Design and characterization of macroporous alumina membranes for 1

#### passive samplers of water contaminants. 2

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#### 11 ABSTRACT

12 Macroporous ceramic membranes with hierarchical pore structure were developed and 13 characterized for the time-integrated analysis of water contaminants. The alumina membranes 14 were boosted to allow a suitable diffusion rate for envisaged applications. Sacrificial fugitives 15 as cornstarch and graphite and partial sintering technique were utilized for the control of pore 16 structure and morphology. The present study includes: (i) Development of ceramic membrane 17 preparation methods based on wet forming techniques and partial sintering in order to control 18 of membrane parameters such as porosity, tortuosity factor and thickness; (ii) Microstructural 19 characterization of the ceramic membranes using two complementary techniques: Hg-20 porosimetry and SEM; (iii) the diffusion tests performed under laboratory conditions by an 21 innovative, easy and accurate method based in the use of methylene blue  $(C_{16}H_{18}CIN_3S)$  water 22 solutions as test compound to study the characteristics of ceramic membranes. Using slip 23 casting and partial sintering processing three different Al<sub>2</sub>O<sub>3</sub> membrane configurations with 24 high values 34.5, 36.9 and 39.6 vol% of interconnected porosity and 1.75, 1.7 and 1.3 mm 25 thickness were prepared. The pore morphology consisted of spherical cavities of 5-10 µm 26 diameter and small pores of 70-200 nm. This study also gathers the basis for the calculation of 27 diffusion coefficients, membrane geometrical factor and sampling rates using methylene blue 28 water solutions whose concentration can be accurately determined by optical absorption measurements at 664 nm. Diffusion coefficients for methylene blue range from 2.3±0.5E-8 to 29 30 3.0±0.5E-8 cm<sup>2</sup> s<sup>-1</sup> increasing as connected porosity increases and membrane thickness decreases. The membrane geometrical factors are low, between 0.6 and 1E-02 and Archie's 31 32 constants large 4.5 to 5.5. This has been associated to the bimodal pore structure with 33 coexistence of large and small pores that may increase the tortuosity factors in membranes. 34 The macroporous ceramic membranes herein developed are suitable for the fabrication of 35 passive samplers aimed at preconcentrating organic contaminants in water.

#### 36 SHORT ABSTRACT

Macroporous ceramic membranes with hierarchical pore structure were developed and 37 38 characterized for the time-integrated analysis of water contaminants. The alumina membranes 39 were boosted to allow a suitable diffusion rate for envisaged applications. Sacrificial fugitives 40 as cornstarch and graphite and partial sintering technique were utilized for the control of pore 41 structure and morphology. The present study includes: (i) Development of ceramic membrane preparation methods based on wet forming techniques and partial sintering in order to control 42 43 of membrane parameters such as porosity, tortuosity factor and thickness; (ii) Microstructural 44 characterization of the ceramic membranes using two complementary techniques: Hg-45 porosimetry and SEM; (iii) the diffusion tests performed under laboratory conditions by an 46 innovative, easy and accurate method based in the use of methylene blue (C16H18CIN3S) 47 water solutions as test compound to study the characteristics of ceramic membranes.

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49 *Keywords:* Macroporous Ceramic Membrane; Hierarchical Pore Structure; Al<sub>2</sub>O<sub>3</sub>; Diffusion

50 coefficient measurement; Passive Sampler.

## 51 **1. Introduction**

52 Concern on the increasing presence of toxic water contaminants has led to the 53 necessity to develop new monitoring tools. Emerging and priority organic contaminants are 54 found in wastewaters and rivers at concentrations that produce impairment. However, their 55 incidence and real environmental risks are often neglected due in part to sampling difficulties. 56 In order to properly formulate preservation regulations, more information about the presence 57 of these contaminants in the environment is necessary. Environmental monitoring tools require the use of efficient and reliable sampling and analytical techniques that should be 58 59 straightforward to use, robust and stable under monitoring conditions. Currently, grab sampling is the most common way to monitor surface waters although it might not be 60 61 completely representative if contamination fluctuates with time. For snap-shot sampling a 62 given volume of water is collected at a given time using bailers or pumps. Limitations to active 63 sampling are well known and have been described elsewhere [1]. Alternatively, passive or 64 diffusive samplers (PS) offer information on average pollution levels over several hours or days 65 [2]. PS devices allow the diffusion of contaminants through the membrane due to the difference in chemical potentials between the external and internal surface of the sampler [3]. 66 67 A sorbent inside the sampler retains the diffusing species. The device efficiency and 68 performance depends on the characteristics of the membrane as well as the polarity and 69 solubility of the contaminants, the nature of the sampled water and properties of the receiving 70 phase. The main advantages of this technique are that it provides time-integrated 71 concentration values, providing more descriptive information in environments where the 72 concentration of the contaminants is not constant [1], autonomy as external energy source is 73 not needed and easy use. Several PS devices have been developed and applied for the analysis 74 of contaminants in water [4] at levels between 0.01 and 1200 μg L<sup>-1</sup>, and thus, represent a valid 75 method for water monitoring provided they are properly calibrated using controlled 76 conditions.

77 Ceramic dosimeters were first developed at the University of Tübingen [5] and validated for the analysis of polycyclic aromatic hydrocarbons (PAHs) in groundwater [6,7] and 78 79 flame retardants in rivers [8]. They are compact, robust, portable and inexpensive and offer 80 information on average pollution levels from few hours to months. They do not require 81 supervision and can be used in hazardous environments. The ceramic dosimeters consist of a 82 porous cylindrical permeable ceramic membrane containing an adsorbent inside. After the 83 sampling period, the adsorbed analytes are extracted and analyzed. State of the art ceramic 84 membranes have a configuration of 50 mm of length, 1.5 mm thickness and 10 mm outer 85 diameter and the pore diameter is of 5 nm [6]. Tested for PAHs in groundwater, the flow rate of these ceramic dosimeters was of 1.5-2.5 mL d<sup>-1</sup>. The same system provided a similar flow 86 87 rate (0.39-3.7 mL d<sup>-1</sup>) when used for flame-retardants in surface waters, although fouling was 88 observed after 6 days deployment [8].

89 This study focuses on the development of a new ceramic membrane for passive 90 sampling where novelty relies in the achievement of pore structures different than that used 91 previously. The motivation is as follow. In the CPS device the water impregnated at the porous 92 ceramic tube acts as a diffusion barrier for the contaminants, which are thereafter retained by 93 the adsorbent or receiving phase. Since the flow of contaminants and the accumulation in the 94 receiving phase depends on the chemical gradient and the diffusivity of the pollutants through 95 the membrane, there is a critical dependence of the device performance on the membrane 96 structure. In particular, porosity, inertness, mechanical resistance, pore size and diffusion 97 length are the key parameters defining the ceramic membrane. Preferentially, pore volume 98 should be large enough to retain high water volumes also aiming to decrease the unwanted 99 effect of interaction of solute molecules with the ceramic walls. In addition, good 100 interconnectivity between pores enhances solvent diffusion but pore size has to allow 101 capillarity forces to keep the water in the membrane ceramic walls avoiding swelling effects 102 and water flux. Furthermore, the matrix material has to be chemically and mechanically stable 103 for long deployment times in natural waters. The membrane has to present enough robustness 104 to resist tight sealing, water turbulences, etc.

105 All these requirements can be achieved through appropriate choice of materials and 106 forming techniques but keeping in mind the industrial scalability and final price of the device. In this work we produced monolithic ceramic membranes of  $AI_2O_3$  with a combination of 107 108 macro-pore morphologies using sacrificial pore former phases and partial sintering techniques. 109 These porous ceramics are referred to as materials with hierarchical porosity. The fabrication 110 and properties of ceramics with hierarchical porosity has been extensively described in some 111 reviews [9]. To better understand the role of pore structure and membrane dimensions on the 112 up-taking ratio of the CPS, three sets of macro-porous ceramic membranes with different 113 bimodal distribution of pores and membrane thickness were prepared and were calibrated for 114 the time-weighted average determination of contaminant concentrations in water. Their 115 performance was tested using methylene blue water solutions as test compound that allow us 116 to determine some useful membrane parameters such as diffusivity, sampling rate, tortuosity, 117 etc.

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## 119 **2. Experimental**

120 *2.1. Materials and fabrication methods.* 



121

**Fig. 1**. CPS device fabricated with the ceramic membranes described in this paper.

Figure 1 shows a CPS sampler fabricated with one of the ceramic membranes studied in this paper. Observe the rubber caps closing the inner membrane volume where concentrator is placed. The ceramic membranes were fabricated by slip casting method. This is a relatively cheap and reproducible method suitable for large-scale production of nearly to 127 final shape green ceramics. Critical aspect of the fabrication process is the preparation of 128 ceramic suspensions. Ceramic suspensions were prepared using aluminum oxide powders ( $\alpha$ -Al2O3, Ceralox APA-0.5, 8.1 m<sup>2</sup>/g, Sasol North America, Inc., grain size D<sub>50</sub> = 300 nm) and 129 130 water. Suspensions were stabilized using Duramax D-3005 as dispersant (1% in solid weight). 131 Two stable suspensions CP1 and CP2 with different sacrificial phases as pore generator agents, 132 cornstarch or cornstarch and graphite (Alfa Aesar, 325 mesh, 44 µm, 99.9%), were prepared. Suspension compositions are given in Table 1. The morphology of starch powders used in this 133 134 work has been recently published [10]. They present irregular prismatic shapes with a 135 relatively narrow particle size distribution  $D_{50}$  = 10  $\mu$ m as measured by laser diffraction in a 136 Mastersizer 2000 equipment. The rheology of suspensions was controlled to achieve the 137 optimum processing conditions. In particular the composition of the colloids was adjusted to get a viscosity for proper filling of mold without entrapping of air bubbles. 138

139

140 **Table 1**.

141 Suspension composition in weight %.

	CP1	CP2
Al <sub>2</sub> O <sub>3</sub>	58.5	58.0
Water	29.25	27.2
Graphite	0	2.8
Cornstarch	12.25	12
Sacrificial phase volume (%)	37	40

142

143 The volume of the solid organic phases was calculated using the following solid phase density values: alumina density 3.98 g cm<sup>-3</sup>, starch density 1.43 g cm<sup>-3</sup> and graphite density 144 2.16 g cm<sup>-3</sup>. Dispersant and water solvent were first mixed by magnetic stirring for 15 min. 145 146 Alumina powders and pore formation agents were poured into the mix and stirred for 30 min. 147 The resulting paste is treated with ultrasounds for 1 minute and magnetic stirred again for 30 148 min. The stable ceramic paste was poured in a plaster mold with the desired cylindrical 149 geometry. The time that the suspension remains in the mold determines the thickness of the 150 raw piece, and the composition of the suspension determines its porosity and microstructure. 151 After casting the green membranes were pre-sintered in air at 950°C for 4 hours and gently 152 rectified and polished. Thermogravimetric analysis (TGA) confirmed that all organics have been burnt out at temperatures above 560°C. The membranes were partially sintered at 1300°C for 153 3 hours dwelling time and heating and cooling rates of 2°C min<sup>-1</sup>. 154

155 Three different membrane families were made. Two thicker membranes, 1.75 mm 156 thickness, MCPS1 and MCPS2 were prepared with pastes CP1 and CP2 respectively and permanence on mold times of 12 min. A thinner membrane, 1.3 mm thick, MCPS3 was 157 158 fabricated with paste CP2 and permanence time of 6 min. The dimensions are specified in 159 Table 2. All the membranes have the same length, about 50 mm, but as the sealing caps fill 160 part of the cylinder inner space the inner membrane volume occupied by liquid was 161 determined measuring the extracted liquid volume. Inner volume is the same for MCPS1 and 162 MCPS3, about 3.5-3.6 mL, and smaller, 2.7 mL in MCPS2 membranes. The external radius of 163 the cylinders is determined by the slip casting mold dimensions and the contraction during 164 sintering. It ranges between 7.1 and 7.35 mm.

165 **Table 2**.

166 Dimensions and porosity values of the ceramic membranes.

	MCDC 1			
	IVICPS 1	IVICPS 2	IVICPS 3	Ref [6]
				Nei. [0]
Length (mm)	50±2	50±2	50±2	50
Density (g/cm <sup>3</sup> )	2.37	2.30	2.38	-
External diameter (mm)	14.6±0.3	14.15±0.35	14.7±0.2	7
Thickness (mm)	1.75±0.2	1.7±0.17	1.15±0.05	1.5
Pore size (nm)	80; 2000	100; 2000	100; 3000	5
Total porosity (%)*	40	42	40	-
Connected porosity (%)	34.5	36.9	39.6	30.5%

167 \* Calculated from comparison between measured ceramic density and dense alumina density

168 - not indicated

169

# 170 2.2. Microstructure characterization

The microstructure of the porous ceramic was studied in polished transverse membrane cross-sections using a JEOL 6301F scanning electron microscope (SEM). The SEM specimens were embedded in epoxy resin and polished in cross-sections according to a polishing procedure previously published [11]. Open and connected porosity was measured by means of an Hg porosimeter (Poremaster, Quantachrome; maximum pressure 30,000 psia). Total porosity was determined by gravimetric density measurement.

177

178 2.3 Diffusion test

179 To test the performance of the cylindrical ceramic membranes and calculate the 180 sampling rates of the device we used a water solution of methylene blue ( $C_{16}H_{18}CIN_3S$ ) as 181 model compound because it is a stable solution, it gives intense color when diffuses into the 182 porous membrane giving information about homogeneity in pore structure, presence of cracks 183 or other structural defects and its concentration can be accurately computed by measuring the 184 optical absorption spectra. The absorption of methylene blue solutions was measured in an 185 HP-Agilent 8453 (Agilent Technologies, CA, USA) spectrophotometer with a diode array 186 detector (DAD). The value of the absorbance at 664 nm, which corresponds to the maximum of 187 one absorption band of this compound, was used as an accurate measure of the 188 concentration. In figure 2 we give the spectroscopic calibration of several solutions of methylene blue giving a molar extinction coefficient of 6.4  $\pm$  0.05 x 10<sup>4</sup> cm<sup>-1</sup>M<sup>-1</sup> for the 189 190 measurement conditions used here.







**Figure 2**. Methylene blue spectroscopic calibration curve\_used in this study. Optical absorption of  $C_{16}H_{18}CIN_3S$ solutions at 664 nm as a function of concentration.

195 Using methylene blue dye and the optical absorption technique we studied the water solution diffusion through the membranes. A beaker with 4 L of mineral water was dosed with 196 methylene blue at 10 mg L<sup>-1</sup>. This concentration was used in the optimization protocol as it 197 198 shows an intense blue color and it can be accurately measured. Several identical cylinders 199 saturated with clean water to create the water membrane were placed in the methylene blue 200 solution. The membranes were closed with thermoplastic rubber caps, placed in a handmade 201 support made of tulle net and then placed inside the methylene blue solution. After each exposure period (2, 5, 6, 9 and 12 d), the solution inside the sampler corresponding to the 202 203 different experiments performed, was collected with a Pasteur pipette and placed in a glass 204 vial. Measuring the concentration of methylene blue inside the sampler as a function of the 205 exposure time permits to establish the flow and diffusion rates of membranes. The external 206 solution from the beaker was also monitored to control the stability of methylene blue

throughout the deployment time. To quantify the concentration of methylene blue inside thecylindrical membranes, we used the calibration values given above.

209

# 210 2.4 Theory and data calculation

211 Analysis of the MCPS up-takings yielded the accumulated mass of solute  $M_s(t)$  diffused 212 through the cylindrical membrane surface over the sampling time t. If during the sampling 213 period the mass flow of contaminants across the water membrane is diffusion limited and 214 diffusion is radial (long cylinder approach) the concentration is only function of radius r and 215 time t. The diffusion equation is [12].

216 
$$\frac{\partial C}{\partial t} = \frac{1}{r} \frac{\partial}{\partial r} \left( r D_e \frac{\partial C}{\partial r} \right)$$
(1)

where  $D_e$  is the effective diffusion coefficient of solute in solvent, measured in cm<sup>2</sup>s<sup>-1</sup> and stands for the diffusion of analytes through the water filled porous ceramic membrane.  $\frac{\partial C}{\partial r}$  is the concentration gradient [13].

220 Consider a cylindrical membrane of inner and outer radii a and b. If the concentration inside 221 cylinder is maintained close to zero, for example if a sorbent is used, and the matrix solute 222 concentration  $C(b)=C_0$  is constant, the steady state mass flux at r=a is proportional to the 223 concentration gradient. The accumulated solute mass  $M_s(t)$  depends linearly on time. It can be 224 given by:

225 
$$M_s(t) = \frac{2\pi L D_e C_0}{\ln(b/a)} \cdot t = k \cdot t$$
 (2)

where *L* is the sampling cylinder length and the slope of gives the uptake rate, k(M/t) that is directly related with the sampling rate,  $R_s(V/t)$ . Linear up taking dynamics are usually observed in PS devices.

229 In the absence of any concentrator the analyte concentration inside the closed cylinder 230 will increase and the concentration gradient will decrease with deployment time. Since the 231 solution of the diffusion equation for a hollow cylinder in the non-steady state is rather 232 complicated we have used that of a solid cylinder instead. The approach could be justified as 233 any mass exchange with the exterior is only made through the membrane wall. Actually, the 234 solution of the diffusion equation for a solid cylinder of infinite length and it has been 235 previously applied, for example, to the study of the reduction kinetics of ceramic composites 236 with cylindrical shape [14,15].

For the solid cylinder, if the diffusion coefficient is constant for initial concentration inside the membrane zero and if L>>b the solution for Eq. (1) is given by Eq. 5.22 in reference [12]

240 
$$\frac{C(a,t)}{C_0} = 1 - \frac{2}{b} \sum_{n=1}^{\infty} \frac{exp(-D_e \alpha_n^2 t) J_0(a\alpha_n)}{\alpha_n J_1(b\alpha_n)}$$
(3)

241 where  $J_i(x)$  are the Bessel functions of the first kind and  $b\alpha_n$  the positive roots of  $J_0(x)$ .  $\alpha_n$  is 242 given in cm<sup>-1</sup> units.

243

252

244 3. Results

# 245 *3.1. Physical characteristics of the membranes*

Three porous ceramic membranes were prepared with two different pore former compositions (see section 2.1 and table 2). SEM studies indicate that the morphology of the pore microstructure is essentially the same in the three cases. Figure 3 shows a typical SEM image of a MCPS2 ceramic membrane transverse cross-section where both cornstarch and graphite pore formers were used. A homogeneous and continuous distribution of spheroid cavities around 10 µm diameter through all the membrane depth is observed.





Figure 4 shows the detailed image of the connecting contact area or necks between cavities. Effect of partial sintering on alumina microstructure can also be seen. Polygonal

- alumina grains indicate that strong struts have been produced but leaving some intergranular
- 257 porosity. This porosity is a consequence of partial sintering and it is related with the alumina
- 258 grain size being 2 to 5 times smaller than alumina grains in agreement with conventional
- sintering theories [16].



260

261 Figure 4. SEM image showing the detail of the cavity and grain structure.

262 Total porosity of the membrane estimated through density measurements is of around 40% (see table 2) in fair agreement with the volume of sacrificial phase (table 1) indicating that 263 264 most of pore volume is produced by pore former phase. However, this relatively high porosity 265 value does not endanger the mechanical stability of the membranes that can be safely handled 266 and sealed. On the other hand, as the diffusion of analytes takes place through the connected 267 pore network and closed pores or blind pores do not contribute to transport of matter it is 268 crucial to know the amount of connected pores. The connected pore volume and pore size has 269 been measured using Hg porosimetry. In these measurements, the intruded mercury volume is 270 obtained as a function of the liquid (mercury) pressure (see figure 5).





Fig. 5. Percentage of intruded volume as a function of pore diameter as measured by Hg porosimetry. Green,
 MCPS1; Blue, MCPS2; Red, MCPS3

274 Due to the capillarity effect the mercury pressure needed to intrude a given cavity is 275 directly related with the size of conduits connecting the pores [17]. However, it has to be taken 276 into account that mercury porosimetry also has some limitations. It provides values of the 277 diameter of the entrance to a pore not of the pore size itself. Also blind holes connected to 278 external surface can be intruded then measured by porosimetry, even if they are not 279 connected to the pore network. Thus, values of connected pores could be overestimated in 280 these measurements, which can be more important for the thinner samplers. Data on total 281 and open porosity obtained as indicated above for the three studied cases are compiled in 282 table 2.

283

# 284 *3.2. Testing and calibration of the membranes*

285 As indicated in section 2.3, the analyte permeation and diffusion rates were studied 286 using methylene blue solution as standard. Two membranes of each configuration (table 2) 287 were placed in the methylene blue solution during 6 days. During exposure period, the 4 L 288 beaker was placed in an orbital shaker (KS500, Janke&Kunkel, IKA WERK, Staufen, Germany) 289 with a constant temperature of 22-23ºC. The orbital shaker has a circular shaking motion at 290 low speed (100 rpm), which does not create vibrations nor produce heat, and it was used to 291 induce a liquid movement simulating the circulating water in a river or wastewater. To 292 determine the water motion effect, we kept 2 more membranes in the same conditions but 293 without motion. In each condition, the concentration of the methylene blue solution inside 294 and outside the cylinder was measured after 6 d by spectrophotometry using the experimental 295 conditions described in section 2.3. The accumulated mass, among the 3 membrane 296 configurations studied, is given in Figure 6.



299

Fig. 6. Total mass of methylene blue accumulated inside the membranes after 6 days of deployment time in a watersolution of 8.85 mg/L concentration.

It is observed that the thinner membranes with the highest connected pore volume (MCPS 3) provided the higher accumulation potential at 6 d deployment time. On the contrary the MCPS1 membranes are less efficient although the geometrical parameters are about the same than those of MCPS2 membranes that shows a better performance owing to their higher concentration of connected pores. Motion has not any meaningful effect in the accumulated mass inside the membrane.

308 To insight the kinetics of mass transport in the membranes the following experiments 309 were performed. Ten MCPS3 cylinders initially filled with clean water were placed inside the 310 methylene blue solution. After 2, 5, 6, 9 and 12 days, two cylinders were removed and the 311 concentration, or accumulated mass of methylene blue inside the cylinder M<sub>s</sub>(t) was 312 determined. The concentration in the external solution was also monitored, being  $C(b)=8.4 \pm$ 0.5 mgL<sup>-1</sup> nearly constant over time. Because no adsorbent was introduced inside the 313 314 membrane, the diffusion experiments would correspond to boundary conditions and transport 315 kinetics described by Eq. 3. In figure 7 we plot C(a,t)/C(b) as a function of the sampling time. 316 The data are well fit by Eq. 3 where up to n=8 terms have to be taken to get a good 317 convergence in the series, specifically for the short deployment times. This result justifies "a posteriori" the approach of a transport model based in that of a solid cylinder instead of a 318 319 hollow cylinder leading to Eq. 3. The best fitting values of De are given in table 3. It is 320 interesting to realize that the effective diffusion values obtained with this more accurate 321 model coincide with those calculated from the sampling rate measurements assuming Eq. 2 is 322 valid which only holds for the short deployment times.



324

**Fig. 7.** Ratio of methylene blue concentration accumulated inside the MCPS3 membranes as a function of deployment time. Blue points experimental values, green points calculated values obtained from Eq.3 with effective diffusion coefficient of  $D_e=3.0 \text{ E-8 cm}^2$ /s. Dark line is a best-fit line to theoretical values.

328

# 329 4. Discussion

### 330 4.1 Pore structure

331 The role of the ceramic membrane in the MCPS device is complex. On one side, the 332 porous ceramic holds the water membrane that is actually the diffusion barrier for 333 contaminants transport from the external solution to be monitored to the concentrator 334 confined by the membrane. The ceramic membrane is also the physical container that holds 335 and protects the sorbent material that accumulates the analyte. Consequently, the ceramic 336 microstructure has to be designed to support the largest water volume without weakening the 337 mechanical integrity of the membrane, so that organic contaminants can diffuse across the 338 membrane with minimal interaction with the ceramic material whereas the inner sorbent 339 (diameter >30  $\mu$ m) does not escape from the cylinder. The ceramic membrane has to be also a 340 barrier for microorganisms and mud entering the inner chamber. In our design, porosity, 341 inertness, pore size and diffusion thickness, which are the key parameters defining the ceramic membrane, were amended. The microstructure of the macro-porous membrane was designed 342 343 to get a porous ceramic shell with a hierarchical pore structure consisting of relatively large 344 spheroid cavities of about 10 µm diameter connected by smaller pores.

Following current knowledge on macro-porous ceramic processing [18, 9] we used starch as sacrificial template to create the large cavities. Starch pore forming is a low-cost and simple technique appropriated synthesis of macro-porous ceramics with pores of less than 20

348 μm size, difficult to achieve with other pore forming techniques such as polymer replica, wood 349 replica or direct foaming [19,16]. In particular cornstarch powder was dispersed in the alumina 350 suspension to further processing by wet colloidal route using slip casting. We have also 351 explored graphite as pore former. The procedure we used to incorporate graphite to the 352 alumina suspension is similar to that used with starch. In our case the effect of graphite former 353 simply adds to that of starch. The organics including pore former were smoothly removed from 354 the green body by pyrolysis treatment. Any cracking effect in the ceramic was observed. In 355 SEM images the spheroidal cavities resulting from pore formers are clearly seen (figs. 3 and 4). 356 Also Hg-porosimetry results (fig. 5) show the presence of a large amount of pores with sizes 357 between 200 to 2000 nm that could be associated to the entrances to spheroidal cavities as 358 shown in fig. 4. It is interesting to realize that addition of graphite as pore former produces an 359 increase in the size of these pores. Porosimetry measurements show the presence of a small 360 amount of large pores about 5 to 10  $\mu$ m diameter more evident in samples MCPS1 and MCPS3 361 in fig. 5 that could be associated either to cavities with large entrances or to cavities at the 362 membrane surface. To increase connectivity between cavities small inter-cavity channels were 363 produced using partial sintering conditions. These small pores can be seen in the SEM images 364 at the grain boundaries of alumina grains (see fig. 4). According to porosimetry results (fig. 5) 365 there contribution to the total pore volume is modest, less than 10% volume, but increased 366 pore connectivity. The size of these small pores is between 70 to 200 nm.

367 Finally, it is interesting to point out that using these suspension compositions and 368 forming procedures there is not any significant dimensional change during drying and 369 demoulding process. The size of spherical pores corresponds to that of starch pore formers 370 indicating that there is not significant swelling or water uptake by starch particles during the 371 process. In fact both starch and graphite are rather insoluble in water at ambient 372 temperatures. The open (connected) porosity values of these membranes are quite large  $\varepsilon$  = 373 0.345, 0369 and 0.396 for MCPS1, MCPS2 and MCPS3 respectively. In fact, in the case of the 374 thinnest MCPS3 membrane essentially all the pores are accessible to Hg infiltration.

375 The parameters describing the pore structure of the MCPS used in this study are 376 compared in Table 2 with those of state of art sampler [6]. Main difference between both 377 systems arises in the higher porosity values and much larger pore size in the MCPS membranes 378 studied in this work. As a consequence of this particular microstructure the present MCPS 379 membranes can be easily saturated with large water volumes and the interaction between 380 analyte and pore walls is expected to be minimum. The ceramic membrane thickness is also much larger than the diffusion barrier length of the sampled chemicals that has been 381 382 estimated by Gale of the order of 100 to 400 nm [20]. In addition, the size of pores is small enough to avoid microorganisms entering the dosimeter and any net flow of water. As a consequence we expect that the mechanism governing the contaminants transport through the membrane is in our case a true diffusion rate-limiting process and data can be analyzed using ad hoc diffusion theory.

387

# 388 4.2 Diffusion of analytes.

389 Using the data given in fig. 6 the uptake rates for sampling times of 6 days were calculated 390 assuming a linear dependence with deployment time as predicted by Eq. 2. Taking into 391 account the concentration of external solution we calculated the sampling rates for methylene 392 blue that are given in table 3. As expected the sampling rates are modest, as any concentrator 393 is used in this experiment, but clearly show the increases through the MCPS1, MCPS2 and 394 MCPS3 sequence. At a first sight and according to porosity values in table 2 it could be 395 concluded that sampling rates increases with membrane porosity. However, sampling rates 396 also depend on membrane geometrical factors such as thickness and inner volume.

397

**Table 3.** MCPS membrane characterization parameters:  $D_e$ , effective diffusion coefficient;  $D_w$ , diffusion coefficient in water (theoretical estimation); *F*, membrane geometrical factor; *m*, Archie's exponent value;  $R_s$ , sampling rate; and uptake rate; *k*.

	k	R <sub>s</sub>	<i>D<sub>e</sub></i> (Eq. 2)	<i>D<sub>e</sub></i> (Eq. 3)	F	т
	(µg day <sup>-1</sup> )	(mL day <sup>-1</sup> )	(cm <sup>2</sup> s <sup>-1</sup> )	(cm <sup>2</sup> s <sup>-1</sup> )		
MCPS1	1.6	0.18	2.3±0.5E-8		1±0.5E-2	4.5±0.5
MCPS2	2.05	0.23	2.9±0.5E-8		0.75±0.1E-2	4.9±0.1
MCPS3	2.4	0.27	2.7±0.1E-8	3.0±0.5E-8	0.6±0.1E-2	5.5±0.1

<sup>401</sup> 

The dependence of sampling rates in the membrane geometrical factors can be eliminated if the effective diffusion parameter through the membrane is calculated. In fact, following previous studies of solute diffusion through porous media [21] it is evident that the effective diffusion parameter  $D_e$  is related with the actual diffusion coefficient in water  $D_w$  by the membrane geometrical factor *F*.

$$407 D_e = D_w.F (4)$$

According to Delgado et al., [22]

408

409

 $F = \varepsilon . \delta / \tau$ 

(5)

410  $\tau$  (>1) is the tortuosity factor, a geometrical factor related with the pore geometry,  $\delta$  ( $\leq$ 1) is 411 the constriction factor assumed to be unity in the present case as the predominant large 412 cavities and wide channels makes collision between molecules much more probable than with 413 the pore walls and  $\mathcal{E}$  stands for the value of effective or connected porosity. Alternatively, the 414 most extensively used relationship to obtain diffusion coefficients of contaminants in water in 415 relation with geometrical factors of the membrane is the Archie's law [23], typically used by 416 geologists to link the electrical resistivity of a rock to its porosity and to the resistivity of the 417 water that saturates its pores.

418  $F = \varepsilon^m$ 

(6)

Where m is known as the cementation exponent. Both eq. 5 and 6 are equivalent in order to
account for the membrane geometrical factor that relates the pore structure with the
sampling rates of the MCPS.

422 The calculation of the effective diffusion coefficient  $D_e$  can be done either using the 423 simplest linear time dependence predicted by Eq. 2 or the most accurate model of Eq. 3. It is 424 interesting to notice that Eq. 3 also predicts a linear dependence with time for short 425 deployment times. Using Eq. 2, the geometrical dimensions in table 2, the effective membrane 426 length estimated from the measure of the spiked liquid inside membrane and inner radius and 427 the calculated sampling rates we get a value of the effective diffusion coefficient of methylene 428 blue in those membranes given in table 3. Errors involved in determination of  $D_e$  are large 429 owing to the uncertainties in the measure of the accumulation rates. Comparison of MCPS 1 430 and 2 membranes seems to indicate an increase on diffusion factor in the later, which could be 431 associated with the higher porosity produced by graphite pore formers. However this 432 implementation with increasing porosity has not been seen in the MCPS3 membranes. It might 433 be possible that a part of the measured porosity in the Hg-porosimetry experiments would 434 correspond to near to surface pores that would be isolated from the pore network. In fact, the 435 porosity curves for MCPS2 and MCPS3 in fig. 5 are very similar except for some amount of 436 large cavities that are detected in the MCPS3 membranes and that as it has been pointed out 437 before could correspond to cavities near surface with large entrance pores. We have to realize 438 that MCPS3 membranes are the thinnest ones so with larger surface to volume ratios.

In order to calculate the membrane geometrical factor, F, we can apply eq. 4. We have not found values of diffusion coefficient of methylene blue in water in literature. Consequently, we used the estimated diffusion coefficient in water at room temperature resulting from equation  $D_w=3.59E-07 T(K)/M^{0.531}$  that gives a theoretical estimation of the diffusion coefficient of methylene blue in water  $D_w=5.1E-06 \text{ cm}^2\text{s}^{-1}$ . Then, using eqs. 4 and 6 we can obtain the membrane geometrical factors an Archie's exponent values given in Table 3.

In spite of the quite large connected porosity values the membrane geometrical factors are low and the Archie's coefficients high. That results in high values of the tortuosity factor in these ceramics. Unexpectedly, tortuosity seems to increase with porosity. One possible explanation could be related with the bimodal pore morphology in these ceramics. In fact it has been reported that a distribution of pores with varying cross sections may cause
abnormally high tortuosity factors [24] although, to our best knowledge, this important aspect
has not been studied in detail up to now.

However, the pore microstructure developed in these ceramic membranes gives 452 453 enough uptake capacity and stability for being used in the fabrication of CPS devices for the 454 analysis of organic contaminants in river waters or wastewater. Recently, it has been reported 455 that CPS fabricated with the "a priory" less optimized MCPS1 membranes, were successfully used for the monitoring of cytostatic drugs in water [25]. These devices achieved sampling 456 rates between 0.825 and 3.350 mL day<sup>-1</sup> that allowed for the first time the detection of 457 458 cyclophosphamide and mycophenolic acid at concentrations of  $19 \pm 3$  and  $136 \pm 28$  ng L<sup>-1</sup> in effluent waters of a Waste Water Treatment Plant (WWTP) in Barcelona. 459

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### 461 **5. Conclusions**

462 Macroporous alumina membranes have been developed. Using pore formers and 463 partial sintering techniques a hierarchical pore morphology consisting of large cavities 5-10 μm 464 diameter and small pores of 70-200 nm provides high pore volume and good connectivity 465 between pores. The diffusion properties and sampling rates of the ceramic membranes were 466 determined using methylene blue as test compound as a function of the connected porosity 467 values, pore former composition and ceramic thickness. Ceramics with greater values of 468 connected porosity give better diffusivities. Thinner membranes give greater sampling ratio 469 values. This pore structure has been proved appropriated for development of passive samplers 470 for detection of low-concentration emergent organic contaminants in water.

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