometric measurements at birth: a systematic review and meta-analysis. Environ Sci Pollut Res [Internet]. 2019;26(20):20137–47. Available from: https://doi.org/10.1007/s11356-019-05338-0
- 87. Yuan L, Zhang Y, Gao Y, Tian Y. Maternal fine particulate matter (PM2.5) exposure and adverse birth outcomes: an updated systematic review based on cohort studies. Environ Sci Pollut Res Int [Internet]. 2019 May [cited 2020 Mar 16];26(14):13963–83. Available from: http://www.ncbi.nlm.nih.gov/pubmed/30891704
- 88. Li X, Huang S, Jiao A, Yang X, Yun J, Wang Y, et al. Association between ambient fine particulate matter and preterm birth or term low birth weight: An updated systematic review and meta-analysis. Environ Pollut [Internet]. 2017 Aug [cited 2020 Mar 16];227:596–605. Available from: http://www.ncbi.nlm.nih.gov/pubmed/28457735
- Malmqvist E, Liew Z, Källén K, Rignell-Hydbom A, Rittner R, Rylander L, et al. Fetal growth and air pollution - A study on ultrasound and birth measures. Environ Res [Internet]. 2017 Jan 1 [cited 2020 Mar 26];152:73–80. Available from: http://www.ncbi.nlm.nih.gov/pubmed/27741452
- 90. Klepac P, Locatelli I, Korošec S, Künzli N, Kukec A. Ambient air pollution and pregnancy outcomes: A comprehensive review and identification of environmental public health challenges [Internet]. Vol. 167, Environmental Research. Academic Press Inc.; 2018 [cited 2020 Mar 16]. p. 144–59. Available from: http://www.ncbi.nlm.nih.gov/pubmed/30014896
- Siddika N, Rantala AK, Antikainen H, Balogun H, Amegah AK, Ryti NRI, et al. Synergistic effects of prenatal exposure to fine particulate matter (PM2.5) and ozone (O3) on the risk of preterm birth: A population-based cohort study. Environ Res [Internet]. 2019 Sep 1 [cited 2020 Mar 16];176:108549. Available from: http://www.ncbi.nlm.nih.gov/pubmed/31252204
- 92. He XJ, Qin F yun, Hu CL, Zhu M, Tian CQ, Li L. Is gestational diabetes mellitus an independent risk factor for macrosomia: a meta-analysis? [Internet]. Vol. 291, Archives of Gynecology and Obstetrics. Springer Verlag; 2015 [cited 2020 Mar 16]. p. 729–35. Available from: http://www.ncbi.nlm.nih.gov/pubmed/25388922
- 93. Xu X, Liu C, Xu Z, Tzan K, Zhong M, Wang A, et al. Long-term exposure to ambient fine particulate pollution induces insulin resistance and mitochondrial alteration in adipose tissue. Toxicol Sci [Internet]. 2011 Nov [cited 2020 Mar 16];124(1):88–98. Available from: http://www.ncbi.nlm.nih.gov/pubmed/21873646

- 94. Clemente DBP, Vrijheid M, Martens DS, Bustamante M, Chatzi L, Danileviciute A, et al. Prenatal and childhood traffic-related air pollution exposure and telomere length in european children: The HELIX project. Environ Health Perspect [Internet]. 2019 [cited 2020 Mar 16];127(8):87001. Available from: http://www.ncbi.nlm.nih.gov/pubmed/31393792
- 95. Saenen ND, Plusquin M, Bijnens E, Janssen BG, Gyselaers W, Cox B, et al. In utero fine particle air pollution and placental expression of genes in the brain-derived neurotrophic factor signaling pathway: An ENVIRONAGE birth cohort study. Environ Health Perspect [Internet]. 2015 Aug 4 [cited 2020 Mar 16];123(8):834–40. Available from: http://www.ncbi.nlm.nih.gov/pubmed/25816123
- 96. Klocke C, Allen JL, Sobolewski M, Mayer-Pröschel M, Blum JL, Lauterstein D, et al. Neuropathological Consequences of Gestational Exposure to Concentrated Ambient Fine and Ultrafine Particles in the Mouse. Toxicol Sci [Internet]. 2017 [cited 2020 Mar 16];156(2):492–508. Available from: http://www.ncbi.nlm.nih.gov/pubmed/28087836
- 97. Guxens M, Lubczyńska MJ, Muetzel RL, Dalmau-Bueno A, Jaddoe VWV, Hoek G, et al. Air Pollution Exposure During Fetal Life, Brain Morphology, and Cognitive Function in School-Age Children. Biol Psychiatry [Internet]. 2018 Aug 15 [cited 2020 Mar 16];84(4):295–303. Available from: http://www.ncbi.nlm.nih.gov/pubmed/29530279
- 98. Ritz B, Liew Z, Yan Q, Cui X, Virk J, Ketzel M, et al. Air pollution and Autism in Denmark. Environ Epidemiol (Philadelphia, Pa) [Internet]. 2018 Dec [cited 2020 Mar 31];2(4). Available from: http://www.ncbi.nlm.nih.gov/pubmed/31008439
- 99. Mínguez-Alarcón L, Gaskins AJ. Female exposure to endocrine disrupting chemicals and fecundity: A review [Internet]. Vol. 29, Current Opinion in Obstetrics and Gynecology. Lippincott Williams and Wilkins; 2017 [cited 2020 Mar 16]. p. 202–11. Available from: http://www.ncbi.nlm.nih.gov/pubmed/28557831
- 100. Yang C, Song G, Lim W. A mechanism for the effect of endocrine disrupting chemicals on placentation. Vol. 231, Chemosphere. Elsevier Ltd; 2019. p. 326–36.
- 101. Kahn LG, Trasande L. Environmental Toxicant Exposure and Hypertensive Disorders of Pregnancy: Recent Findings [Internet]. Vol. 20, Current Hypertension Reports. Current Medicine Group LLC 1; 2018 [cited 2020 Mar 16]. p. 87. Available from: http://www.ncbi.nlm.nih.gov/pubmed/30090982
- 102. Sabra S, Malmqvist E, Saborit A, Gratacós E, Gomez Roig MD. Heavy metals exposure levels and their correlation with different clinical forms of fetal growth restriction. PLoS One [Internet]. 2017 Oct 1 [cited 2020 Mar 16];12(10):e0185645. Available from: http://www.ncbi.nlm.nih.gov/pubmed/28985223
- 103. Xu Y, Cook TJ, Knipp GT. Effects of di-(2-ethylhexyl)-phthalate (DEHP) and its metabolites on fatty acid homeostasis regulating proteins in rat placental HRP-1 trophoblast cells. Toxicol Sci [Internet]. 2005 Apr [cited 2020 Mar 16];84(2):287–300. Available from: http://www.ncbi.nlm.nih.gov/pubmed/15647598
- Kaufmann P, Black S, Huppertz B. Endovascular Trophoblast Invasion: Implications for the Pathogenesis of Intrauterine Growth Retardation and Preeclampsia. Biol Reprod. 2003 Jul 1;69(1):1–7.
- 105. Tait S, Tassinari R, Maranghi F, Mantovani A. Bisphenol A affects placental layers morphology and angiogenesis during early pregnancy phase in mice. J Appl Toxicol [Internet]. 2015 Nov 1 [cited 2020 Mar 16];35(11):1278–91. Available from: http://www.ncbi.nlm.nih.gov/pubmed/26063408
- 106. Al-Saleh I, Al-Doush I, Alsabbaheen A, Mohamed GED, Rabbah A. Levels of DDT and its metabolites in placenta, maternal and cord blood and their potential influence on neonatal anthropometric measures. Sci Total Environ [Internet]. 2012 Feb 1 [cited 2020

Mar 16];416:62-74. Available from: http://www.ncbi.nlm.nih.gov/pubmed/22192892

- 107. Tyagi V, Garg N, Mustafa MD, Banerjee BD, Guleria K. Organochlorine pesticide levels in maternal blood and placental tissue with reference to preterm birth: a recent trend in North Indian population. Environ Monit Assess [Internet]. 2015 Jul 1 [cited 2020 Mar 16];187(7):471. Available from: http://www.ncbi.nlm.nih.gov/pubmed/26122123
- 108. Von Ehrenstein OS, Ling C, Cui X, Cockburn M, Park AS, Yu F, et al. Prenatal and infant exposure to ambient pesticides and autism spectrum disorder in children: Population based case-control study. BMJ [Internet]. 2019 [cited 2020 Mar 31];364:I962. Available from: http://www.ncbi.nlm.nih.gov/pubmed/30894343
- 109. Li ZM, Hernandez-Moreno D, Main KM, Skakkebæk NE, Kiviranta H, Toppari J, et al. Association of in Utero Persistent Organic Pollutant Exposure with Placental Thyroid Hormones. Endocrinology [Internet]. 2018 Oct 1 [cited 2020 Mar 16];159(10):3473–81. Available from: http://www.ncbi.nlm.nih.gov/pubmed/30059991
- 110. Cornelius DC. Preeclampsia: From inflammation to immunoregulation. Clin Med Insights Blood Disord. 2018 Jan 1;11.
- 111. Zhu Y, Tan YQ, Wang CC, Leung LK. The flame retardant 2,2",4,4"-Tetrabromodiphenyl ether enhances the expression of corticotropin-releasing hormone in the placental cell model JEG-3. Chemosphere [Internet]. 2017 May [cited 2020 Mar 16];174:499–505. Available from: http://www.ncbi.nlm.nih.gov/pubmed/28189027
- 112. Baldacci S, Gorini F, Santoro M, Pierini A, Minichilli F, Bianchi F. Environmental and individual exposure and the risk of congenital anomalies: A review of recent epidemiological evidence. Epidemiol Prev [Internet]. 2018 [cited 2020 Mar 31];42(3– 4):1–34. Available from: http://www.ncbi.nlm.nih.gov/pubmed/30066535
- 113. Spinder N, Prins JR, Bergman JEH, Smidt N, Kromhout H, Boezen HM, et al. Congenital anomalies in the offspring of occupationally exposed mothers: a systematic review and meta-analysis of studies using expert assessment for occupational exposures. Hum Reprod [Internet]. 2019 May 1 [cited 2020 Mar 31];34(5):903–19. Available from: http://www.ncbi.nlm.nih.gov/pubmed/30927411
- 114. Bowen SE, Hannigan JH. Developmental toxicity of prenatal exposure to toluene. AAPS J. 2006 Jun;8(2):E419–24.
- Vandenberg LN, Colborn T, Hayes TB, Heindel JJ, Jacobs DR, Lee DH, et al. Hormones and endocrine-disrupting chemicals: Low-dose effects and nonmonotonic dose responses. Vol. 33, Endocrine Reviews. 2012. p. 378–455.
- 116. Van Maele-Fabry G, Lantin AC, Hoet P, Lison D. Childhood leukaemia and parental occupational exposure to pesticides: A systematic review and meta-analysis [Internet]. Vol. 21, Cancer Causes and Control. 2010 [cited 2020 Mar 31]. p. 787–809. Available from: http://www.ncbi.nlm.nih.gov/pubmed/20467891
- 117. Grohs MN, Reynolds JE, Liu J, Martin JW, Pollock T, Lebel C, et al. Prenatal maternal and childhood bisphenol a exposure and brain structure and behavior of young children. Environ Heal A Glob Access Sci Source [Internet]. 2019 Oct 15 [cited 2020 Mar 31];18(1):85. Available from: http://www.ncbi.nlm.nih.gov/pubmed/31615514
- 118. Ejaredar M, Lee Y, Roberts DJ, Sauve R, Dewey D. Bisphenol A exposure and children's behavior: A systematic review. J Expo Sci Environ Epidemiol [Internet]. 2017 Mar 1 [cited 2020 Mar 31];27(2):175–83. Available from: http://www.ncbi.nlm.nih.gov/pubmed/26956939
- 119. Basner M, Babisch W, Davis A, Brink M, Clark C, Janssen S, et al. Auditory and nonauditory effects of noise on health. Vol. 383, The Lancet. Lancet Publishing Group; 2014. p. 1325–32.

- 120. Münzel T, Gori T, Babisch W, Basner M. Cardiovascular effects of environmental noise exposure. Eur Heart J. 2014;35(13):829–36.
- 121. van Kempen E, Casas M, Pershagen G, Foraster M. WHO environmental noise guidelines for the European region: A systematic review on environmental noise and cardiovascular and metabolic effects: A summary [Internet]. Vol. 15, International Journal of Environmental Research and Public Health. MDPI AG; 2018 [cited 2020 Mar 16]. Available from: http://www.ncbi.nlm.nih.gov/pubmed/29470452
- 122. Münzel T, Sørensen M, Gori T, Schmidt FP, Rao X, Brook FR, et al. Environmental stressors and cardio-metabolic disease: part II-mechanistic insights. Eur Heart J [Internet]. 2017 Feb 21 [cited 2020 Mar 16];38(8):557–64. Available from: http://www.ncbi.nlm.nih.gov/pubmed/27460891
- 123. WHO/Europe | Noise [Internet]. [cited 2020 Mar 16]. Available from: http://www.euro.who.int/en/health-topics/environment-and-health/noise
- 124. Auger N, Duplaix M, Bilodeau-Bertrand M, Lo E, Smargiassi A. Environmental noise pollution and risk of preeclampsia. Environ Pollut [Internet]. 2018 Aug 1 [cited 2020 Mar 16];239:599–606. Available from: http://www.ncbi.nlm.nih.gov/pubmed/29704672
- 125. Pedersen M, Halldorsson TI, Olsen SF, Hjortebjerg D, Ketzel M, Grandström C, et al. Impact of road traffic pollution on pre-eclampsia and pregnancy-induced hypertensive disorders. Epidemiology [Internet]. 2017 Jan 1 [cited 2020 Mar 16];28(1):99–106. Available from: http://www.ncbi.nlm.nih.gov/pubmed/27648591
- 126. Bechi N, letta F, Romagnoli R, Focardi S, Corsi I, Buffi C, et al. Estrogen-Like Response to p-Nonylphenol in Human First Trimester Placenta and BeWo Choriocarcinoma Cells. Toxicol Sci [Internet]. 2006 Jun 21;93(1):75–81. Available from: https://doi.org/10.1093/toxsci/kfl043
- 127. Paulesu L, Rao C V., letta F, Pietropolli A, Ticconi C. HCG and its disruption by environmental contaminants during human pregnancy. Vol. 19, International Journal of Molecular Sciences. MDPI AG; 2018.
- 128. Ferguson KK, McElrath TF, Cantonwine DE, Mukherjee B, Meeker JD. Phthalate metabolites and bisphenol-A in association with circulating angiogenic biomarkers across pregnancy. Placenta [Internet]. 2015 Jun 1 [cited 2020 Mar 17];36(6):699–703. Available from: http://www.ncbi.nlm.nih.gov/pubmed/25913709
- Ferguson KK, Cantonwine DE, McElrath TF, Mukherjee B, Meeker JD. Repeated measures analysis of associations between urinary bisphenol-A concentrations and biomarkers of inflammation and oxidative stress in pregnancy. Reprod Toxicol [Internet].
 2016 Dec 1 [cited 2020 Mar 17];66:93–8. Available from: http://www.ncbi.nlm.nih.gov/pubmed/27751756
- 130. Sieppi E, Vähäkangas K, Rautio A, letta F, Paulesu L, Myllynen P. The xenoestrogens, bisphenol A and para-nonylphenol, decrease the expression of the ABCG2 transporter protein in human term placental explant cultures. Mol Cell Endocrinol [Internet]. 2016 Jul 5 [cited 2020 Mar 17];429:41–9. Available from: http://www.ncbi.nlm.nih.gov/pubmed/27036933
- 131. Peltier MR, Klimova NG, Arita Y, Gurzenda EM, Murthy A, Chawala K, et al. Polybrominated diphenyl ethers enhance the production of proinflammatory cytokines by the placenta. Placenta [Internet]. 2012 Sep [cited 2020 Mar 17];33(9):745–9. Available from: http://www.ncbi.nlm.nih.gov/pubmed/22749501
- 132. Ruis MT, Rock KD, Hall SM, Horman B, Patisaul HB, Stapleton HM. PBDEs Concentrate in the Fetal Portion of the Placenta: Implications for Thyroid Hormone Dysregulation. Endocrinology [Internet]. 2019 Nov 1 [cited 2020 Mar 17];160(11):2748–58. Available from: http://www.ncbi.nlm.nih.gov/pubmed/31555822

- 133. Suter MA, Aagaard KM, Coarfa C, Robertson M, Zhou G, Jackson BP, et al. Association between elevated placental polycyclic aromatic hydrocarbons (PAHs) and PAH-DNA adducts from Superfund sites in Harris County, and increased risk of preterm birth (PTB). Biochem Biophys Res Commun [Internet]. 2019 Aug 20 [cited 2020 Mar 17];516(2):344–9. Available from: http://www.ncbi.nlm.nih.gov/pubmed/31208719
- 134. Punshon T, Li Z, Jackson BP, Parks WT, Romano M, Conway D, et al. Placental metal concentrations in relation to placental growth, efficiency and birth weight. Environ Int [Internet]. 2019 May 1 [cited 2020 Mar 17];126:533–42. Available from: http://www.ncbi.nlm.nih.gov/pubmed/30851484
- 135. Olsson D, Mogren I, Forsberg B. Air pollution exposure in early pregnancy and adverse pregnancy outcomes: A register-based cohort study. BMJ Open [Internet]. 2013 [cited 2020 Mar 17];3(2). Available from: http://www.ncbi.nlm.nih.gov/pubmed/23386578
- 136. Saenen ND, Vrijens K, Janssen BG, Madhloum N, Peusens M, Gyselaers W, et al. Placental Nitrosative Stress and Exposure to Ambient Air Pollution During Gestation: A Population Study. Am J Epidemiol [Internet]. 2016 [cited 2020 Mar 17];184(6):442–9. Available from: http://www.ncbi.nlm.nih.gov/pubmed/27601048
- Neven KY, Saenen ND, Tarantini L, Janssen BG, Lefebvre W, Vanpoucke C, et al. Placental promoter methylation of DNA repair genes and prenatal exposure to particulate air pollution: an ENVIRONAGE cohort study. Lancet Planet Heal [Internet]. 2018 Apr 1 [cited 2020 Mar 17];2(4):e174–83. Available from: http://www.ncbi.nlm.nih.gov/pubmed/29615218
- 138. García-Mantrana I, Bertua B, Martínez-Costa C, Collado MC. Perinatal nutrition: How to take care of the gut microbiota? Clin Nutr Exp [Internet]. 2016;6:3–16. Available from: http://dx.doi.org/10.1016/j.yclnex.2016.02.002
- Reece-Stremtan S, Marinelli KA. ABM clinical protocol #21: guidelines for breastfeeding and substance use or substance use disorder, revised 2015. Breastfeed Med [Internet]. 2015 Apr;10(3):135–41. Available from: https://pubmed.ncbi.nlm.nih.gov/25836677
- Rowe H, Baker T, Hale TW. Maternal Medication, Drug Use, and Breastfeeding [Internet]. Vol. 60, Pediatric Clinics of North America. 2013 [cited 2020 Mar 17]. p. 275– 94. Available from: http://www.ncbi.nlm.nih.gov/pubmed/23178070
- 141. Eidelman AI, Schanler RJ. Breastfeeding and the use of human milk. Vol. 129, Pediatrics. 2012.
- 142. Esteban M, Schindler BK, Jiménez JA, Koch HM, Angerer J, Rosado M, et al. Mercury analysis in hair: Comparability and quality assessment within the transnational COPHES/DEMOCOPHES project. Environ Res. 2015 Aug 1;141:24–30.
- 143. Aids P. Scientific Opinion on the risks to public health related to the presence of bisphenol A (BPA) in foodstuffs. EFSA J. 2015;13(1).
- 144. Migeot Virginie A4 Dupuis, Antoine A4 Cariot, Axelle A4 Albouy-Llaty, Marion A4 Pierre, Fabrice A4 Rabouan, Sylvie VA-M. Bisphenol A and Its Chlorinated Derivatives in Human Colostrum. Environ Sci Technol. 2013;v. 47(23):13791-13797–2013 v.47 no.23.
- 145. Mercogliano R, Santonicola S. Investigation on bisphenol A levels in human milk and dairy supply chain: A review [Internet]. Vol. 114, Food and Chemical Toxicology. Elsevier Ltd; 2018 [cited 2020 Mar 17]. p. 98–107. Available from: http://www.ncbi.nlm.nih.gov/pubmed/29448092
- 146. Dualde P, Pardo O, Corpas-Burgos F, Kuligowski J, Gormaz M, Vento M, et al. Biomonitoring of bisphenols A, F, S in human milk and probabilistic risk assessment for breastfed infants. Sci Total Environ [Internet]. 2019 Jun 10 [cited 2020 Mar 17];668:797–

805. Available from: http://www.ncbi.nlm.nih.gov/pubmed/30870748

- 147. Mallozzi M, Bordi G, Garo C, Caserta D. The effect of maternal exposure to endocrine disrupting chemicals on fetal and neonatal development: A review on the major concerns [Internet]. Vol. 108, Birth Defects Research Part C - Embryo Today: Reviews. John Wiley and Sons Inc.; 2016 [cited 2020 Mar 17]. p. 224–42. Available from: http://www.ncbi.nlm.nih.gov/pubmed/27653964
- 148. Dualde P, Pardo O, Corpas-Burgos F, Kuligowski J, Gormaz M, Vento M, et al. Biomonitoring of parabens in human milk and estimated daily intake for breastfed infants. Chemosphere [Internet]. 2020 Feb 1 [cited 2020 Mar 17];240:124829. Available from: http://www.ncbi.nlm.nih.gov/pubmed/31563722
- 149. Sanchis Y, Coscollà C, Corpas-Burgos F, Vento M, Gormaz M, Yusà V. Biomonitoring of bisphenols A, F, S and parabens in urine of breastfeeding mothers: Exposure and risk assessment. Environ Res [Internet]. 2020 Jun 1 [cited 2020 Apr 13];185:109481. Available from: https://www.sciencedirect.com/science/article/abs/pii/S0013935120303741
- 150. Iszatt N, Janssen S, Lenters V, Dahl C, Stigum H, Knight R, et al. Environmental toxicants in breast milk of Norwegian mothers and gut bacteria composition and metabolites in their infants at 1 month. Microbiome [Internet]. 2019 Feb 27 [cited 2020 Mar 17];7(1):34. Available from: http://www.ncbi.nlm.nih.gov/pubmed/30813950
- 151. Lenters V, Iszatt N, Forns J, Čechová E, Kočan A, Legler J, et al. Early-life exposure to persistent organic pollutants (OCPs, PBDEs, PCBs, PFASs) and attention-deficit/hyperactivity disorder: A multi-pollutant analysis of a Norwegian birth cohort. Environ Int [Internet]. 2019 Apr 1 [cited 2020 Mar 17];125:33–42. Available from: http://www.ncbi.nlm.nih.gov/pubmed/30703609
- 152. Sabarwal A, Kumar K, Singh RP. Hazardous effects of chemical pesticides on human health–Cancer and other associated disorders. Vol. 63, Environmental Toxicology and Pharmacology. Elsevier B.V.; 2018. p. 103–14.
- 153. Kao CC, Que DE, Bongo SJ, Tayo LL, Lin YH, Lin CW, et al. Residue levels of organochlorine pesticides in breast milk and its associations with cord blood thyroid hormones and the offspring's neurodevelopment. Int J Environ Res Public Health [Internet]. 2019 Apr 2 [cited 2020 Mar 17];16(8). Available from: http://www.ncbi.nlm.nih.gov/pubmed/31018505
- 154. Escuder-Vieco D, Garcia-Algar Ó, Joya X, Marchei E, Pichini S, Pacifici R, et al. Breast Milk and Hair Testing to Detect Illegal Drugs, Nicotine, and Caffeine in Donors to a Human Milk Bank. J Hum Lact. 2016;32(3):542–5.