Check for updates

OPEN ACCESS

EDITED AND REVIEWED BY Kenneth Fields, University of Kentucky, United States

*CORRESPONDENCE Harapriya Mohapatra Mm@niser.ac.in Jesús Arenas jarenasbusto@gmail.com

RECEIVED 24 August 2023 ACCEPTED 29 August 2023 PUBLISHED 07 September 2023

CITATION

Mohapatra H and Arenas J (2023) Editorial: Novel insights on the role of bacterial membrane proteins in virulence and pathogenesis. *Front. Cell. Infect. Microbiol.* 13:1282672. doi: 10.3389/fcimb.2023.1282672

COPYRIGHT

© 2023 Mohapatra and Arenas. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Editorial: Novel insights on the role of bacterial membrane proteins in virulence and pathogenesis

Harapriya Mohapatra^{1,2*} and Jesús Arenas^{3,4*}

¹School of Biological Sciences, National Institute of Science Education and Research, Jatani, India, ²Homi Bhabha National Institute, BARC Training School Complex, Mumbai, India, ³Unit of Microbiology and Immunology, University of Zaragoza, Faculty of Veterinary, Zaragoza, Spain, ⁴Institute Agrofood of Aragón-IA2, University of Zaragoza-Centro de Investigación y Tecnología Agroalimentaria de Aragón (CITA), Zaragoza, Spain

KEYWORDS

infection, membrane proteins, virulence, pathogenesis, host-pathogen interaction

Editorial on the Research Topic

Novel insights on the role of bacterial membrane proteins in virulence and pathogenesis

In 1884, Christian Gram's staining revealed the differential nature of cell envelopes in two classes of bacteria. Almost a decade later Glauert and Thornley (1969) using an electron -microscopy technique evidenced a distinct two-layered structure of Gramnegative bacteria versus a thick layer of Gram-positive bacteria. Later Miura and Mizushima (1968) and Osborn et al. (1972) developed biochemical methods to characterize cytoplasmatic and outer membrane layers. Bacterial membranes are not only important to maintain cell integrity and regulate the import of nutrients/ antimicrobials but, also play relevant functions in pathogenesis functioning as a platform to expose a repertoire of virulence factors. These factors include associated or integral membrane proteins, as those studied in this Research Topic. Besides, the discovery of membrane vesicles reiterated the significant role of the composition of both membranes in virulence and pathogenesis. With this background, the current thematic issue was aimed to invite novel findings on how bacterial membrane proteins facilitate and/or modulate pathogenesis. At the end, five articles were edited; four of which are original research articles. Two articles report novel functions of cell surface collagen binding adhesins in Gram-positive pathogens Streptococcus sp while the other two unveil the role of outer membrane proteins in the Gram-negative pathogen Haemophilus influenzae.

Original research by Naka et al., elucidated the role of collagen-binding protein Cnm of *Streptococcus mutans*. Investigations of the Cnm-deficient isogenic mutant strains showed that indeed, Cnm is a relevant protein for collagen binding in this bacterium. Diverse electron microscopy assays showed that the protein is located around the bacteria cell wall and produces protrusions at the membrane surface. Comparative RNA-seq assays evidenced that the lack of Cnm production altered the expression of several genes including biofilm formation-associated genes, and, indeed, biofilm formation assays evidenced that Cnm contributes to biofilm formation in the absence of collagen.

10.3389/fcimb.2023.1282672

Streptococcus suis encodes a collagen-binding protein called Cba. By exploring the phenotypes of a Cba-depletion mutant, authors demonstrated that Cba has implications in biofilm formation and phagocytic resistance to macrophages with relevant implications in virulence as revealed by an attenuated phenotype in mice infection models. Immunization of mice with purified protein induced strong antibody responses, but resulted in no protection and caused organ lessons and mortality emulating a phenomenon called antibody-dependent enhancement of infection, previously described for other proteins (Halstead et al., 2010).

The cell wall is an interconnected matrix of peptidoglycan that surrounds the cytoplasmic membrane and protects the cell from osmotic lysis. Peptidoglycan biosynthesis comprises the synthesis of precursors in the cytoplasm, their transport to the outer face of the cytoplasmic membrane, and their subsequent cross-linking into the existing peptidoglycan layer. The latter process is mediated by enzymes with peptidoglycan-glycosyl transferase and transpeptidase activities. The most studied enzymes are penicillinbinding proteins (PBPs), which are classified as PBP type A and PBP type B. Early work showed that the activity of some PBPs is regulated by lipoproteins, i.e. LpoA and LpoB. Both proteins activate transpeptidase and the peptidoglycan polymerization of PBP1a and PBP1b, respectively (Sardis et al., 2021). A research article by Jalalvand et al. investigated the role of LpoA in the formation of outer membrane vesicles (OMVs) in the respiratory tract pathogen Haemophilus influenzae strain 3655. Fluorescence microscopy assays revealed that LipoA accumulated in OMVs and was located in the septum during cell division. Through reporter fusion protein studies, the authors have unveiled that the C-terminal domain of the protein is required for its transportation to OMVs.

Further, in a brief report Su et al., investigated the outer membrane protein of P5 from non-typeable *Haemophilus influenzae.* P5 is structured in a β -barrel transmembrane domain that contains four extracellular loops and a C-terminal domain (Novotny and Bakaletz, 2003). The authors showed that P5 binds to the peptidoglycan layer. Deletion of P5 generated a viable mutant that exhibited a reduced abundance of virulence factors in the outer membrane and periplasmic chaperones. Depletion of P5 caused a variety of phenotypes including, i) reduced adherence to cells, ii) reduced binding to fibronectin, iii) reduced survival to complement mediate killing, and iv) enhanced OMV production. This effect was majorly caused by the depletion of its C-terminal domain.

The development of new ideas and strategies to counter bacterial pathogens is of utmost importance in this post-antibiotic era, where unprecedented escalation of drug-resistant organisms is being observed. Capsular vaccines and bacterins gained attention as these function by stimulating immune response. The perspective by Ewasechko et al., provides insights from the study of transferring binding proteins of human and animal pathogens with demonstrated potential in vaccine development. The authors also speculate that the presence of these receptors in pathogenic bacteria of the upper respiratory tract of human and porcine pathogens could be related to the differential expression of transferrin between the subepithelial space and the air-liquid interface that varies iron availability. Therefore, transferrin-binding proteins could contribute to iron acquisition during invasion. This is supported by the presence of these proteins in human and porcine pathogens and their lack of commensal species.

All together, we hope this Research Topic provides valuable insights into the role of bacterial membranes in virulence and pathogenesis and the development of novel vaccines.

Author contributions

HM: Writing – original draft, Writing – review & editing. JA: Writing – original draft, Writing – review & editing.

Funding

JA received funding from Gobierno de Aragón (Department of Science, University and knowledge Society) (Project TRANSIT, Grant agreement LMP58_21) and from Ministerio de Ciencia e Innovación/Agencia Española de Investigación MCIN/AEI/ 10.13039/501100011033 (Project ABC-VACCINESs, Grant agreement PID2020-114617RB-100).

Acknowledgments

We thank the authors of the papers published on this Research Topic.

Conflict of interest

JA is co-author of a patent for human vaccines and received funding for research from diverse vaccine companies.

The remaining author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

Glauert, A. M., and Thornley, M. J. (1969). The topography of the bacterial cell wall. Ann. Rev. Micro 23, 159–198. doi: 10.1146/annurev.mi.23.100169.001111

Halstead, S. B., Mahalingam, S., Marovich, M. A., Ubol, S., and Mosser, D. M. (2010). Intrinsic antibody-dependent enhancement of microbial infection in macrophages: disease regulation by immune complexes. *Lancet Infect. Dis.* 10, 712–722. doi: 10.1016%2FS1473-3099(10)70166-3

Miura, T., and Mizushima, S. (1968). Separation by density gradient centrifugation of two types of membranes from spheroplast membrane of *Escherichia coli* K12. *BBA- Biomembranes* 150, 159–161. doi: 10.1016/0005-2736(68)90020-5

Novotny, L. A., and Bakaletz, L. O. (2003). The fourth surface-exposed region of the outer membrane protein P5-homologous adhesin of nontypable *Haemophilus influenzae* is an immunodominant but nonprotective decoying epitope. J. Immunol. 171, 1978–1983. doi: 10.4049/jimmunol.171.4.1978

Osborn, M. J., Gander, J. E., Parisi, E., and Carson, J. (1972). Mechanism of Assembly of the Outer Membrane of *Salmonella typhimurium*: isolation and characterization of cytoplasmic and outer membrane. *J. Biol. Chem.* 247, 3962–3972. doi: 10.1016/S0021-9258(19)45127-2

Sardis, M. F., Bohrhunter, J. L., Greene, N. G., and Bernhardt, T. G. (2021). The LpoA activator is required to stimulate the peptidoglycan polymerase activity of its cognate cell wall synthase PBP1a. *P.N.A.S.* 118, e2108894118. doi: 10.1073/pnas.2108894118