



Arctium Species Secondary Metabolites Chemodiversity and Bioactivities

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Arctium species are known for a variety of pharmacological effects due to their diverse volatile and non-volatile secondary metabolites. Representatives of *Arctium* species contain non-volatile compounds including lignans, fatty acids, acetylenic compounds, phytosterols, polysaccharides, caffeoylquinic acid derivatives, flavonoids, terpenes/terpenoids and volatile compounds such as hydrocarbons, aldehydes, methoxypyrazines, carboxylic and fatty acids, monoterpenes and sesquiterpenes. *Arctium* species also possess bioactive properties such as anti-cancer, anti-diabetic, anti-oxidant, hepatoprotective, gastroprotective, antibacterial, antiviral, antimicrobial, anti-allergic, and anti-inflammatory effects. This review aims to provide a complete overview of the chemistry and biological activities of the secondary metabolites found in therapeutically used *Arctium* species. Summary of pharmacopeias and monographs contents indicating the relevant phytochemicals and therapeutic effects are also discussed, along with possible safety considerations.

Keywords: *Arctium* species, secondary metabolites, volatile compounds, non-volatile compounds, chemodiversity, bioactivity

INTRODUCTION

Botanical and Ethnobotanical Aspects

The genus *Arctium* L. (Asteraceae/Compositae, tribe Cardueae, subtribe Carduinae), together with the related genera *Cousinia* Cass., *Hypacanthium* Juz. and *Schmalhausenia* C. Winkl, forms the so-called *Arctium*–*Cousinia* group (de Souza et al., 2004). The species of the *Arctium* genus, also known as ‘burdock,’ comprise biennial herbs occurring in waste places, streams and roadsides, less

often in wood and forests, in temperate regions of Europe and Asia and sporadically in subtropical and tropical regions (European Scientific Cooperative on Phytotherapy, 2003). In North and South America, the genus is considered as naturalized, whereas in Africa it is quite rare. The name of the genus comes from the Greek ‘*arctēion*’ which means ‘bear,’ alluding to the plant habitus characterized by pronounced hairiness.

According to the Plant List¹, this genus encompasses 18 recognized species among which five are considered as hybrid species due to the frequent outbreeding occurring between its allogamous representatives (Lopez-Vinyallonga et al., 2010).

Arctium species are represented by hemicryptophyte plants equipped with a stout, erect taproot and entire (sporadically as dentate), rough, unarmed, alternate, tomentose, and cordate leaves. The stem is usually stout, erect, grooved, branched, and reddish. Inflorescences are formed by solitary or corymbose ovoid-conical to spherical capitula equipped with involucre made up of bracts ending with hooked apices. Receptacles are composed of numerous, hard scales. Florets are only tubulose, hermaphrodite, purple or white. Pollination is allowed by insects, mostly belonging to Lepidoptera. Fruits are having oblong, rugose achenes equipped with a golden-yellow pappus (European Scientific Cooperative on Phytotherapy, 2003).

The *Arctium* genus is highly polymorphic due to variability occurring in hairiness of leaves and capitula, length of floral peduncles, and color of capitula and florets. As a consequence, a sharp distinction between its members cannot occasionally be defined. In the Euro-Mediterranean area, six main species are found: *A. atlanticum* (Pomel) H. Lindb., *A. lappa* L., *A. minus* (Hill) Bernh., *A. nemorosum* Lej., *A. palladini* (Marcow) R.E.Fr. & Soderb. and *Arctium tomentosum* Mill. (European Scientific Cooperative on Phytotherapy, 2003; **Figure 1**).

Arctium lappa enjoys a longstanding use in the traditional medicine (mainly roots, and, to a lesser extent, leaves and seeds) due to the bioactive properties of its metabolites (Zhao et al., 2014). To the best of our knowledge, the most utilized species for therapeutic purposes, are on first place *A. lappa*, also known as ‘greater burdock,’ and, to a minor extent, *A. minus* (lesser burdock) and *A. tomentosum* (wooly burdock).

Arctium lappa is an herbaceous biennial plant up to 150 cm tall, with pubescent to subglabrous epigeal parts. Basal leaves are up to 50 cm in diameter, ovate, cordate and with hollow petioles. Stems are branched and end with corymbose capitula. Florets are as long as the involucre bracts. *A. minus* differs for the shape of inflorescence (solitary terminal capitula), dimensions of involucre bracts and capitula (smaller and shorter, respectively) and consistency of petiole (hollow). Furthermore, the bracts show a relatively hinted hairiness. *A. tomentosum* is characterized by petioles and peduncles covered with wooly tufts; petioles are solid. The involucre bracts are similar in dimensions to those of *A. minus*, but they show a dense covering of hairs. Florets are longer than bracts as in *A. minus* (European Scientific Cooperative on Phytotherapy, 2003). These three species are quite common in central Europe where often they undergo interspecific

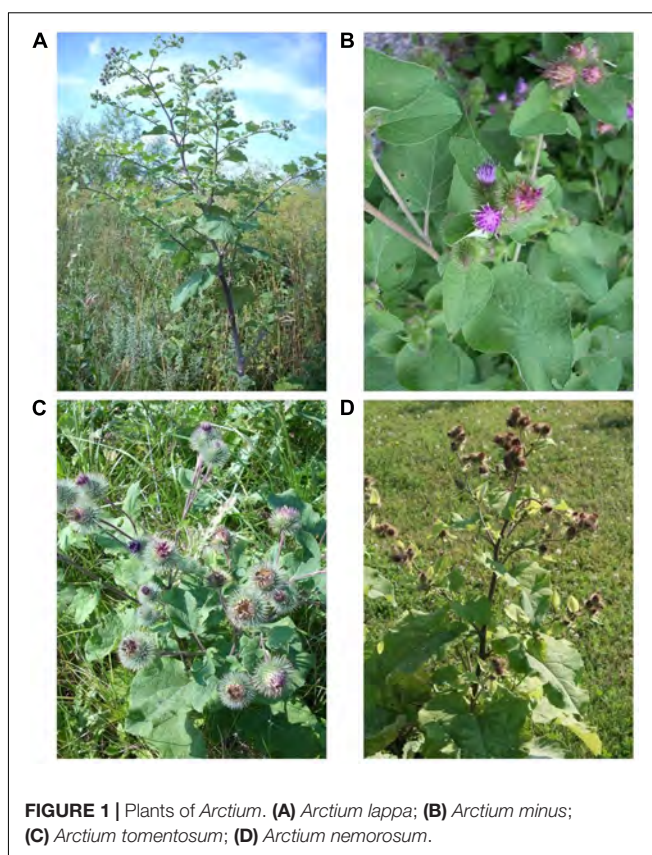


FIGURE 1 | Plants of *Arctium*. **(A)** *Arctium lappa*; **(B)** *Arctium minus*; **(C)** *Arctium tomentosum*; **(D)** *Arctium nemorosum*.

hybridization giving rise to questions about their integrity (European Scientific Cooperative on Phytotherapy, 2003).

Greater burdock (*A. lappa*) has been traditionally used in both Asian and European medicines as depurative, diuretic, carminative, anti-inflammatory, and anti-tubercular agent (Zhao et al., 2014). For therapeutic purposes, its different parts such as roots, fruits, and leaves are used. The latter have been used to treat ulcers and fester wounds (Jaric et al., 2007). They are also applied externally on the forehead to cure headache and fever, on the scalp to treat bruises and hair loss, mixed with oil and honey and applied on the chest to heal cough. In addition, under infusion, they are taken orally to treat enuresis in children (Pieroni et al., 2011). Fruits of burdock (*Arctii fructus*) are used to purify the blood (Lans and Turner, 2011) and to treat respiratory and infectious diseases (Bai et al., 2016). In addition, *A. lappa* roots, together with aerial parts of *Rumex acetosella* L., leaves of *Ulmus rubra* Muhl. and rhizomes of *Rheum officinale* Baill., are used to make ‘Essiac,’ a tea used by the Ojibwa tribe of Canada for the treatment of cancer (Leonard et al., 2006). In the veterinary medicine, the root is used to treat mastitis (Lans et al., 2007), whereas the whole plant is applied against endoparasites in poultry (Lans and Turner, 2011). Besides therapeutic uses, *A. lappa* is also appreciated as an edible plant. For the latter purpose, young leaves, and stalks are eaten raw or cooked (Pieroni et al., 2011).

Lesser burdock (*A. minus*) leaves are traditionally used externally to treat rheumatic pains, fever, sunstroke, wounds,

¹<http://www.theplantlist.org>

general infections, skin and body inflammations, alopecia, and bladder diseases (de Souza et al., 2004; Erdemoglu et al., 2009; Neves et al., 2009). They are also disposed above the body of the patient, wetted with vinegar or milk, to stimulate sweating (Sezik et al., 2001). Roots and leaves, under infusion, are also used against snake and scorpion bites and to purify the blood (Mosaddegh et al., 2012). Basal leaves and stems are also eaten raw as a snack or stewed (Tardio et al., 2005). Due to their bitter taste, they are also used to stimulate the appetite and liver functions (Tardio et al., 2005).

Wooly burdock (*A. tomentosum*) leaves are used as vulnerary, to treat skin rash, ulcers, abscesses, mouth sores and against rheumatic pains, whereas root is applied against alopecia and to wash hairs (Sezik et al., 2004; Saric-Kundalic et al., 2010). Roots are also employed to make a tea used for digestive problems, ulcers, rheumatisms, to purify the blood and increase sweating and as diuretic (Saric-Kundalic et al., 2010).

Medicinal Uses of *Arctium* Species in Pharmacopeias and Monographs

Burdock species and in particular *A. lappa* are used in traditional medicine for different purposes. The main traditional use of the roots of *A. lappa* in Europe comprises treatment of dermatological disorders (Saric-Kundalic et al., 2010; Miglani and Manchanda, 2014) whereas in other Eastern and Asian countries *A. lappa* fruits and roots are used as an antidiabetic remedy (Tousch et al., 2014; Xu et al., 2014, 2015; Ahangarpour et al., 2017). In Traditional Chinese Medicine (TCM), apart from the antidiabetic activity, the roots of *A. lappa* are considered as a blood detoxifying agent (Qin et al., 2014). In the Japanese pharmacopeia, the fruit is included as a traditional herbal medicine with recent studies revealing the potential of its extracts in oncology (Ikeda et al., 2016). The leaves of *A. lappa* have also been reported as an anti-inflammatory agent to relieve gastrointestinal disorders in Brazilian traditional medicine (de Almeida et al., 2013).

According to international institutions that work in the validation of traditional herbal medicines, such as the European Medicines Agency (EMA) and the European Scientific Cooperative on Phytotherapy (ESCOP), *A. lappa* is recommended and approved for different indications. For example, the EMA monograph approves the use of roots of *A. lappa*, *A. minus*, and *A. tomentosum* as an adjuvant in minor urinary tract complaints, in temporary loss of appetite and for seborrheic skin conditions (European Medicines Agency, 2011). All these indications are based upon long-standing use. In 2016, ESCOP released a monograph where the roots of all the former three species are indicated to be internally and externally used for seborrheic skin, eczema, furuncles, acne, psoriasis and internally for minor urinary tract disorders (European Scientific Cooperative on Phytotherapy-The Scientific Foundation for Herbal Medicinal Products, 2016). For oral and internal administration, the herbal drug can be used as an infusion, extract, tincture or decoction but the fresh pulp of the roots or a decoction can also be directly applied to the skin. The later monograph reveals that *A. lappa* preparations

should not be ingested during pregnancy, lactation, or in case of hypersensitivity to the Compositae and in patients with oedema due to impaired heart or kidney function. Although certain preclinical studies can be found in the literature, clinical trials are not available for these indications approved by ESCOP and EMA.

PHYTOCHEMISTRY

Non-volatile Compounds

Till date, more than two hundred non-volatile compounds have been isolated from *Arctium* genus. These chemical compounds include lignans, terpenoids, sterols, flavonoids, phenolics, lactones, polyacetylenes, quinic acids, and sugars (polysaccharides). In particular, lignans are the most characteristic components in the *Arctium* genus. The details of chemical compounds, their occurrence in different plant parts, and the analytical methods used for their qualitative determinations are briefly summarized in Table 1 whereas their description is provided in this section. The chemical structures of some compounds from *Arctium* species are shown in Figure 2.

Lignans

Major biologically active lignans include mainly arctigenin (a dietary phytoestrogen) and its glycoside, arctiin (lignanoides) occurring commonly in seeds, roots, fruits, and leaves of *A. lappa* and *A. tomentosum* (Yu et al., 2003; Ming et al., 2004; Liu et al., 2005, 2012, 2015; Wang et al., 2005; Matsumoto et al., 2006; Gao et al., 2008; Boldizar et al., 2010; Ferracane et al., 2010; Zhou et al., 2011; Qin et al., 2014; Su et al., 2015; Lou et al., 2016). In addition, seeds and roots are distributed with low levels of dilignans and sesquilignans. For the first time, two new sesquilignans, namely lappaol A and B were isolated and characterized from *A. lappa* seeds (Ichihara et al., 1976). Later, 3 more sesquilignans, namely, lappaol C, D, and E, and two dilignans, namely lappaol F and H, were structurally determined from the seeds of *A. lappa* (Ichihara et al., 1977, 1978; Yong et al., 2007; Su et al., 2015). Lappaol A, C, and F are also found in the fruits of *A. tomentosum* (Kardosova et al., 2003). Two new lignans, neoarctin A and B, along with other recognized compounds including arctiin, arctigenin, daucosterol, lappaol F, isolappaol C and matairesinol were identified in seeds of *A. lappa* (Wang and Yang, 1995; Kardosova et al., 2003; Yong et al., 2007; Gao et al., 2008; Qin et al., 2014; Su et al., 2015). A simple RP-HPLC method was developed to identify the presence of arctiin in fruits of *A. lappa* (Yu et al., 2003; Boldizar et al., 2010). Using bioactivity-guided fractionation, lappaol A, C and F, arctiin and arctigenin E were isolated and characterized from the ethanolic extract (95%) of *A. lappa* seeds (Ming et al., 2004). Likewise, HPLC/UPLC/LC/MS/MS methods have been developed to identify arctigenin and arctiin in the seeds, leaves and roots of *A. lappa* (Yu et al., 2003; Liu et al., 2005; Ferracane et al., 2010; Lou et al., 2010a,b; Predes et al., 2011). Further, a supercritical fluid extraction procedure was found to be superior for extracting arctiin from *A. lappa*

TABLE 1 | The known non-volatile constituents of *Arctium* species.

| No | Compound name | Formula | Species | Plant origin/part | Analytical method | References |
|----|---|---|--|------------------------------|---|---|
| 1 | Lignans Diarctigenin | C ₄₂ H ₄₆ O ₁₂ | <i>A. lappa</i> | Fruits, roots, seeds | IR/NMR/MS/TLC | Han et al., 1994; Park et al., 2007; Qin et al., 2014 |
| 2 | Arctiin | C ₂₇ H ₃₄ O ₁₁ | <i>A. lappa</i> , <i>A. tomentosum</i> | Leaves, fruits, roots, seeds | UV/IR/MS/NMR/HPLC/LCMS/MALDI-QIT-TOF MS | Wang and Yang, 1993; Ting-Guo et al., 2001; Yu et al., 2003; Ming et al., 2004; Liu et al., 2005, 2012, 2015; Wang et al., 2005; Matsumoto et al., 2006; Boldizar et al., 2010; Ferracane et al., 2010; Zhou et al., 2011; Qin et al., 2014; Su et al., 2015; Lou et al., 2016; Al-Shammaa et al., 2017 |
| 3 | Arctigenin | C ₁₂ H ₂₄ O ₇ | <i>A. lappa</i> , <i>A. tomentosum</i> | Leaves, fruits, seeds, roots | UV/MS/NMR/HPLC/LCMS/MALDI-QIT-TOF MS/HRESI-MS | Umehara et al., 1993; Wang and Yang, 1993; Liu et al., 2005, 2012, 2015; Matsumoto et al., 2006; Gao et al., 2008; Boldizar et al., 2010; Ferracane et al., 2010; Predes et al., 2011; Zhou et al., 2011; Qin et al., 2014; Su et al., 2015; Al-Shammaa et al., 2017 |
| 4 | Arctigenin-4-O-β-D-gentiobioside | C ₁₈ H ₃₂ O ₁₆ | <i>A. lappa</i> | Fruits | NMR/UV/IR/ORD/HRESIMS | Yang et al., 2015 |
| 5 | Arctigenin-4-O-α-D-galactopyranosyl-(1→6)-O-β-D-glucopyranoside | C ₁₈ H ₃₂ O ₁₆ | <i>A. lappa</i> | Fruits | NMR/UV/IR/ORD/HRESIMS | Yang et al., 2015 |
| 6 | Arctigenin-4-O-β-D-apiofuranosyl-(1→6)-O-β-D-glucopyranoside | C ₃₂ H ₄₂ O ₁₅ | <i>A. lappa</i> | Fruits | NMR/UV/IR/ORD/HRESIMS | Yang et al., 2015 |
| 7 | 3-benzyl-6-(1-hydroxyethyl)-2,5-piperazinedione | C ₁₃ H ₁₆ N ₂ O ₃ | <i>A. lappa</i> | Fruits | IR/HR-ESI-MS/NMR/CD | Yang et al., 2012 |
| 8 | 3-benzyl-2,5- piperazinedione | C ₁₃ H ₁₆ N ₂ O ₂ | <i>A. lappa</i> | Fruits | IR/HR-ESI-MS/NMR/CD | Yang et al., 2012 |
| 9 | 5'-propanediolmatairesinoside | C ₂₉ H ₃₈ O ₁₃ | <i>A. lappa</i> | Fruits | NMR/UV/IR/ORD/HRESIMS | Yang et al., 2015 |
| 10 | (7'R,8R,8'R)-rafanotrachelogenin-4-O-β-D-glucopyranoside | C ₂₇ H ₃₄ O ₁₂ | <i>A. lappa</i> | Fruits | NMR/UV/IR/ORD/HRESIMS | Yang et al., 2015 |
| 11 | (7'S,8R,8'R)-rafanotrachelogenin-4-O-β-D-glucopyranoside | C ₂₇ H ₃₄ O ₁₂ | <i>A. lappa</i> | Fruits | NMR/UV/IR/ORD/HRESIMS | Yang et al., 2015 |
| 12 | (7S,8S,8'R)-4,7-dihydroxy-3,3',4-trimethoxyl-9-oxo benzylbutyrolactone lignan-4-O-β-D-glucopyranoside | C ₂₇ H ₃₄ O ₁₂ | <i>A. lappa</i> | Fruits | NMR/UV/IR/ORD/HRESIMS | Yang et al., 2015 |

(Continued)

TABLE 1 | Continued

| No | Compound name | Formula | Species | Plant origin/part | Analytical method | References |
|----|---|---|--|-------------------|--|---|
| 13 | (7S,8S,8'R)-4,7- dihydroxy-3,3',4'-trimethoxyl-9-oxo dibenzylbutyrolactone lignin | C ₂₁ H ₂₄ O ₇ | <i>A. lappa</i> | Fruits | NMR/UV/IR/ORD/HRESIMS | Yang et al., 2015 |
| 14 | (7R,8S,8'R)-4,7,4'-trihydroxy-3,3'- dimethoxyl-9-oxo dibenzylbutyrolactone lignan-4-O-β-D-glucopyranoside | C ₂₆ H ₃₂ O ₁₂ | <i>A. lappa</i> | Fruits | NMR/UV/IR/ORD/HRESIMS | Yang et al., 2015 |
| 15 | 7,8-didehydroarctigenin | C ₂₁ H ₂₂ O ₅ | <i>A. lappa</i> | Fruits | HRFAB/EIMS/NMR | Matsumoto et al., 2006 |
| 16 | Arctiidilactone | C ₂₀ H ₂₀ O ₈ | <i>A. lappa</i> | Fruits | NMR/UV/IR/ORD/HRESIMS | Yang et al., 2015 |
| 17 | Arctiapolignan A | C ₂₀ H ₂₈ O ₁₀ | <i>A. lappa</i> | Fruits | NMR/UV/IR/ORD/HRESIMS | Yang et al., 2015 |
| 18 | Arctiisiquinolignan A | C ₄₂ H ₅₂ O ₁₉ | <i>A. lappa</i> | Fruits | NMR/UV/IR/ORD/HRESIMS | Yang et al., 2015 |
| 19 | Arctiisiquinolignan B | C ₃₆ H ₄₆ O ₁₆ | <i>A. lappa</i> | Fruits | UV/IR/HRESIMS/NMR | He et al., 2016 |
| 20 | Arctiiphenolglycoside A | C ₁₉ H ₂₈ O ₁₃ | <i>A. lappa</i> | Fruits | UV/IR/HRESIMS/NMR | He et al., 2016 |
| 21 | Arctignan A | C ₃₀ H ₃₄ O ₁₀ | <i>A. lappa</i> | Seeds | UV/MS/NMR/HPLC | Umehara et al., 1993 |
| 22 | Arctignan B | C ₃₀ H ₃₄ O ₁₀ | <i>A. lappa</i> | Seeds | UV/MS/NMR/HPLC | Umehara et al., 1993 |
| 23 | Arctignan C | C ₃₀ H ₃₂ O ₁₀ | <i>A. lappa</i> | Seeds | UV/MS/NMR/HPLC | Umehara et al., 1993 |
| 24 | Arctignan D | C ₃₀ H ₃₄ O ₁₀ | <i>A. lappa</i> | Seeds | UV/MS/NMR/HPLC/LCMS/MALDI-QIT-TOF MS | Umehara et al., 1993; Liu et al., 2012 |
| 25 | Arctignan E | C ₄₀ H ₄₄ O ₁₃ | <i>A. lappa</i> | Seeds | UV/IR/MS/NMR/HPLC | Umehara et al., 1993; Ming et al., 2004; Ferracane et al., 2010; Qin et al., 2014 |
| 26 | Lappaol A | C ₃₀ H ₃₂ O ₉ | <i>A. lappa</i> , <i>A. tomentosum</i> | Seeds/fruits | TLC/UV/IR/MS/NMR/HPLC | Ichihara et al., 1976; Ting-Guo et al., 2001; Ming et al., 2004; Ferracane et al., 2010; Liu et al., 2012; Qin et al., 2014; Su et al., 2015 |
| 27 | Lappaol B | C ₃₁ H ₃₄ O ₉ | <i>A. lappa</i> | Seeds/fruits | NMR/MS/TLC/HPLC | Ichihara et al., 1976; Qin et al., 2014 |
| 28 | Isolappaol C | C ₃₀ H ₃₄ O ₁₀ | <i>A. lappa</i> , <i>A. tomentosum</i> | Seeds/fruits | NMR/MS/TLC | Park et al., 2007; Qin et al., 2014; Su et al., 2015 |
| 29 | Lappaol C | C ₃₀ H ₃₄ O ₁₀ | <i>A. lappa</i> | Seeds | TLC/UV/IR/MS/NMR | Ichihara et al., 1977; Ting-Guo et al., 2001; Ming et al., 2004; Park et al., 2007; Ferracane et al., 2010; Liu et al., 2012; Su et al., 2015 |
| 30 | Lappaol D | C ₃₁ H ₃₆ O ₁₀ | <i>A. lappa</i> | Seeds | NMR/MS/TLC | Ichihara et al., 1977; Park et al., 2007 |
| 31 | Lappaol E | C ₃₀ H ₃₄ O ₁₀ | <i>A. lappa</i> | Seeds | NMR/MS/TLC | Ichihara et al., 1977; Park et al., 2007 |
| 32 | Lappaol F | C ₄₂ H ₄₆ O ₁₂ | <i>A. lappa</i> , <i>A. tomentosum</i> | Fruits, seeds | TLC/UV/IR/MS/NMR/HPLC | Ting-Guo et al., 2001; Ming et al., 2004; Park et al., 2007; Ferracane et al., 2010; Qin et al., 2014 |
| 33 | Lappaol H | C ₄₀ H ₄₆ O ₁₄ | <i>A. lappa</i> | Seeds/fruits | UV/MS/NMR/HPLC/LCMS/MALDI-QIT-TOF MS | Liu et al., 2012; Qin et al., 2014 |
| 34 | Neoarctin A | C ₄₂ H ₄₆ O ₁₂ | <i>A. lappa</i> | Seeds | UV, IR, 1H-NMR, 13C-NMR, DEPT, 2D-NMR and MS | Wang and Yang, 1995; Yong et al., 2007 |

(Continued)

TABLE 1 | Continued

| No | Compound name | Formula | Species | Plant origin/part | Analytical method | References |
|----|---|---|-----------------|-------------------|--|---|
| 35 | Neoarctiin B | C ₄₂ H ₄₆ O ₁₂ | <i>A. lappa</i> | Seeds | UV, IR, 1H-NMR, 13C-NMR, DEPT, 2D-NMR and MS | Wang and Yang, 1993 |
| 36 | Matairesinoside | C ₂₆ H ₃₂ O ₁₁ | <i>A. lappa</i> | Fruits | UV/IR/HPLC | Boldizar et al., 2010 |
| 37 | Matairesinol | C ₂₀ H ₂₂ O ₆ | <i>A. lappa</i> | Seeds/fruits | UV/MS/NMR/HPLC/LCMS/MALDI-QIT-TOF MS | Wang and Yang, 1993; Boldizar et al., 2010; Ferracane et al., 2010; Liu et al., 2012; Qin et al., 2014; Su et al., 2015 |
| 38 | Matairesinol-4,4'-di-O-β-D-glucopyranoside | C ₂₇ H ₃₄ O ₁₂ | <i>A. lappa</i> | Fruits | NMR/UV/IR/ORD/HRESIMS | Yang et al., 2015 |
| 39 | Pinoresinol | C ₂₀ H ₂₂ O ₆ | <i>A. lappa</i> | Fruits | UV/IR/HPLC | Boldizar et al., 2010 |
| 40 | Phylligenin | C ₂₁ H ₂₄ O ₆ | <i>A. lappa</i> | Fruits | UV/IR/HPLC | Boldizar et al., 2010 |
| 41 | Styraxlignolide E | C ₂₆ H ₃₂ O ₁₁ | <i>A. lappa</i> | Fruits | NMR/UV/IR/ORD/HRESIMS | Yang et al., 2015 |
| 42 | Styraxlignolide D | C ₂₆ H ₃₂ O ₁₁ | <i>A. lappa</i> | Fruits | NMR/UV/IR/ORD/HRESIMS | Yang et al., 2015 |
| 43 | Syringaresinol | C ₂₂ H ₂₆ O ₈ | <i>A. lappa</i> | Roots | UV/IR/ESIMS/NMR | Han et al., 2013 |
| 44 | (7S, 8R)-4,7,9,9'-tetrahydroxy-3,3'-dimethoxy-7'-oxo-8-4'-oxyneolignan-4-O-β-D-glucopyranoside | C ₂₆ H ₃₄ O ₁₃ | <i>A. lappa</i> | Roots | IR/HR-ESI-MS/NMR/CD | Yang et al., 2012 |
| 45 | (7'S, 8'R, 8S)-4,4',9'-trihydroxy-3,3'-dimethoxy-7',9-epoxylignan-7-oxo-4-O-β-D-glucopyranosyl-4'-O-β-D-glucopyranoside | C ₃₂ H ₄₂ O ₁₇ | <i>A. lappa</i> | Roots | IR/HR-ESI-MS/NMR/CD | Yang et al., 2012 |
| 46 | (7S, 8R)-4,7,9,9'-tetrahydroxy-3,3'-dimethoxy-8-O-4'-neolignan-9'-O-β-D-apiofuranosyl-(1 → 6)-O-β-D-glucopyranoside | C ₃₁ H ₄₄ O ₁₆ | <i>A. lappa</i> | Fruits | IR/HR-ESI-MS/NMR/CD | Huang K. et al., 2015; Huang X.Y. et al., 2015 |
| 47 | (8R)-4,9,9'-trihydroxy-3,3'-dimethoxy-7-oxo-8-O-4'-neolignan-4-O-β-D-glucopyranoside | C ₂₆ H ₃₄ O ₁₂ | <i>A. lappa</i> | Fruits | IR/HR-ESI-MS/NMR/CD | Huang K. et al., 2015; Huang X.Y. et al., 2015 |
| 48 | (7R, 8S)-dihydrodehydrodiconiferyl alcohol-7'-oxo-4-O-β-D-glucopyranoside | C ₂₆ H ₃₂ O ₁₂ | <i>A. lappa</i> | Fruits | IR/HR-ESI-MS/NMR/CD | Huang K. et al., 2015; Huang X.Y. et al., 2015 |
| 49 | (7'S, 8'R, 8S)-4,4',9'-trihydroxy-3,3'-dimethoxy-7',9-epoxylignan-7-oxo-4-O-β-D-glucopyranoside | C ₂₆ H ₃₂ O ₁₂ | <i>A. lappa</i> | Fruits | IR/HR-ESI-MS/NMR/CD | Huang K. et al., 2015; Huang X.Y. et al., 2015 |
| 50 | Trachelogenin Terpenes/Terpenoids | C ₂₁ H ₂₄ O ₇ | <i>A. lappa</i> | Fruits | – | Ichikawa et al., 1986 |
| 51 | β-eudesmol | C ₁₅ H ₂₆ O | <i>A. lappa</i> | Fruits | – | Yayli et al., 2005 |
| 52 | Ursolic acid | C ₃₀ H ₄₈ O ₃ | <i>A. lappa</i> | Root | UV/IR/ESIMS/NMR | Han et al., 2013 |

(Continued)

TABLE 1 | Continued

| No | Compound name | Formula | Species | Plant origin/part | Analytical method | References |
|------------|--|---|---------------------------------------|---------------------|----------------------------|--|
| 53 | Oleanolic acid | C ₃₀ H ₄₈ O ₃ | <i>A. lappa</i> | Roots | UV/IR/ESIMS/NMR | Han et al., 2013 |
| 54 | Arctiopicrin | C ₁₉ H ₂₆ O ₆ | <i>A. lappa</i> , <i>A. minus</i> | Leaves | TLC/NMR | Savina et al., 2006 |
| 55 | Onopordopicrin | C ₁₉ H ₂₄ O ₆ | <i>A. lappa</i> , <i>A. nemorosum</i> | Leaves/aerial parts | TLC/HPLC/NMR/HR-ESI-TOF-MS | Barbosa et al., 1993; Savina et al., 2006; Machado et al., 2012; Zimmermann et al., 2012 |
| 56 | Dehydrovomifolol | C ₁₃ H ₁₈ O ₃ | <i>A. lappa</i> | Leaves | NMR/HR-ESI-TOF-MS | Machado et al., 2012 |
| 57 | Loliolide | C ₁₁ H ₁₆ O ₃ | <i>A. lappa</i> | Leaves | NMR/HR-ESI-TOF-MS | Machado et al., 2012 |
| 58 | Dehydromelitensin-8-(4'-hydroxymethacrylate) | C ₁₅ H ₂₄ O ₆ | <i>A. lappa</i> | Leaves | NMR/HR-ESI-TOF-MS | Machado et al., 2012 |
| 59 | Dehydromelitensin | C ₁₅ H ₂₀ O ₄ | <i>A. lappa</i> | Leaves | NMR/HR-ESI-TOF-MS | Machado et al., 2012 |
| 60 | Melitensin | C ₁₅ H ₂₂ O ₄ | <i>A. lappa</i> | Leaves | NMR/HR-ESI-TOF-MS | Machado et al., 2012 |
| 61 | 3 α -acetoxyp-22(29)-ene | C ₃₀ H ₄₉ O ₂ | <i>A. lappa</i> | Leaves | NMR, IR and MS | Jeelani and Khuroo, 2012 |
| 62 | 3 α -hydroxylanosta-5,15-diene | C ₃₀ H ₅₀ O | <i>A. lappa</i> | Leaves | NMR, IR and MS | Jeelani and Khuroo, 2012 |
| Flavonoids | | | | | | |
| 63 | Baicalin | C ₂₁ H ₁₈ O ₁₁ | <i>A. lappa</i> | Leaves/roots | UPLC/LC/MS/MS | Uchiyama et al., 2005 |
| 64 | Luteolin | C ₂₅ H ₂₄ O ₁₂ | <i>A. lappa</i> | | | Ferracane et al., 2010; Lou et al., 2010a; Tang et al., 2014 |
| 65 | Rutin | C ₂₇ H ₃₀ O ₁₆ | <i>A. lappa</i> , <i>A. minus</i> | Leaves | TLC/UPLC/LC/MS/MS | Saleh and Bohm, 1971; Lou et al., 2010a,b |
| 66 | Quercitrin | C ₂₁ H ₂₀ O ₁₁ | <i>A. lappa</i> | Leaves/roots | UPLC/LC/MS/MS | Lou et al., 2010a |
| 67 | Quercetin | C ₁₅ H ₁₀ O ₇ | <i>A. lappa</i> | Leaves/roots | UPLC/LC/MS/MS/HRESI-MS | Lou et al., 2010a; Predes et al., 2011; Tang et al., 2014 |
| 68 | Quercetin 3-O-glucuronide | C ₂₁ H ₁₈ O ₁₃ | <i>A. lappa</i> | Roots | HPTLC/LC/ESI-MS/MS | Rajasekharan et al., 2015 |
| 69 | Quercetin 3-vicianoside | C ₂₆ H ₂₈ O ₁₆ | <i>A. lappa</i> | Roots | HPTLC/LC/ESI-MS/MS | Rajasekharan et al., 2015 |
| 70 | Quercetin rhamnoside | C ₂₁ H ₂₀ O ₁₁ | <i>A. lappa</i> | roots | HPLC/LC/MS/MS | Ferracane et al., 2010 |
| 71 | Quercimeritrin | C ₂₁ H ₂₀ O ₁₂ | <i>A. minus</i> | Leaves | TLC | Saleh and Bohm, 1971 |
| 72 | Isoquercetin | C ₂₁ H ₂₀ O ₁₂ | <i>A. minus</i> | Leaves | TLC | Saleh and Bohm, 1971 |
| 73 | Astragalin | C ₂₁ H ₂₀ O ₁₁ | <i>A. minus</i> | Leaves | TLC | Saleh and Bohm, 1971 |
| 74 | Kaempferol-3-o-rhamnoglucoside | C ₂₇ H ₃₀ O ₁₅ | <i>A. minus</i> | Leaves | TLC | Saleh and Bohm, 1971 |
| 75 | Biachanin A | C ₁₆ H ₁₂ O ₅ | <i>A. lappa</i> | Roots | – | Tamayo et al., 2000; Eberding et al., 2007 |
| 76 | Genestein | C ₁₅ H ₁₀ O ₅ | <i>A. lappa</i> | Roots | – | Tamayo et al., 2000; Eberding et al., 2007 |
| 77 | Nobiletin | C ₂₁ H ₂₂ O ₈ | <i>A. lappa</i> | Roots | – | Tamayo et al., 2000; Eberding et al., 2007 |
| 78 | Tangeretin | C ₂₀ H ₂₀ O ₇ | <i>A. lappa</i> | Roots | – | Tamayo et al., 2000 |

(Continued)

TABLE 1 | Continued

| No | Compound name | Formula | Species | Plant origin/part | Analytical method | References |
|----------------------|-----------------------------------|---|--|--------------------|---|---|
| Sterols | | | | | | |
| 79 | β -sitosterol | C ₂₉ H ₅₀ O | <i>A. lappa</i> , <i>A. tomentosum</i> | Seeds/roots/fruits | UV/IR/MS/NMR/HPLC | Ting-Guo et al., 2001; Ming et al., 2004; Han et al., 2013 |
| 80 | Sitosterol-beta-D-glucopyranoside | C ₃₅ H ₆₀ O ₆ | <i>A. lappa</i> | Roots | IR/NMR/EI-MS | Miyazawa et al., 2005 |
| 81 | Daucosterol | C ₃₅ H ₆₀ O ₆ | <i>A. lappa</i> , <i>A. tomentosum</i> | Seeds/fruits | UV, IR, 1H-NMR, 13C-NMR, DEPT, 2D-NMR and MS/HPLC | Wang and Yang, 1993; Ting-Guo et al., 2001; Han et al., 2013 |
| Fatty acids | | | | | | |
| 82 | Docosanoic acid | C ₂₂ H ₄₄ O ₂ | <i>A. tomentosum</i> | Seeds | GCMS | Zong et al., 2013 |
| 83 | Eicosanoic acid | C ₂₀ H ₄₀ O ₂ | <i>A. tomentosum</i> | Seeds | GCMS | Zong et al., 2013 |
| 84 | <i>cis</i> -13-eicosenoic acid | C ₂₀ H ₃₈ O ₂ | <i>A. tomentosum</i> | Seeds | GCMS | Zong et al., 2013 |
| 85 | Methyl palmitate | C ₁₇ H ₃₄ O ₂ | <i>A. lappa</i> | – | IR/NMR/EI-MS | Miyazawa et al., 2005 |
| 86 | Methyl linoleate | C ₁₉ H ₃₄ O ₂ | <i>A. lappa</i> | – | IR/NMR/EI-MS | Miyazawa et al., 2005 |
| 87 | Methyl linolenate | C ₁₉ H ₃₂ O ₂ | <i>A. lappa</i> | Roots | IR/NMR/EI-MS/GCMS | Miyazawa et al., 2005; Kuo et al., 2012 |
| 88 | Methyl stearate | C ₁₉ H ₃₈ O ₂ | <i>A. lappa</i> | – | IR/NMR/EI-MS | Miyazawa et al., 2005 |
| 89 | Methyl oleate | C ₁₉ H ₃₆ O ₂ | <i>A. lappa</i> | Roots | IR/NMR/EI-MS/GCMS | Miyazawa et al., 2005; Kuo et al., 2012 |
| 90 | Hexadecanoic acid | C ₁₆ H ₃₂ O ₂ | <i>A. lappa</i> , <i>A. tomentosum</i> | Fruits/seeds | UV/TLC/IR/NMR/EIMS/GCMS | Miyazawa et al., 2005; Boldizar et al., 2010; Zong et al., 2013 |
| 91 | 9-hexadecenoic acid | C ₁₆ H ₃₀ O ₂ | <i>A. tomentosum</i> | Seeds | GCMS | Zong et al., 2013 |
| 92 | Linoleic acid | C ₁₈ H ₃₂ O ₂ | <i>A. lappa</i> | Roots | IR/NMR/EI-MS/GCMS | Miyazawa et al., 2005; Boldizar et al., 2010; Kuo et al., 2012 |
| 93 | Linolenic acid | C ₁₈ H ₃₀ O ₂ | <i>A. lappa</i> | Fruits | IR/NMR/EI-MS; GCMS | Miyazawa et al., 2005 |
| 94 | Stearic acid | C ₁₇ H ₃₅ CO ₂ H | <i>A. lappa</i> | Fruits | IR/NMR/EI-MS | Miyazawa et al., 2005 |
| 95 | 9,12-octadecadienoic acid | C ₁₈ H ₃₂ O ₂ | <i>A. tomentosum</i> | Seeds | GCMS | Zong et al., 2013 |
| 96 | Oleic acid | C ₁₈ H ₃₄ O ₂ | <i>A. lappa</i> | Fruits | UV/IR/HPLC/NMR/EI-MS | Miyazawa et al., 2005; Boldizar et al., 2010 |
| 97 | Oxiraneoctanoic acid | C ₁₉ H ₃₆ O ₃ | <i>A. tomentosum</i> | Seeds | GCMS | Zong et al., 2013 |
| 98 | Tetracosanoic acid | C ₂₄ H ₄₈ O ₂ | <i>A. tomentosum</i> | Seeds | GCMS | Zong et al., 2013 |
| Acetylenic compounds | | | | | | |
| 99 | Arctinone-a | C ₁₃ H ₁₀ O ₂ S ₂ | <i>A. lappa</i> | Roots | UV/TLC/IR/NMR/MS | Washino et al., 1986 |
| 100 | Arctinone-b | C ₁₃ H ₁₀ OS ₂ | <i>A. lappa</i> | Roots | UV/TLC/IR/NMR/MS | Washino et al., 1986 |
| 101 | Arctinol-a | C ₁₃ H ₁₂ O ₂ S ₂ | <i>A. lappa</i> | Roots | UV/TLC/IR/NMR/MS | Washino et al., 1986 |
| 102 | Arctinol-b | C ₁₃ H ₁₂ O ₂ S ₂ | <i>A. lappa</i> | Roots | UV/TLC/IR/NMR/MS | Washino et al., 1986 |
| 103 | Arctinal | C ₁₂ H ₈ OS ₂ | <i>A. lappa</i> | Roots | UV/TLC/IR/NMR/MS | Washino et al., 1986 |
| 104 | Arctic acid-b | C ₁₃ H ₈ O ₃ S ₂ | <i>A. lappa</i> | Roots | UV/TLC/IR/NMR/MS | Washino et al., 1986 |

(Continued)

TABLE 1 | Continued

| No | Compound name | Formula | Species | Plant origin/part | Analytical method | References |
|---|--------------------------------|---|-----------------|--------------------|--|---|
| 105 | Arctic acid-c | C ₁₃ H ₁₀ O ₃ S ₂ | <i>A. lappa</i> | Roots | UV/TLC/IR/NMR/MS | Washino et al., 1986 |
| 106 | Methyl arctate-b | C ₁₄ H ₁₀ O ₃ S ₂ | <i>A. lappa</i> | Roots | UV/TLC/IR/NMR/MS | Washino et al., 1986 |
| 107 | Arctinone-a acetate | C ₁₅ H ₁₀ O ₃ S ₂ | <i>A. lappa</i> | Roots | UV/TLC/IR/NMR/MS | Washino et al., 1986 |
| 108 | Dehydrodihydrocostus lactone | C ₁₅ H ₂₁ O ₂ | <i>A. lappa</i> | Roots | UV/TLC/IR/NMR/MS | Washino et al., 1986, 1987 |
| 109 | Dehydrocostus lactone | C ₁₅ H ₁₉ O ₂ | <i>A. lappa</i> | Roots | UV/TLC/IR/NMR/MS | Washino et al., 1986, 1987 |
| 110 | Lappaphen-a | C ₂₇ H ₂₆ O ₄ S | <i>A. lappa</i> | Roots | UV/TLC/IR/NMR/MS | Washino et al., 1986, 1987 |
| 111 | Lappaphen-b | C ₂₇ H ₂₆ O ₄ S | <i>A. lappa</i> | Roots | UV/TLC/IR/NMR/MS | Washino et al., 1986, 1987 |
| Carboxylic acids/Quinic acids and derivatives | | | | | | |
| 112 | Caffeic acid | C ₉ H ₈ O ₄ | <i>A. lappa</i> | Seeds/leaves/roots | TLC/HPLC/UPLC/LC/MS/ HRESI-MS | Chen et al., 2004; Lin and Harnly, 2008; Lou et al., 2010a,b; Ferracane et al., 2010; Predes et al., 2011; Tang et al., 2014; Lou et al., 2016; Al-Shammaa et al., 2017 |
| 113 | Caffeic acid 4-O-glucoside | C ₁₅ H ₁₈ O ₉ | <i>A. lappa</i> | Roots | LC-DAD-ESI/MS | Lin and Harnly, 2008; Lou et al., 2016 |
| 114 | Chlorogenic acid | C ₁₆ H ₁₈ O ₉ | <i>A. lappa</i> | Seeds/leaves/roots | TLC/HPTLC/HPLC/UPLC/LC/MS/MS/ QIT-TOF MS/ HRESI-MS | Wang et al., 2001; Chen et al., 2004; Lin and Harnly, 2008; Ferracane et al., 2010; Lou et al., 2010a,b; Predes et al., 2011; Liu et al., 2012; Haghi et al., 2013; Qin et al., 2014; Liu et al., 2015; Lou et al., 2016; Al-Shammaa et al., 2017 |
| 115 | <i>p</i> -coumaric acid | C ₉ H ₈ O ₃ | <i>A. lappa</i> | Seeds/leaves/roots | UPLC/EIMS | Lou et al., 2010a,b; Tang et al., 2014 |
| 116 | Coumaroylquinic acid | C ₁₆ H ₁₈ O ₈ | <i>A. lappa</i> | Roots | HPTLC/LC/ESI-MS/MS | Rajasekharan et al., 2015 |
| 117 | Benzoic Acid | C ₇ H ₆ O ₂ | <i>A. lappa</i> | Leaves | UPLC/EIMS | Lou et al., 2010a,b |
| 118 | Cynarin | C ₂₅ H ₂₄ O ₁₂ | <i>A. lappa</i> | Seeds/leaves/roots | UPLC/LC/MS | Ferracane et al., 2010; Lou et al., 2010a, 2016; Tang et al., 2014 |
| 119 | Caffeoyl-hexose-hydroxyphenol | C ₂₁ H ₂₁ O ₁₀ | <i>A. lappa</i> | Roots | HPTLC/LC/ESI-MS/MS | Rajasekharan et al., 2015 |
| 120 | 1-O-caffeoylquinic acid | C ₁₆ H ₁₈ O ₉ | <i>A. lappa</i> | Roots | GCMS/LC-DAD-ESI/MS | Lin and Harnly, 2008; Tusch et al., 2014 |
| 121 | 3-O-caffeoylquinic acid | C ₁₆ H ₁₈ O ₉ | <i>A. lappa</i> | Roots | GCMS/LC-DAD-ESI/MS | Lin and Harnly, 2008; Tusch et al., 2014 |
| 122 | 4-O-caffeoylquinic acid | C ₁₆ H ₁₈ O ₉ | <i>A. lappa</i> | Roots | GCMS/LC-DAD-ESI/MS | Lin and Harnly, 2008; Tusch et al., 2014 |
| 123 | 5-O-caffeoylquinic acid | C ₁₆ H ₁₈ O ₉ | <i>A. lappa</i> | Roots | GCMS/LC-DAD-ESI/MS | Lin and Harnly, 2008; Jaiswal and Kuhnert, 2011; Tusch et al., 2014 |
| 124 | 1-O-,5-O-dicaffeoylquinic acid | C ₂₅ H ₂₄ O ₁₂ | <i>A. lappa</i> | Roots | HPLC/NMR/MS | Maruta et al., 1995; Wang et al., 2001; Han et al., 2013; Rajasekharan et al., 2015 |

(Continued)

TABLE 1 | Continued

| No | Compound name | Formula | Species | Plant origin/part | Analytical method | References |
|-----|--|---|-----------------|--------------------|--------------------------------------|---|
| 125 | 1-O-, 5-O-dicaffeoyl-3-O-succinylquinic acid | C ₃₅ H ₄₀ O ₁₅ | <i>A. lappa</i> | Roots | NMR/EI-MS | Maruta et al., 1995 |
| 126 | 1-O-, 5-O-dicaffeoyl-4-O-succinylquinic acid | C ₂₉ H ₃₅ O ₁₅ | <i>A. lappa</i> | Roots | NMR/MS | Maruta et al., 1995 |
| 127 | 1-O-, 5-O-dicaffeoyl-3-O- | C ₃₃ H ₃₉ O ₁₈ | <i>A. lappa</i> | Roots | NMR/MS | Maruta et al., 1995 |
| 128 | 4-O-disuccinylquainic acid and 1-O-, 3-O-, 5-O-tricaffeoyl-4-O-succinylquinic acid | C ₃₈ H ₄₁ O ₁₈ | <i>A. lappa</i> | Roots | NMR/MS | Maruta et al., 1995 |
| 129 | 1,3-di-O-caffeoylquinic acid | C ₂₅ H ₂₄ O ₁₂ | <i>A. lappa</i> | Seeds/roots | LCMS/ MALDI-QIT-TOF MS | Lin and Harnly, 2008; Liu et al., 2012 |
| 130 | 1,5-di-O-caffeoylquinic acid | C ₂₅ H ₂₄ O ₁₂ | <i>A. lappa</i> | Leaves/Seeds/roots | UPLC/HPLC/PDA/LCMS/ MALDI-QIT-TOF MS | Maruta et al., 1995; Lin and Harnly, 2008; Liu et al., 2012; Haghi et al., 2013; Tusch et al., 2014 |
| 131 | 1,5-di-O-caffeoyl-4-O-maloylquinic acid | C ₂₉ H ₂₇ O ₁₆ | <i>A. lappa</i> | Roots | LCMS/ MALDI-QIT-TOF MS | Jaiswal and Kuhnert, 2011; Liu et al., 2012; Tusch et al., 2014 |
| 132 | 1,5-di-O-caffeoyl-3-O-maloylquinic acid | C ₂₅ H ₂₇ O ₁₆ | <i>A. lappa</i> | Roots | LCMS/ MALDI-QIT-TOF MS | Jaiswal and Kuhnert, 2011; Liu et al., 2012; Tusch et al., 2014 |
| 133 | 1,5-di-O-caffeoyl-3-O-succinoylquinic acid | C ₂₉ H ₂₇ O ₁₅ | <i>A. lappa</i> | Roots | LCMS/ MALDI-QIT-TOF MS | Maruta et al., 1995; Jaiswal and Kuhnert, 2011; Liu et al., 2012; Tusch et al., 2014 |
| 134 | 1,5-di-O-caffeoyl-3,4-di-O-succinoylquinic acid | C ₃₃ H ₃₁ O ₁₈ | <i>A. lappa</i> | Roots | LCMS/ MALDI-QIT-TOF MS | Liu et al., 2012; Tusch et al., 2014 |
| 135 | 1,3,5-tri-O-caffeoyl-4-O-succinoylquinic acid | C ₃₈ H ₃₃ O ₁₈ | <i>A. lappa</i> | Roots | GCMS/LCMS/ MALDI-QIT-TOF MS | Jaiswal and Kuhnert, 2011; Liu et al., 2012; Tusch et al., 2014 |
| 136 | 1,3,5-tri-O-caffeoylquinic acid | C ₃₄ H ₂₉ O ₁₅ | <i>A. lappa</i> | Roots | GCMS/LCMS/ MALDI-QIT-TOF MS | Liu et al., 2012; Tusch et al., 2014 |
| 137 | 1,5-di-O-caffeoyl-3-O-succinoyl-4-O-maloylquinic acid | – | <i>A. lappa</i> | Roots | LCMS/ MALDI-QIT-TOF MS | Liu et al., 2012 |
| 138 | 5-sinapoylquinic acid | C ₁₈ H ₂₂ O ₁₀ | <i>A. lappa</i> | Roots | LC-DAD-ESI/MS | Lin and Harnly, 2008 |
| 139 | 3-sinapoyl-5-caffeoylquinic acid | C ₂₇ H ₂₈ O ₁₃ | <i>A. lappa</i> | Roots | LC-DAD-ESI/MS | Lin and Harnly, 2008 |
| 140 | 3-sinapoyl-5-caffeoyl-1-methoxyoxaloylquinic acid | – | <i>A. lappa</i> | Roots | LC-DAD-ESI/MS | Lin and Harnly, 2008 |

(Continued)

TABLE 1 | Continued

| No | Compound name | Formula | Species | Plant origin/part | Analytical method | References |
|-----|--|---|-----------------|-------------------|--------------------|--|
| 141 | 4-sinapoyl-5-caffeoyl-1-methoxyoxaloylquinic acid | – | <i>A. lappa</i> | Roots | LC-DAD-ESI/MS | Lin and Harnly, 2008 |
| 142 | 3,4-dicaffeoylquinic acid | C ₂₅ H ₂₄ O ₁₂ | <i>A. lappa</i> | Roots/seeds | LC-DAD-ESI/MS | Lin and Harnly, 2008 |
| 143 | 1,4-di- <i>O</i> -caffeoylquinic acid | C ₂₅ H ₂₃ O ₁₂ | <i>A. lappa</i> | Roots | GCMS/LC-DAD-ESI/MS | Lin and Harnly, 2008; Jaiswal and Kuhnert, 2011; Tusch et al., 2014 |
| 144 | 3,5-di- <i>O</i> -caffeoylquinic acid | C ₂₅ H ₂₄ O ₁₂ | <i>A. lappa</i> | Roots | GCMS/LC-DAD-ESI/MS | Lin and Harnly, 2008; Jaiswal and Kuhnert, 2011; Tusch et al., 2014 |
| 145 | 4,5-dicaffeoylquinic acid | C ₂₅ H ₂₄ O ₁₂ | <i>A. lappa</i> | Roots/seeds | LC-DAD-ESI/MS | Lin and Harnly, 2008 |
| 146 | 3,5-dicaffeoyl-1-methoxyoxaloylquinic acid | – | <i>A. lappa</i> | Roots | LC-DAD-ESI/MS | Lin and Harnly, 2008 |
| 147 | 3-feruloyl-5-caffeoylquinic acid | – | <i>A. lappa</i> | Roots | LC-DAD-ESI/MS | Lin and Harnly, 2008 |
| 148 | 4,5-dicaffeoyl-1-methoxyoxaloylquinic acid | – | <i>A. lappa</i> | Roots | LC-DAD-ESI/MS | Lin and Harnly, 2008 |
| 149 | 3-sinapoyl-5-caffeoyl-4-methoxyoxaloylquinic acid | – | <i>A. lappa</i> | Roots | LC-DAD-ESI/MS | Lin and Harnly, 2008 |
| 150 | 1,4,5-tricaffeoylquinic acid | C ₃₄ H ₃₀ O ₁₅ | <i>A. lappa</i> | Roots | LC-DAD-ESI/MS | Lin and Harnly, 2008 |
| 151 | 3,4,5-tricaffeoylquinic acid | – | <i>A. lappa</i> | Roots | LC-DAD-ESI/MS | Lin and Harnly, 2008 |
| 152 | 1,4,5-tricaffeoyl-3-methoxyoxaloylquinic acid | – | <i>A. lappa</i> | Roots | LC-DAD-ESI/MS | Lin and Harnly, 2008 |
| 153 | 3-succinoyl-4,5-dicaffeoyl | – | <i>A. lappa</i> | Roots | LCMS | Jaiswal and Kuhnert, 2011 |
| 154 | 1,5-dicaffeoyl-3-succinoylquinic acid | – | <i>A. lappa</i> | Roots | HPLC/LCMS | Wang et al., 2001; Jaiswal and Kuhnert, 2011 |
| 155 | 1,5-di- <i>O</i> -caffeoyl-4- <i>O</i> -succinoylquinic acid | C ₂₉ H ₂₇ O ₁₅ | <i>A. lappa</i> | Roots | GCMS/LCMS | Maruta et al., 1995; Jaiswal and Kuhnert, 2011; Liu et al., 2012; Tusch et al., 2014 |
| 156 | 3,4-dicaffeoyl-5-succinoylquinic acid | C ₂₉ H ₂₈ O ₁₅ | <i>A. lappa</i> | Roots | LCMS | Jaiswal and Kuhnert, 2011 |
| 157 | 1,3-dicaffeoyl-5-fumaroylquinic acid | – | <i>A. lappa</i> | Roots | LCMS | Jaiswal and Kuhnert, 2011 |
| 158 | 1,5-dicaffeoyl-4-fumaroylquinic acid | – | <i>A. lappa</i> | Roots | LCMS | Jaiswal and Kuhnert, 2011 |
| 159 | 1,5-dicaffeoyl-3-maloylquinic acid | – | <i>A. lappa</i> | Roots | LCMS | Jaiswal and Kuhnert, 2011 |

(Continued)

TABLE 1 | Continued

| No | Compound name | Formula | Species | Plant origin/part | Analytical method | References |
|-----|---|---|-----------------|-------------------|-------------------|---|
| 160 | 1,4-di-O-caffeoyl-3-O-maloylquinic Acid | C ₂₉ H ₂₇ O ₁₆ | <i>A. lappa</i> | Roots | LCMS | Jaiswal and Kuhnert, 2011 |
| 161 | 1,3-di-O-caffeoyl-4,5-di-O-maloylquinic | C ₃₃ H ₃₁ O ₂₀ | <i>A. lappa</i> | Roots | GCMS/LCMS | Jaiswal and Kuhnert, 2011; Tusch et al., 2014 |
| 162 | 1,5-dicaffeoyl-4-maloylquinic acid | – | <i>A. lappa</i> | Roots | LCMS | Jaiswal and Kuhnert, 2011 |
| 163 | 1,4-di-O-maloyl-3,5-di-O-caffeoylquinic acid | C ₃₁ H ₃₃ O ₂₀ | <i>A. lappa</i> | Roots | GCMS/LCMS | Jaiswal and Kuhnert, 2011; Tusch et al., 2014 |
| 164 | 1,3,5-tricaffeoyl-4-succinoylquinic acid | – | <i>A. lappa</i> | Roots | LCMS | Jaiswal and Kuhnert, 2011 |
| 165 | 1,5-dicaffeoyl-3,4-disuccinoylquinic acid | – | <i>A. lappa</i> | Roots | LCMS | Jaiswal and Kuhnert, 2011 |
| 166 | 1,5-dicaffeoyl-3-fumaroyl-4-succinoylquinic acid | – | <i>A. lappa</i> | Roots | LCMS | Jaiswal and Kuhnert, 2011 |
| 167 | 1-fumaroyl-3,5-dicaffeoyl-4-succinoylquinic acid | – | <i>A. lappa</i> | Roots | LCMS | Jaiswal and Kuhnert, 2011 |
| 168 | 1,5-di-O-caffeoyl-3-O-succinoyl-4-O-maloylquinic acid | C ₃₃ H ₃₁ O ₁₉ | <i>A. lappa</i> | Roots | GCMS/LCMS | Jaiswal and Kuhnert, 2011; Tusch et al., 2014 |
| 169 | Dimaloyl-dicaffeoylquinic acid isomer 1 | C ₃₃ H ₃₁ O ₂₀ | <i>A. lappa</i> | Roots | GCMS/LCMS | Tusch et al., 2014 |
| 170 | Succinoyl-tricaffeoylquinic acid isomer | C ₃₈ H ₃₃ O ₁₈ | <i>A. lappa</i> | Roots | GCMS/LCMS | Tusch et al., 2014 |
| 171 | Maloyl-dicaffeoylquinic acid isomer | C ₂₉ H ₂₇ O ₁₅ | <i>A. lappa</i> | Roots | GCMS/LCMS | Tusch et al., 2014 |
| 172 | Dicaffeoyl-succinoyl-malonylquinic acid isomer 1 | C ₃₃ H ₃₁ O ₁₉ | <i>A. lappa</i> | Roots | GCMS/LCMS | Tusch et al., 2014 |
| 173 | Dicaffeoyl-succinoyl-malonylquinic acid isomer 2 | C ₃₃ H ₃₁ O ₂₀ | <i>A. lappa</i> | Roots | GCMS/LCMS | Tusch et al., 2014 |
| 174 | Dimaloyl-dicaffeoylquinic acid isomer 2 | C ₃₃ H ₃₁ O ₂₀ | <i>A. lappa</i> | Roots | GCMS/LCMS | Tusch et al., 2014 |
| 175 | Dimaloyl-dicaffeoylquinic acid isomer 3 | C ₃₃ H ₃₁ O ₂₀ | <i>A. lappa</i> | Roots | GCMS/LCMS | Tusch et al., 2014 |

(Continued)

TABLE 1 | Continued

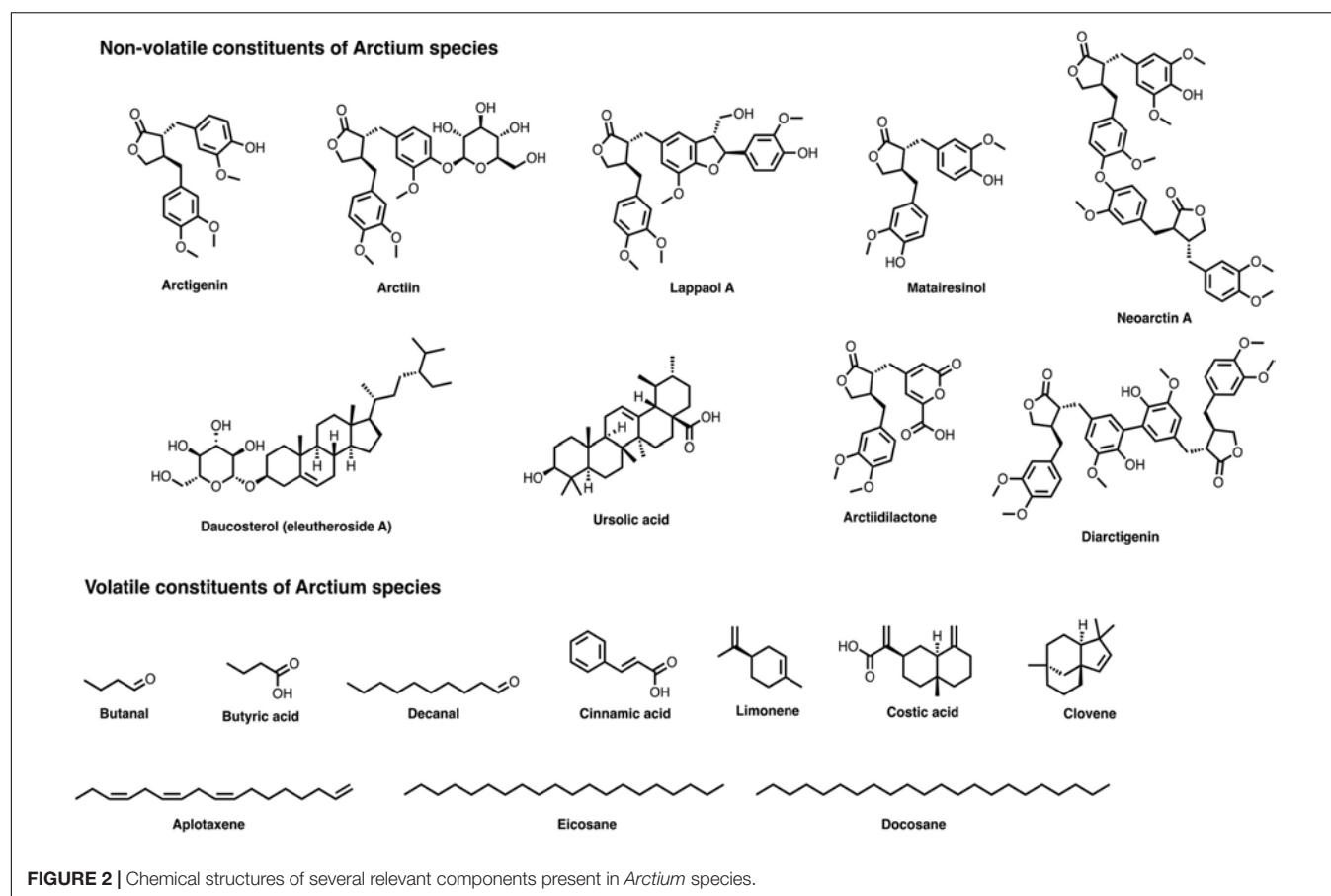
| No | Compound name | Formula | Species | Plant origin/part | Analytical method | References |
|-----|--|---|--|---------------------|-----------------------------------|---|
| 176 | Maloyl-tricaffeoylquinic isomer | C ₂₈ H ₃₂ O ₁₉ | <i>A. lappa</i> | Roots | GCMS/LCMS | Tousch et al., 2014 |
| 177 | 1,3,5-tri-O-caffeoyl-4-O-maloylquinic Acid | C ₃₈ H ₃₃ O ₁₉ | <i>A. lappa</i> | Roots | GCMS/LCMS | Tousch et al., 2014 |
| 178 | 5-hydroxymaltol | C ₆ H ₆ O ₄ | <i>A. lappa</i> | Roots | UV/IR/ESIMS/NMR | Han et al., 2013 |
| 179 | Succinic acid | C ₄ H ₆ O ₄ | <i>A. lappa</i> | Roots | UV/IR/ESIMS/NMR | Han et al., 2013 |
| 180 | Saccharides/Polysaccharides | | | | | |
| 180 | Rhamnogalacturonan | C ₁₁₇ H ₁₇₈ O ₁₀₁ | <i>A. lappa</i> , <i>A. minus</i> | Roots/leaves | Chromatography/NMR/sugar analysis | Kato and Watanabe, 1993; Carlotto et al., 2016 |
| 181 | Xylan | (C ₅ H ₈ O ₄) _n | <i>A. lappa</i> , <i>A. minus</i> | Roots/leaves | Chromatography/NMR/sugar analysis | Kato and Watanabe, 1993 |
| 182 | Arabinan | C ₉ H ₁₃ N ₃ O ₅ | <i>A. lappa</i> , <i>A. minus</i> | Roots/leaves | Chromatography/NMR/sugar analysis | Kato and Watanabe, 1993; Carlotto et al., 2016 |
| 183 | Arabinogalactan | C ₂₀ H ₃₆ O ₁₄ | <i>A. lappa</i> , <i>A. minus</i> | Roots/leaves | Chromatography/NMR/sugar analysis | Kato and Watanabe, 1993; Carlotto et al., 2016 |
| 184 | Galactan | C ₁₈ H ₃₂ O ₁₆ | <i>A. lappa</i> , <i>A. minus</i> | Roots/leaves | Chromatography/NMR/sugar analysis | Kato and Watanabe, 1993 |
| 185 | Cellulose | C ₆₄ H ₁₂₄ O ₃₀ | <i>A. lappa</i> , <i>A. minus</i> | Roots/leaves | Chromatography/NMR/sugar analysis | Kato and Watanabe, 1993 |
| 186 | Xyloglucan | C ₅₁ H ₈₆ O ₄₂ | <i>A. lappa</i> , <i>A. minus</i> | Roots/leaves | Chromatography/NMR/sugar analysis | Kato and Watanabe, 1993 |
| 187 | Galacturonic acid | C ₆ H ₁₀ O ₇ | <i>A. lappa</i> | Roots/leaves | Chromatography/ NMR | Carlotto et al., 2016 |
| 188 | Galacturonic acid | C ₆ H ₁₀ O ₇ | <i>A. lappa</i> | Roots | Chromatography | Fuchigami et al., 1990 |
| 189 | Galactose | C ₆ H ₁₂ O ₆ | <i>A. lappa</i> | Roots/leaves/fruits | Chromatography/ NMR | Fuchigami et al., 1990; Kardosova et al., 2003; Boldizar et al., 2010; Carlotto et al., 2016 |
| 190 | Glucose | C ₆ H ₁₂ O ₆ | <i>A. lappa</i> | Roots/leaves/fruits | UV/NMR/HPLC/GCMS | Kardosova et al., 2003; Boldizar et al., 2010; Li et al., 2013; Carlotto et al., 2016 |
| 191 | Mannose | C ₆ H ₁₂ O ₆ | <i>A. lappa</i> | Roots/leaves | NMR | Carlotto et al., 2016 |
| 192 | Sucrose | C ₁₂ H ₂₂ O ₁₁ | <i>A. lappa</i> | Roots | UV/NMR/HPLC/GCMS | Boldizar et al., 2010; Li et al., 2013 |
| 193 | Raffinose | C ₁₈ H ₃₂ O ₁₆ | <i>A. lappa</i> | Fruits | UV/NMR/HPLC/GCMS | Boldizar et al., 2010 |
| 194 | Rhamnose | C ₆ H ₁₂ O ₅ | <i>A. lappa</i> | Roots/leaves/fruits | UV/NMR/HPLC/GCMS | Boldizar et al., 2010; Carlotto et al., 2016 |
| 195 | Arabinose | C ₅ H ₁₀ O ₅ | <i>A. lappa</i> | Roots/leaves/fruits | UV/NMR/HPLC/GCMS | Fuchigami et al., 1990; Kardosova et al., 2003; Boldizar et al., 2010; Carlotto et al., 2016 |
| 196 | Inulin (fructan) | (C ₆ H ₁₀ O ₅) _n | <i>A. lappa</i> , <i>A. tomentosum</i> | Roots | HPTLC/MS/NMR/HPLC-ELSD | Kardosova et al., 2003; Turdumambetov et al., 2004; Milani et al., 2011; Olennikov and Tankhaeva, 2011; Li et al., 2013; Liu et al., 2014 |
| 197 | Fructose | C ₆ H ₁₂ O ₆ | <i>A. lappa</i> | Roots | HPLC-ELSD | Li et al., 2013 |

(Continued)

TABLE 1 | Continued

| No | Compound name | Formula | Species | Plant origin/part | Analytical method | References |
|-----|----------------------|---|--|-------------------|-------------------|-----------------------|
| 198 | Sorbitol | C ₆ H ₁₄ O ₆ | <i>A. lappa</i> | Fruits | UV/NMR/HPLC/GCMS | Boldizar et al., 2010 |
| 199 | Mannitol | C ₆ H ₁₄ O ₆ | <i>A. lappa</i> | Fruits | UV/NMR/HPLC/GCMS | Boldizar et al., 2010 |
| 200 | Others | | | | | |
| 200 | Crocin | C ₄₄ H ₆₄ O ₂₄ | <i>A. lappa</i> | Leaves | UPLC | Lou et al., 2016 |
| 201 | <i>b</i> -asparagine | C ₄ H ₈ N ₂ O ₃ | <i>A. lappa</i> , <i>A. tomentosum</i> | Roots | IR/NMR | Boev, 2005 |

fruits (Carlotto et al., 2016). A high-speed counter-current chromatography was employed to obtain the pure compound arctiin from the fruit extracts of *A. lappa*. Authors obtained 49% of arctiin identified based on LC-MS and NMR techniques (Wang et al., 2005). A novel butyrolactone lignan compound named diartigenin was found to occur in the methanolic seed extracts of *A. lappa* (Han et al., 1994). The fruits of *A. lappa* are reported to contain a total of 13 compounds including 5 new natural products (Umehara et al., 1993). Among them, 6 compounds were identified as arctignan A-E and arctigenin. Later, the occurrence of arctigenin and arctiin was also established from the leaves and seeds of *A. lappa* (Umehara et al., 1993; Liu et al., 2005; Matsumoto et al., 2006). Besides, the active extract resulted from the bioassay-guided fractionation of seed methanolic extract contained five active compounds including a new sesquilignan named isolappaol C and four known sesquilignan and dilignans namely, diartigenin, and lappaol C, D, and F (Ferracane et al., 2010). Further, improved methods of extraction and analysis revealed that the seeds and roots of *A. lappa* contain arctigenin, arctiin, arctignan E, matareisinol, lappaol A, C, and F (Ferracane et al., 2010; Lou et al., 2010a, 2016; Liu et al., 2015; Su et al., 2015). The occurrence of 8 lignans in seeds and 1 lignan in roots of *A. lappa* was determined. The identified lignans were arctiin, arctigenin arctignan D and E, lappaol A, C, and H, isolappaol C and matairesinol (Liu et al., 2012; Haghi et al., 2013). Likewise, syringaresinol was reported in the chloroform fraction of *A. lappa* roots (Han et al., 2013). A rare butyrolactone lignan named arctiidilactone, and 11 novel butyrolactone lignans [arctigenin-4-*O*-β-*D*-gentiobioside, arctigenin-4-*O*-α-*D*-galactopyranosyl-(1→6)-*O*-β-*D*-glucopyranoside, arctigenin-4-*O*-β-*D*-apiofuranosyl-(1→6)-*O*-β-*D*-glucopyranoside, 5'-propanediolmatairesinoside, (7'*R*,8*R*,8'*R*)-rafanotrachelogenin-4-*O*-β-*D*-glucopyranoside, (7'*S*,8*R*,8'*R*)-rafanotrachelogenin-4-*O*-β-*D*-glucopyranoside, (7*S*,8*S*,8'*R*)-4,7-dihydroxy-3,3', 4-trimethoxyl-9-oxo benzylbutyrolactone lignan-4-*O*-β-*D*-glucopyranoside, (7*R*,8*S*,8'*R*) -4,7,4'-trihydroxy-3,3'-dimethoxyl-9-oxo dibenzylbutyrolactone lignan-4-*O*-β-*D*-glucopyranoside, (7*S*,8*S*,8'*R*)-4,7-dihydroxy-3,3',4'-trimethoxyl-9-oxo dibenzylbutyrolactone lignan, arctiidilactone, arctiapolignan A and arctiisesquineolignan A] were determined in *A. lappa* fruits (Yang et al., 2015). Phylligenin, matairesinoside and pinoresinol were reported only in the fruits of *A. lappa* (Boldizar et al., 2010). Also, 2 secolignans, styraxlignolide D and styraxlignolide E (Yang et al., 2015) and 4 new neolignan glucosides namely, (8*R*)-4,9,9'-trihydroxy-3,3'-dimethoxy-7-oxo-8-*O*-4'-neolignan-4-*O*-β-*D*-glucopyranoside, (7*S*,8*R*)-4,7,9,9'-tetrahydroxy-3,3'-dimethoxy-8-*O*-4'-neolignan-9'-*O*-β-*D*-apiofuranosyl-(1→6)-*O*-β-*D*-glucopyranoside, (7'*S*,8'*R*,8*S*)-4,4',9'-trihydroxy-3,3'-dimethoxy -7',9-epoxylignan-7-oxo-4-*O*-β-*D*-glucopyranoside and (7*R*, 8*S*)-dihydrodehydrodiconiferyl alcohol-7'-oxo-4-*O*-β-*D*-glucopyranoside are reported from *A. lappa* fruits (Huang X.Y. et al., 2015). Besides, phytochemical analysis of *A. lappa* fruits revealed the existence of 2 more lignans named arctiisesquineolignan B and arctiiphenolglycoside A (He et al., 2016). Bioassay-guided separation and purification of hydroethanolic extracts of *A. lappa* fruits allowed to identify a



new lignan, (+)-7,8-didehydroarctigenin along with arctigenin and matairesinol identified previously (Matsumoto et al., 2006).

Fatty Acids and Esters

In search of α -glucosidase inhibitory compounds, Miyazawa et al. (2005) found 11 compounds in *A. lappa* methanol extract. Among them, 10 compounds belonged to fatty acids. The identified compounds were linolenic acid, linoleic acid, methyl linoleate, methyl oleate, methyl linolenate, oleic acid, palmitic acid, methyl palmitate, methyl stearate, and stearic acid. Methanol extract from *Arctium lappa* L. which was found to contain sitosterol- β -D-glucopyranoside, methyl palmitate, methyl linoleate and methyl linolenate showed an inhibitory activity against α -glucosidase at 97.3, 73.4, 66.5, and 68.5% respectively at a concentration of 200.0 μ M (Miyazawa et al., 2005). Later, Kuo et al. (2012) identified methyl methyl α -linolenate, linolenic acid and methyl oleate as the chief constituents in the *n*-hexane fraction of *A. lappa* root (Kuo et al., 2012). The presence of linoleic acid, oleic acid, palmitic acid and stearic acid were also reported from *A. lappa* fruits (Boldizsar et al., 2010). Fatty acid composition of *A. tomentosum* seeds showed the occurrence of docosanoic acid, hexadecanoic acid, 9-hexadecenoic acid, 9,12-octadecadienoic acid, oxiraneoctanoic acid, eicosanoic acid, *cis*-13-eicosenoic acid, and tetracosanoic acid (Zhou et al., 2011).

Acetylenic Compounds

From the roots of *A. lappa*, Washino et al. (1986) isolated and characterized 9 sulfur-containing acetylenic compounds namely, arctinone-a, arctinone-b, arctinol-a, arctinol-b, arctinal, arctic acid-b, arctic acid-c, methyl arctate-b, and arctinone-a acetate. Based on the chemical and spectral analysis, it was found that all these compounds were derivatives of 5'-(1-propynyl)-2,2'-bithienyl-5-yl. Later, the occurrence of few guaianolides linked with a sulfur-containing acetylenic compounds namely dehydridihydrocostus lactone, dehydrocostus lactone, lappaphen-a and lappaphen-b were discovered from the acetone extracts of *A. lappa* roots (Washino et al., 1986, 1987). Several bioactivities of the key *A. lappa* constituents have been well-described in literature including antibacterial and antifungal activities of acetylenic compounds (Takasugi et al., 1987) and anti-edematogenic activity on carrageenan-induced paw edema (Carlotto et al., 2016).

Phytosterols

Daucosterol, a natural phytosterol-like compound, was obtained from the seeds of *A. lappa* (Ahangarpour et al., 2017). The fruits of *A. tomentosum* are reported to contain 2 steroids, such as daucosterol and β -sitosterol. Using bioactivity-guided fractionation, daucosterol and β -sitosterol were recovered from the ethanolic extract (95%) of *A. lappa* seeds (Ming et al., 2004). Later, sitosterol-beta-D-glucopyranoside was found in the

methanolic extracts of *A. lappa* (Miyazawa et al., 2005). Also, daucosterol and β -sitosterol compounds were detected from the chloroform extracts of *A. lappa* roots (Han et al., 2013). It was shown that phytosterol daucosterol inhibited cancer cell proliferation by inducing autophagy through reactive oxygen species-dependent manner (Zhao et al., 2015), and exhibited immunoregulatory activity by inducing protective Th1 immune response (Lee et al., 2007).

Polysaccharides

For the first time, Fuchigami et al. (1990; Ferracane et al., 2010) determined the pectic polysaccharides in edible *A. lappa* roots. Later investigations revealed the occurrence of several kinds of polysaccharides such as pectic substance, rhamnogalacturonan with neutral sugars, hemicellulose (arabinan, arabinogalactan, galactan, xylan, and xyloglucan), galacturonic acid, glucose, galactose, arabinose, rhamnose, mannose, and cellulose in cell walls of *A. lappa* and *A. minus* roots and leaves (Kato and Watanabe, 1993; Carlotto et al., 2016). Also, arabinose, glucose, galactose, rhamnose, and raffinose are reported from fruits of *A. lappa* (Boldizar et al., 2010). The xyloglucan characterized from *A. minus* comprised a repeated unit of oligosaccharides of hepta-(Glc-Xyl = 4:3), deca-(Glc-Xyl-Gal-Fuc = 4:3:2:1) and nona-(Glc-Xyl-Gal-Fuc = 4:3:1:1) saccharides in the ratio of 14:5:12 (Kato and Watanabe, 1993). Biologically active inulin-type fructofuranans and other fructooligosaccharides have been identified from the roots of *A. lappa* (Kardosova et al., 2003). Inulin, a fiber comprising oligomers and polymers of fructose units linked by $\beta(2 \rightarrow 1)$ fructosyl-fructose bonds, has also been reported in the roots of *A. lappa* (Rajasekharan et al., 2015). A water-soluble polysaccharide fructan with a molecular weight of 4,600 Da, named as ALP1, was purified from *A. lappa* root and was composed of fructose and glucose in the molar ratio of 13:1. They were linked in $\rightarrow(1)\text{-Fruf-}(2)\rightarrow$, $\text{Fruf-}(2)\rightarrow$ and $\text{Glc-}(1)\rightarrow$ (Liu et al., 2014). The structure was similar to the crude fructan obtained previously by Kardosova et al. (2003). In *A. tomentosum*, the glucofructans content is 24%, constituted by a polymer of 2 inulin type (GF-A and GF-B) and 1 graminan (a mixed type of glucofructans containing 1,2- and 2-6 bonds) type polysaccharides. HPTLC method was developed by Olennikov and Tankhaeva (2011) to quantify fructans in *A. tomentosum* and *A. lappa* (Olennikov and Tankhaeva, 2011). Two sugar alcohols, mannitol and sorbitol were reported from the fruits of *A. lappa* (Boldizar et al., 2010). The yield of inulin from *A. lappa* root was successfully increased by adopting an ultrasonic extraction technology (Milani et al., 2011). It was indicated that water-soluble polysaccharide from *A. lappa* could significantly ameliorate the dysregulation of pro-inflammatory cytokines (IL-1 β , IL-6, and TNF- α) and anti-inflammatory cytokine (IL-10) caused by colitis (Wang et al., 2019).

Caffeoylquinic Acid Derivatives (Carboxylic Acids)

Caffeoylquinic acids are the major bioactive phenolic compounds of *Arctium* species and impart superior antioxidant properties to the plant. The roots of *A. lappa* were reported to contain caffeoylquinic acid derivatives such

as 1-*O*-,5-*O*-dicaffeoylquinic acid, 1-*O*-,5-*O*-dicaffeoyl-3-*O*-succinylquinic acid, 1-*O*-,5-*O*-dicaffeoyl-4-*O*-succinylquinic acid, 1-*O*-,5-*O*-dicaffeoyl-3-*O*-,4-*O*-disuccinylquinic acid and 1-*O*-,3-*O*-,5-*O*-tricaffeoyl-4-*O*-succinylquinic acid (Maruta et al., 1995). Chlorogenic acid content is much higher than the caffeic acid and both occur mainly in the skin of *A. lappa* roots (Chen et al., 2004). HPTLC analysis was used as a chemical profiling tool to estimate chlorogenic acid in *A. lappa* roots. The content ranged from 0.107 to 0.140%. Lin and Harnly (2008) and Liu et al. (2012) identified several compounds, including 5-sinapoylquinic acid, 3-sinapoyl-5-caffeoylquinic acid, 3-sinapoyl-5-caffeoyl-1-methoxyoxaloylquinic acid, 4-sinapoyl-5-caffeoyl-1-methoxyoxaloylquinic acid, 1,4-dicaffeoylquinic acid, 3,4-dicaffeoylquinic acid, 4,5-dicaffeoylquinic acid, 3,5-dicaffeoylquinic acid, 3,5-dicaffeoyl-1-methoxyoxaloylquinic acid, 3-feruloyl-5-caffeoylquinic acid, 4,5-dicaffeoyl-1-methoxyoxaloylquinic acid, 3,5-dicaffeoyl-1-methoxyoxaloylquinic acid, 3-feruloyl-5-caffeoylquinic acid, 4,5-dicaffeoyl-1-methoxyoxaloylquinic acid, 3,4,5-tricaffeoylquinic acid, 1,4,5-tricaffeoylquinic acid and 1,4,5-tricaffeoyl-3-methoxyoxaloylquinic acid from the roots of *A. lappa*. Jaric et al. (2007) and Jaiswal and Kuhnert (2011) have characterized succinic, fumaric and malic acid-containing chlorogenic acid from the roots of *A. lappa*. These compounds included 3-succinoyl-4,5-dicaffeoyl, 1,5-dicaffeoyl-4-succinoylquinic acid, 1,5-dicaffeoyl-3-succinoylquinic acid, 3,4-dicaffeoyl-5-succinoylquinic acid, 1,5-dicaffeoyl-4-fumaroylquinic acid, 1,3-dicaffeoyl-5-fumaroylquinic acid, 1,4-dicaffeoyl-3-maloylquinic acid, 1,5-dicaffeoyl-3-maloylquinic acid and 1,5-dicaffeoyl-4-maloylquinic acid, 1,3,5-tricaffeoyl-4-succinoylquinic acid, 1,5-dicaffeoyl-3,4-disuccinoylquinic acid, 1,5-dicaffeoyl-3-fumaroyl-4-succinoylquinic acid, 1-fumaroyl-3,5-dicaffeoyl-4-succinoylquinic acid, 1,5-dicaffeoyl-3-succinoyl-4-dimaloylquinic acid and dicaffeoyldimaloylquinic acid. Further, Liu et al. (2012) isolated and identified 12 caffeoylquinic acids in both seeds and roots of *A. lappa*. The identified compounds included chlorogenic acid, 1,5-di-*O*-caffeoylquinic acid, 1,3-di-*O*-caffeoylquinic acid, dicaffeoyl-maloylquinic acid, dicaffeoyl-maloylquinic acid, 1,3-di-*O*-caffeoylquinic acid, 1,5-di-*O*-caffeoyl-3-*O*-maloylquinic acid, 1,5-di-*O*-caffeoyl-3-*O*-succinoylquinic acid, 1,5-di-*O*-caffeoyl-4-*O*-maloylquinic acid, dicaffeoyl-dimaloylquinic acid, 1,5-di-*O*-caffeoylquinic acid, 1,5-di-*O*-caffeoyl-3-*O*-succinoyl-4-*O*-maloylquinic acid, 1,5-di-*O*-caffeoyl-3,4-di-*O*-succinoylquinic acid, and 1,3,5-tri-*O*-caffeoyl-4-*O*-succinoylquinic acid. In addition, phytochemical analysis of root extracts of *A. lappa* showed the occurrence of 8 additional isomers of hydroxycinnamic acids (Liu et al., 2012; Tusch et al., 2014). An average content of chlorogenic acid, 1-*O*-,5-*O*-dicaffeoylquinic acid and 1,5-dicaffeoyl-3-succinylquinic acid was observed to be between 1.7 and 7.9 mg/g dry weight of roots (Wang et al., 2001). Two new neolignan glucosides named (7*OS*, 8*OR*, 8*S*)-4,40,90-trihydroxy-3,30-dimethoxy-7*O*,9-epoxylignan-7-oxo-4-*O*- β -D-glucopyranosyl-40-*O*- β -D-glucopyranoside and (7*S*, 8*R*)-4,7,9,90-tetrahydroxy-3,30-dimethoxyl-70-oxo-8-40-oxyneolignan-4-*O*- β -D-glucopyranoside were determined from

the fruit extract of *A. lappa* (Yang et al., 2012). The occurrence of phenolic acids, caffeic acid, cynarin and chlorogenic acid has been reported for the first time in *A. lappa* seeds and leaves (Ferracane et al., 2010; Tang et al., 2014; Lou et al., 2016) and later in both seeds and roots (Predes et al., 2011). Chlorogenic acid was determined by Tardio et al. (2005) in the seeds of *A. lappa*. Further, UPLC analysis revealed the presence of caffeic acid, benzoic acid and *p*-coumaric acid in the leaves of *A. lappa* (Tardio et al., 2005; Lou et al., 2010a,b). From the ethyl acetate and *n*-butanol fractions of *A. lappa*, 1,5-*O*-two caffeoylquinic acids, succinic acid and 5-hydroxy maltol were identified for the first time (Han et al., 2013). Likewise, HPLC and UPLC with photodiode array (PDA) detector were used to quantify caffeoyl esters, chlorogenic acid and 1,5-dicaffeoylquinic acid in aerial parts and root samples of *A. lappa* (Haghi et al., 2013). Two more phenolic compounds, namely, coumaroylquinic acid and caffeoyl-hexose-hydroxyphenol were identified by Rajasekharan et al. (2015) in the root extracts of *A. lappa* (Rajasekharan et al., 2015). It was reported that caffeoylquinic acids and their derivatives show multiple pharmacological activities including decrease in diet-induced obesity *via* modulation of PPAR α and LXR α transcription (Huang K. et al., 2015) and anti-ulcerogenic effect (Lee et al., 2010).

Flavonoids

The reported flavonoids include flavonols, flavones, and their glycosides. Two major constituents, namely rutin and isoquercetin, along with few other minor flavonoids including kaempferol-3-*O*-rhamnoglucoside, quercimeritrin and astragalin were identified in the ethanolic extracts of *A. minus* leaves (Saleh and Bohm, 1971). Likewise, the occurrence of quercetin-3-*O*-rhamnoside was reported from the leaves of *A. lappa*. Later, the presence of phenolic compounds such as quercetin, quercitrin, rutin, and luteolin have been reported in seeds, fruits, leaves and roots of *A. lappa* (Saleh and Bohm, 1971; Tamayo et al., 2000; Yu et al., 2003; Lou et al., 2010a,b, 2016; Predes et al., 2011; Liu et al., 2012; Tang et al., 2014). Also, few isoflavone derivatives including genistein, nobiletin, biachanin A and tangeretin have been detected in *A. lappa* roots (Eberding et al., 2007). A comparative study has shown the existence of chemical differences within the *A. lappa* organs (Ferracane et al., 2010). According to them, luteolin and quercetin rhamnoside were detected in roots whereas rutin, quercetin, quercitrin and luteolin in leaves. On the other hand, no flavonoids were found in the seeds of *A. lappa*. Two more flavonols, namely quercetin 3-*O*-glucuronide and quercetin 3-vicianoside, were identified by Rajasekharan et al. (2015) in the root extracts of *A. lappa*.

Terpenoids

The fruits of *A. lappa* were found to contain β -eudesmol, a sesquiterpene alcohol (Rajasekharan et al., 2015; Yang et al., 2015). Pentacyclic triterpenoids, such as ursolic and oleanolic acids were detected by Han et al. in the ethanolic extract of *A. lappa* roots (Han et al., 2013). Arctiopicroin and onopordopicroin are the sesquiterpene lactones isolated from the leaf extract of *A. lappa* (Barbosa et al., 1993; Machado et al., 2012). Arctiopicroin occurrence is also evidenced in *A. lappa*. Later,

few more sesquiterpene lactones, namely dehydromelitensin-8-(4'-hydroxymethacrylate), dehydromelitensin, and melitensin and a norisoprenoid along with 2 more terpenes such as dehydrovomifoliol and lolilide were identified in *A. lappa* leaf (Machado et al., 2012). Onopordopicroin, a germacranolide sesquiterpene lactone was isolated from the aerial parts of *A. nemorosum* (Zimmermann et al., 2012). Two triterpenoids, namely 3 α -acetoxy-hop-22(29)-ene and 3 α -hydroxylanosta-5, 15-diene were isolated from the leaves of *A. lappa* (Jaiswal and Kuhnert, 2011).

Others

From the concentrated sap obtained from *A. lappa* roots (*A. lappa* and *A. tomentosum*), β -asparagine was isolated for the first time (Boldizar et al., 2010). The carotenoid crocin was reported to occur in the leaves of *A. lappa* (Lou et al., 2016).

Volatile Compounds

A total of 101 volatile chemical constituents were identified in *A. lappa*. The details of these compounds are partially summarized in Table 2 and described in this section. Carboxylic acids and fatty acids were more prevalent in *A. lappa*. On the other hand, there are no available literatures on the identification of volatile components in other *Arctium* species. The chemical structures of some compounds from *Arctium* species are shown in Figure 2.

Hydrocarbons

Fourteen hydrocarbon compounds, aploaxene, clovene, dihydroaploaxene, eicosane, 1-heptadecene, heptacosane, hexacosane, nonadecane, 2-naphthalenemethanol, 1-pentadecene, pentacosane, pentadecane, tetracosane, and tetradecane were detected from the roots, seeds, and leaves of *A. lappa* (Washino et al., 1986; Wang et al., 2004). In addition, docosane, eicosane, heptacosane, hexacosane, tetracosane, and pentadecane were found only in roots and leaves. Docosane was found only in leaves, while seeds of *A. lappa* contained only eicosane.

Aldehydes

Nineteen aldehydes, namely, benzaldehyde, butanal, decanal, dodecanal, heptanal, hexanal, (*Z*)-3-hexenal, (*E*)-2-hexenal, 2-methylpropanal, 3-methylbutanal, nonanal, octanal, (*E*)-2-octanal, phenylacetaldehyde, pentanal, propanal, tridecanal, 4-methoxybenzaldehyde, and undecanal were found as root volatile compounds in *A. lappa* (Washino et al., 1986, 1987; Wang et al., 2004). Interestingly, only the alkyl aldehyde nonanal was present in all plant parts such as roots, leaves, and seeds (Washino et al., 1986, 1987).

Methoxypyrazines

Seven methoxypyrazines, such as 2-methoxy-3-methylpyrazine, 2-methoxy-3-propylpyrazine, 2-isopropyl-3-methoxy-3-methylpyrazine, 2-sec-butyl-3-methoxypyrazine, 2-butyl-3-methoxypyrazine, 2-isobutyl-3-methoxypyrazine, and 2-isoamyl-3-methoxypyrazine were detected in roots of *A. lappa* (Washino et al., 1986, 1987).

TABLE 2 | Volatile constituents of *Arctium* spp.

| S. No | Compound name | Formula | Species | Plant origin/part | Analytical method | References |
|------------------------------|--------------------------------|---|-----------------|--------------------|-------------------|--|
| Hydrocarbons | | | | | | |
| 1 | Apotaxene | C ₁₇ H ₂₈ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 2 | Clovene | C ₁₅ H ₂₄ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 3 | Dihydroaplotaxene | C ₁₇ H ₃₀ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 4 | Docosane | C ₂₂ H ₄₆ | <i>A. lappa</i> | Leaves | GCMS | Aboutabl et al., 2013 |
| 5 | Eicosane | C ₂₀ H ₄₂ | <i>A. lappa</i> | Roots/leaves/seeds | GCMS | Aboutabl et al., 2013 |
| 6 | 1-Heptadecene | C ₁₇ H ₃₄ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 7 | Heptacosane | C ₂₇ H ₅₆ | <i>A. lappa</i> | Roots/leaves | GCMS | Aboutabl et al., 2013 |
| 8 | Hexacosane | C ₂₆ H ₅₄ | <i>A. lappa</i> | Roots/leaves | GCMS | Aboutabl et al., 2013 |
| 9 | Nonadecane | C ₁₉ H ₄₀ | <i>A. lappa</i> | Leaves | GCMS | Aboutabl et al., 2013 |
| 10 | 2-Naphthalenemethanol | C ₁₁ H ₁₀ O | <i>A. lappa</i> | Roots | GCMS | Wang et al., 2004 |
| 11 | 1-Pentadecene | C ₁₅ H ₃₀ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 12 | Pentacosane | C ₂₅ H ₅₂ | <i>A. lappa</i> | Roots | GCMS | Aboutabl et al., 2013 |
| 13 | Pentadecane | C ₁₅ H ₃₂ | <i>A. lappa</i> | Roots/leaves | GCMS | Aboutabl et al., 2013 |
| 14 | Tetracosane | C ₂₄ H ₅₀ | <i>A. lappa</i> | Roots/leaves | GCMS | Aboutabl et al., 2013 |
| 15 | Tetradecane | C ₁₄ H ₃₀ | <i>A. lappa</i> | Leaves | GCMS | Aboutabl et al., 2013 |
| Aldehydes | | | | | | |
| 16 | Benzaldehyde | C ₇ H ₆ O | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987; Wang et al., 2004 |
| 17 | Butanal | C ₄ H ₈ O | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 18 | Decanal | C ₁₀ H ₂₀ O | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 19 | Dodecanal | C ₁₂ H ₂₄ O | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 20 | Heptanal | C ₇ H ₁₄ O | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 21 | Hexanal | C ₆ H ₁₂ O | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 22 | (Z)-3-Hexenal | C ₆ H ₁₀ O | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 23 | (E)-2-Hexenal | C ₆ H ₁₀ O | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 24 | 2-Methylpropanal | C ₄ H ₈ O | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 25 | 3-Methylbutanal | C ₅ H ₁₀ O | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 26 | Nonanal | C ₉ H ₁₈ O | <i>A. lappa</i> | Roots/leaves/seeds | GCMS | Washino et al., 1986, 1987 |
| 27 | Octanal | C ₈ H ₁₆ O | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 28 | (E)-2-Octanal | C ₈ H ₁₄ O | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 29 | Phenylacetaldehyde | C ₈ H ₈ O | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 30 | Pentanal | C ₅ H ₁₀ O | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 31 | Propanal | C ₃ H ₆ O | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 32 | Tridecanal | C ₁₃ H ₂₆ O | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 33 | 4-Methoxybenzaldehyde | C ₈ H ₈ O ₂ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 34 | Undecanal | C ₁₁ H ₂₂ O | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| Methoxypyrazines | | | | | | |
| 35 | 2-Methoxy-3-methylpyrazine | C ₆ H ₈ N ₂ O | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 36 | 2-Isopropyl- 3-methoxypyrazine | C ₈ H ₁₂ N ₂ O | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 37 | 2-Methoxy-3- propylpyrazine | C ₈ H ₁₂ N ₂ O | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 38 | 2-sec-Butyl-3-methoxypyrazine | C ₉ H ₁₄ N ₂ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 39 | 2-Isobutyl-3-methoxypyrazine | C ₉ H ₁₄ N ₂ O | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 40 | 2-Butyl-3- methoxypyrazine | C ₉ H ₁₄ N ₂ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 41 | 2-Isoamyl-3-methoxypyrazine | C ₉ H ₁₄ N ₂ O | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| Fatty acids/Carboxylic acids | | | | | | |
| 42 | Acetic acid | CH ₃ COOH | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 43 | Benzoic acid | C ₇ H ₆ O ₂ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 44 | Butyric acid | C ₄ H ₈ O ₂ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 45 | Cinnamic acid | C ₉ H ₈ O ₂ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 46 | Costic acid | C ₁₅ H ₂₂ O ₂ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 47 | Decanoic acid | C ₁₀ H ₂₀ O ₂ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 48 | Dodecanoic acid | C ₁₂ H ₂₄ O ₂ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |

(Continued)

TABLE 2 | Continued

| S. No | Compound name | Formula | Species | Plant origin/part | Analytical method | References |
|----------------------------|--------------------------------|--|-----------------|--------------------|-------------------|---|
| 49 | Ethyl oleate | C ₂₀ H ₃₈ O ₂ | <i>A. lappa</i> | Seeds | GCMS | Aboutabl et al., 2013 |
| 50 | Hexanoic acid | C ₆ H ₁₂ O ₂ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 51 | Hexadecanoic acid | C ₁₇ H ₃₄ O ₂ | <i>A. lappa</i> | Roots/seeds | GCMS | Washino et al., 1986, 1987; Aboutabl et al., 2013 |
| 52 | (<i>E</i>)-3-Hexenoic acid | C ₆ H ₁₀ O ₂ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 53 | Heptanoic acid | C ₇ H ₁₄ O ₂ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 54 | (<i>E</i>)-3-Heptenoic acid | C ₇ H ₁₂ O ₂ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986; 1987 |
| 55 | Linoleic acid | C ₁₈ H ₃₂ O ₂ | <i>A. lappa</i> | Roots | GCMS | Wang et al., 2004 |
| 56 | 2, 3-Hydroxyoctanoic acid | C ₈ H ₁₆ O ₃ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 57 | 2-Methylpropionic acid | C ₄ H ₈ O ₂ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 58 | 2-Methylbutyric acid | C ₅ H ₁₀ O ₂ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 59 | 3-Methoxybenzoic acid | C ₈ H ₈ O ₃ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 60 | Methyl palmitate | C ₁₇ H ₃₄ O ₂ | <i>A. lappa</i> | Roots/seeds | GCMS | Washino et al., 1986, 1987; Aboutabl et al., 2013 |
| 61 | Methyl linolenate | C ₁₉ H ₃₂ O ₂ | <i>A. lappa</i> | Roots | GCMS | Wang et al., 2004 |
| 62 | Methyl oleate | C ₁₉ H ₃₆ O ₂ | <i>A. lappa</i> | Seeds | GCMS | Aboutabl et al., 2013 |
| 63 | Nonanoic acid | C ₉ H ₁₈ O ₂ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 64 | Nonanedioic acid | C ₉ H ₁₆ O ₄ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986; 1987 |
| 65 | (<i>E</i>)-3-nonenoic acid | C ₉ H ₁₆ O ₂ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 66 | Octanoic acid | C ₈ H ₁₆ O ₂ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 67 | (<i>E</i>)-3-Octenoic acid | C ₈ H ₁₄ O ₂ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 68 | Octadecanoic acid | C ₁₈ H ₃₆ O ₂ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 69 | Octadecanoic acid methyl ester | C ₁₈ H ₃₆ O ₂ | <i>A. lappa</i> | Seeds | GCMS | Aboutabl et al., 2013 |
| 70 | Pentanoic acid | C ₅ H ₁₀ O ₂ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 71 | Phenylacetic acid | C ₈ H ₈ O ₂ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 72 | Phenylpropionic acid | C ₉ H ₁₀ O ₂ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 73 | Propionic acid | C ₃ H ₆ O ₂ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 74 | Pentadecanoic acid | C ₁₅ H ₃₀ O ₂ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 75 | Salicylic acid | C ₇ H ₆ O ₃ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 76 | Tridecanoic acid | C ₁₃ H ₂₆ O ₂ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 77 | Tetradecanoic acid | C ₁₄ H ₂₈ O ₂ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 78 | Undecanoic acid | C ₁₁ H ₂₂ O ₂ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| Terpenes/terpenoids | | | | | | |
| Monoterpenoids | | | | | | |
| 79 | Carvomenthone | C ₁₀ H ₁₈ O | <i>A. lappa</i> | Roots/leaves | GCMS | Aboutabl et al., 2013 |
| 80 | Geraniol | C ₁₀ H ₁₈ O | <i>A. lappa</i> | Seeds | GCMS | Aboutabl et al., 2013 |
| 81 | Linalool | C ₁₀ H ₁₈ O | <i>A. lappa</i> | Seeds | GCMS | Aboutabl et al., 2013 |
| 82 | Thymol | C ₁₀ H ₁₄ O | <i>A. lappa</i> | Seeds | GCMS | Aboutabl et al., 2013 |
| 83 | <i>Z</i> -citral | C ₁₀ H ₁₆ O | <i>A. lappa</i> | Seeds | GCMS | Aboutabl et al., 2013 |
| 84 | <i>E</i> -citral | C ₁₀ H ₁₆ O | <i>A. lappa</i> | Seeds | GCMS | Aboutabl et al., 2013 |
| Sesquiterpenoids | | | | | | |
| 85 | Dehydrocostus lactone | C ₁₅ H ₁₈ O ₂ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 86 | Dehydrodihydrocostus lactone | C ₁₅ H ₂₉ O ₂ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| Oxygenated sesquiterpenes | | | | | | |
| 87 | Caryophyllene oxide | C ₁₅ H ₂₄ O | <i>A. lappa</i> | Roots/leaves | GCMS | Aboutabl et al., 2013 |
| 88 | β-Costol | C ₁₅ H ₂₄ O | <i>A. lappa</i> | Roots | GCMS | Aboutabl et al., 2013 |
| Sesquiterpene Hydrocarbons | | | | | | |
| 89 | Aromadendrene | C ₁₅ H ₂₄ | - | Roots/seeds | GCMS | Aboutabl et al., 2013 |
| 90 | Caryophyllene | C ₁₅ H ₂₄ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 91 | γ-Cadinene | C ₁₅ H ₂₄ | - | Roots/leaves/seeds | GCMS | Aboutabl et al., 2013 |
| 92 | Cyperene | C ₁₅ H ₂₄ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 93 | β-Elementene | C ₁₅ H ₂₄ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987; Aboutabl et al., 2013 |

(Continued)

TABLE 2 | Continued

| S. No | Compound name | Formula | Species | Plant origin/part | Analytical method | References |
|-------|-----------------------------------|-----------------------------------|-----------------|--------------------|-------------------|----------------------------|
| 94 | <i>trans</i> - β -Farnesene | C ₁₅ H ₂₄ | <i>A. lappa</i> | Roots/leaves | GCMS | Aboutabl et al., 2013 |
| 95 | α -Guaiene | C ₁₅ H ₂₄ | <i>A. lappa</i> | Roots | GCMS | Washino et al., 1986, 1987 |
| 96 | Isoaromadendrene epoxide | C ₁₅ H ₂₄ O | – | Roots/leaves/seeds | GCMS | Aboutabl et al., 2013 |
| 97 | Limonene | C ₁₀ H ₁₆ | <i>A. lappa</i> | Leaves/seeds | GCMS | Washino et al., 1986, 1987 |
| 98 | α -Myrcene | C ₁₀ H ₁₆ | <i>A. lappa</i> | Seeds | GCMS | Aboutabl et al., 2013 |
| 99 | α -Pinene | C ₁₀ H ₁₆ | <i>A. lappa</i> | Roots/leaves | GCMS | Aboutabl et al., 2013 |
| 100 | Squalene | C ₃₀ H ₅₀ | <i>A. lappa</i> | Seeds | GCMS | Aboutabl et al., 2013 |
| | Sesquiterpene Alcohol | | | | | |
| 101 | β -Copaen-4 α -ol | C ₁₅ H ₂₄ O | – | Roots/leaves/seeds | GCMS | Aboutabl et al., 2013 |

Carboxylic Acids and Fatty Acids

Twenty-two carboxylic acids namely acetic acid, benzoic acid, butyric acid, cinnamic acid, costic acid, dodecanoic acid, hexanoic acid, (*E*)-3-hexenoic acid, heptanoic acid, (*E*)-3-heptenoic acid, 2, 3-hydroxyoctanoic acid, 2-methylpropionic acid, 2-methylbutyric acid, 3-methoxybenzoic acid, nonanoic acid, nonanedioic acid, pentanoic acid, phenylacetic acid, phenylpropionic acid, propionic acid, salicylic acid, and undecanoic acid were identified in *A. lappa* roots (Washino et al., 1986, 1987; Wang et al., 2004). Fatty acids such as decanoic acid, hexadecanoic acid, linoleic acid, octanoic acid, (*E*)-3-octenoic acid, octadecanoic acid, pentadecanoic acid, tridecanoic acid, and tetradecanoic acid were found in roots while ethyl oleate, methyl oleate, hexadecanoic acid, methyl palmitate, and octadecanoic acid methyl ester were identified in seeds of *A. lappa* (Washino et al., 1986, 1987; Wang et al., 2004).

Monoterpenes and Sesquiterpenes

Three alcoholic and one phenolic monoterpenoids (carvomenthone, geraniol, linalool, and thymol); 2 sesquiterpene lactones (dehydrocostus lactone and dehydrodihydrocostus lactone, isoaromadendrene epoxide); 2 oxygenated sesquiterpenes (caryophyllene oxide and β -costol) and 12 sesquiterpene hydrocarbons namely, aromadendrene, caryophyllene, γ -cadinene, cyperene, β -elemene, *trans*- β -farnesene, α -guaiene, limonene, myrcene, α -pinene, and squalene were identified in *A. lappa* (Washino et al., 1986, 1987; Wang et al., 2004). Geraniol, linalool, thymol, aromadendrene, γ -cadinene, isoaromadendrene epoxide, limonene, α -myrcene, and squalene were identified only in the seeds of *A. lappa*. α -Pinene, isoaromadendrene epoxide, γ -cadinene, carvomenthone, and caryophyllene oxide were found in the roots and leaves.

BIOACTIVITIES OF *Arctium* SPECIES

Arctium lappa is widely used as an ethno-medicinal plant especially in North America, Asia and Europe, and is applied to treat various diseases including diabetes, gout, rheumatism, and skin problems (Chan et al., 2011; Azizov et al., 2012). *A. lappa* roots have been used as a vegetable in Japanese (referred to as 'gobo') and Korean cuisine. Its root has been used to treat constipation, mercury poisoning,

upper respiratory infections, inflammation and oxidative stress in patients with knee osteoarthritis (Maghsoumi-Norouzabad et al., 2016), while the leaves were efficacious in healing burns, rashes, and applied in women with labor condition (Force, 2001; Lewis and Elvin-Lewis, 2003; Amish Burn Study et al., 2014). *A. lappa* has also been found for the treatment of alopecia (loss of hair) among adults (Amish Burn Study et al., 2014). In Western countries, burdock is used as a remedy for several ailments ranging from arthritis, chronic inflammation, and various skin problems (e.g., scaly skin conditions such as psoriasis and eczema) to cancer treatment (Wu et al., 2010; Amish Burn Study et al., 2014).

Studies on the biological activities of extracts of different parts of *A. lappa* and compounds isolated thereof, were carried out and revealed antipyretic, antimicrobial, diuretic, diaphoretic, hypoglycaemic, antioxidant, anti-inflammatory, anti-hepatotoxicity, antiulcer, antimutagenicity, and antitumour activities.

Anticancer Effects

Arctium lappa fruit has been used in traditional medicine, and it is popular for its various anticancer effects. Arctigenin (ATG), a natural lignan product extracted from the seeds of *Arctium lappa*, has been shown to have estrogenic properties, that reduced the risk of osteoporosis, heart disease, and menopausal symptoms (Maxwell et al., 2017). It was found to possess antitumor effect by modulating the protein kinase activation pathway and hence rendering the tumor cells susceptible to effects of the nutrient-deprived environment (Awale et al., 2006). Later on, ATG was shown to induce apoptosis (programmed cell death) of estrogen receptor-negative cancer cells (MDA-MB-231) through the ROS/p38 MAPK pathway and epigenetic regulation of Bcl-2 by upregulating trimethylation of histone H3K9 (Hsieh et al., 2014). It was reported that ATG was able to inhibit cell proliferation and may induce apoptosis and cell cycle arrest at the G0/G1 phase in glioma cells (Maimaitili et al., 2017). In more detail, it was found that ATG increased the expression levels of p21, retinoblastoma and p53 proteins, and significantly decreased the expression levels of cyclin D1 and CDK4 proteins (Maimaitili et al., 2017). Furthermore, ATG was able to induce apoptosis in glioma cells, coupled with increased expression levels of cleaved caspase-3 and the

pro-apoptotic BCL2-associated X protein (Maimaitili et al., 2017). ATG-induced apoptosis was significantly suppressed by the pretreatment of cells with Z-DEVD-FMK, a caspase-3 inhibitor (Maimaitili et al., 2017). More recently, study by Lou et al. (2017) demonstrated ATG to significantly inhibit *in vitro* migration and invasion of human breast cancer cells (MDA-MB-231) by downregulation of MMP-2, MMP-9 and heparanase (Lou et al., 2017).

Extracts from *A. lappa* also showed selective antiproliferative activity against certain human cancer cell lines including K562, MCF-7 and 786-0 (Predes et al., 2011). Lappaol F, a novel natural product isolated from the seeds of *A. lappa*, was found to suppress cancer cell growth in a dose-dependent manner in various human cancer cell lines through induction of G1 and G2 cell-cycle arrest. This effect was associated with strong induction of p21 and p27 and suppression of cyclin-dependent kinase 1 (CDK1) and cyclin B1 (Sun et al., 2014).

A. lappa is one of the herbs widely used by cancer patients in some Canadian populations to improve quality of life (QOL) and prevent cancer progression. *A. lappa* is one of the herbs constituting the two proprietary herbal products: Flor-Essence® and Essiac® suggested for prolong survival and the improvement of QOL among cancer patients (Tamayo et al., 2000).

Antidiabetic Effects

Root of *A. lappa* root has been found to mediate hypoglycemic activities making it a popular choice to be used as a traditional medicine in diabetes. Oral administration of burdock root ethanolic extract in streptozotocin-induced diabetic rats significantly lowered blood glucose and increased insulin level in the diabetic rats compared to the control diabetic group (Cao et al., 2012). Additionally, treatment with *A. lappa* extract also reduced the levels of serum total cholesterol (TC), triglycerides (TG) and low density lipoprotein (LDL), whereas high density lipoprotein (HDL) level was higher in the control rats. More recently in a similar study, Ahangarpour et al. (2017) investigated the antidiabetic and hypolipidemic properties of the root extract of *A. lappa* on nicotinamide-streptozotocin (NA-STZ)-induced type 2 diabetes in mice (Ahangarpour et al., 2017). The results show that root extract of *A. lappa* displays anti-diabetic effect at certain doses. It exerts its effects through hypolipidemic and insulinotropic properties and hence the root extract could serve successfully in treating patients with type 2 diabetes in the future. Moreover, sitosterol- β -D-glucopyranoside from burdock's root acts as a potent inhibitor of alpha-glucosidases, thereby having the potential to reduce glycogenolysis and help to decrease blood glucose level (Tousch et al., 2014). In addition, Zhao and Zhou (2015) demonstrated that trace elements (e.g., Na, K, Mn, Fe, and Mg) present in the root and fruit extracts of *A. lappa* exhibit antidiabetic effects. While *A. lappa* constituents do reduce absorption of glucose, they also elevate inulin content in blood and slow digestion of carbohydrates to confer its anti-diabetic activities. The pharmacological mechanisms of *A. lappa* roots are slightly different from other classes of oral antihyperglycemic agents such as metformin. Metformin decreases hepatic glucose production, decreases intestinal absorption of glucose, and

improves insulin sensitivity by increasing peripheral glucose uptake and utilization (Dumitrescu et al., 2015).

Anti-oxidant, Hepatoprotective and Gastroprotective Activities

It is believed that lignans and caffeoylquinic acids from *A. lappa* are of value because of their antioxidant capacity (Maruta et al., 1995; Mkrtchian et al., 1998; Jaiswal and Kuhnert, 2011) by which they can scavenge free radicals that are thought to play an important role in many diseases.

The hydroalcoholic extracts of burdock roots possess significant antioxidant potential as seen by the application of various assays. Very recently, Fierascu et al. (2018) quantified antioxidant potential of burdock extracts using DPPH (2,2-diphenyl-1-picrylhydrazyl) and phosphomolybdate assays to demonstrate that burdock extracts have very high antioxidative activities, presumably due to the high content of polyphenols (Fierascu et al., 2018). The potent antioxidative property makes these extracts effective inhibitors of lipid peroxidation in rat liver homogenate *in vitro* (Duh, 1998) and an excellent hepatoprotective agent *in vivo* and *in vitro* (Lin et al., 2000). Due to its radical scavenging ability, *A. lappa* is also used to treat gastrointestinal ulcers (da Silva et al., 2013).

Antimicrobial Effects

Extracts of different parts of *A. lappa* have been investigated for their microbial-modulatory properties by many researchers. An organic extract from *A. lappa* has shown inhibiting properties toward the growth of *Pseudomonas aeruginosa*, *Escherichia coli*, *Lactobacillus acidophilus*, *Streptococcus mutans*, and *Candida albicans* residing in the teeth of the oral cavity (Gentil et al., 2006). Pereira et al. (2005) further reported potent growth inhibiting activities of *A. lappa* extract against a broad spectrum of oral microorganisms, specifically those associated with teeth infections, namely *Enterococcus faecalis*, *Staphylococcus aureus*, *Pseudomonas aeruginosa*, *Bacillus subtilis*, and *Candida albicans* (Pereira et al., 2005). Very recently, Fierascu et al. (2018) investigated antifungal potential of hydroalcoholic extract of burdock roots and observed that it is active against the fungal lines *Aspergillus niger* ATCC 15475 and *Penicillium hirsutum* ATCC 52323 (Fierascu et al., 2018). Fruit extract of *A. lappa* was tested (Dias et al., 2017) for *in vitro* antiviral properties against *Herpes simplex virus* type-1 (HSV-1) and was found to decrease viral load significantly at all concentrations tested (400, 50, and 3.125 μ g/mL). At 400 μ g/mL concentration, it showed comparable antiviral activity as acyclovir (50 μ g/mL). Arctigenin, one of the key constituents of *A. lappa* extract, has shown potent activities against human immunodeficiency virus type-1 (HIV-1) both *in vivo* and *in vitro* presumably by increasing the expression of Heme oxygenase-2 (HO-2) and blocking HIV-1gag proteins (Schroder et al., 1990).

Anti-inflammatory Effects

Various parts of *A. lappa* demonstrated anti-inflammatory effects (Liu et al., 2005). Burdock extract is known to

alleviate wound irritation and swelling and therefore has been traditionally used for healing burn wounds. This effect might be mediated through the inhibition of the cyclooxygenase-2 (COX-2) enzyme. Cyclooxygenase is a lipid metabolizing enzyme that catalyzes the oxygenation of polyunsaturated fatty acids. This process forms prostanoids, specifically eicosanoids, which are known to be potent cell signaling molecules connected to inflammatory processes (Charlier and Michaux, 2003). Phenolic compounds present in burdock extract (e.g., arctigenin, lappaol F, diartigenin) are inhibitors of this enzyme (Zhao et al., 2009; Lee and Kim, 2010), thereby suppressing lipopolysaccharide (LPS)-stimulated NO production (Park et al., 2007) and pro-inflammatory cytokines secretion (including TNF- α and IL-6) in a dose-dependent manner (Zhao et al., 2009; Kwon et al., 2016). Arctigenin also strongly inhibited the expression of iNOS (Inducible Nitric Oxide Synthase) and its enzymatic activity (Wang et al., 2007; Zhao et al., 2009). Moreover, it induced endothelial nitric oxide synthase (eNOS) and suppressed in a rat model subarachnoid hemorrhage-induced vasospasm by regulation of the PI3K/Akt signaling pathway (Chang et al., 2015). Among the studied phenolic compounds, diartigenin was found to inhibit the DNA binding ability of NF- κ B and to inhibit NF- κ B-regulated iNOS expression (Kim et al., 2008), thereby overall targeting NF- κ B-activating signaling cascade directly to confer anti-inflammatory response. Luteolin, an important flavonoid from burdock was also reported to possess significant anti-inflammatory properties (Ferracane et al., 2010; Nabavi et al., 2015).

Effects Against Skin Conditions

Leaves of *Arctium* species have been used in traditional medicinal practices in various skin conditions (e.g., rashes, boils, eczema, ichthyosis, acne, psoriasis, and abscesses) presumably due to the presence of various phenolic compounds. The potent antioxidant and anti-inflammatory properties of these compounds serve to detoxify and mediate healing action (Chan et al., 2011). Several hydroxycinnamic acids which are among the active phytochemicals in the *A. lappa* extracts (Liu et al., 2012; Tousch et al., 2014) have been found to act as free radical scavengers and possess antioxidant activities, which confer them potential to serve as skin protectors and wound healers (Graf, 1992; Phan et al., 2001; Taofiq et al., 2017). In addition, hydroxycinnamic acid derivatives also display anti-collagenase, anti-inflammatory, antimicrobial and anti-tyrosinase activities, as well as ultraviolet (UV) protective effects, suggesting that they can be exploited as anti-aging and anti-inflammatory agents, preservatives and hyperpigmentation-correcting ingredients (Ahangarpour et al., 2017). These bioactivities are the reason why burdock extracts find their use in various commercial cosmetic products.

Effect on Potency and Fertility

Diabetes mellitus induces many complications among which dysfunctions male reproductive system is worth mentioning. Glucose metabolism plays an important regulatory role

on the production or development of mature spermatozoa (spermatogenesis) as well as on maintaining specific functions, such as motility and fertilization ability in mature sperm cells. Therefore, it is not surprising that *A. lappa* root extract, which has hypoglycemic and antioxidative properties, would have beneficial effects on male potency and fertility. Ahangarpour et al. (2015) investigated the effect of *A. lappa* root extract on gonadotropin, testosterone, and sperm parameters in nicotinamide/streptozotocin-induced diabetic mice (Ahangarpour et al., 2015). The root extract led to increased level of luteinizing hormone (LH), follicle stimulating hormone (FSH), and testosterone as well as enhancement in sperm viability only in diabetic mice compared with the control group, indicating *A. lappa* root extract to be a potentially effective treatment for male sterility arising from diabetic conditions.

Effect on NO Production

It was reported that arctigenin inhibited NO release by IFN- γ signal, whereas it significantly enhanced lipopolysaccharide-triggered NO production in RAW264.7 cells, suggesting that arctigenin may regulate immune responses in activated macrophages and lymphocytes including TNF- α and NO production and lymphocyte proliferation (Cho et al., 1999). Another study shows that arctigenin suppressed the overproduction of NO through down-regulation of iNOS expression and iNOS enzymatic activity in LPS-stimulated macrophage (Zhao et al., 2009). Besides, lappaol F and diartigenin from *Arctium lappa* were shown to significantly inhibit NO production in the LPS-stimulated RAW264.7 cells with IC₅₀ values of 9.5 and 9.6 μ M, respectively (Park et al., 2007).

Safety Considerations on *Arctium* Species

Several adverse effects have been reported in literature stemming from long-term use of *A. lappa*. For example, contact dermatitis might develop after several days of applying a burdock root plaster to a wound, or even as fast as within 12 h in some cases (Rodriguez et al., 1995). In one instance, anticholinergic poisoning has been reported upon oral consumption of *A. lappa* extract (Force, 2001). However, this poisoning later turned out to be caused by products that have been contaminated with root of belladonna (deadly nightshade). The latter herb contains the poisonous chemical atropine. Long-term consumption of burdock also has led to anaphylaxis in one case (Chan et al., 2011). Root oil made from *A. lappa* was also found to cause unfavorable physiological effects such as redness, and anaphylactic shock (Rodriguez et al., 1995; Lewis and Elvin-Lewis, 2003; Sasaki et al., 2003). Caution is advised for pregnant or nursing women to consume burdock or its extract, as it might have detrimental effects on the fetus (Chan et al., 2011). Burdock can also interfere with blood clotting. People who are already on blood thinning medications are advised not take it without approval from their doctors. Even though burdock is considered a 'safe'

food, consuming it in large amounts should be avoided due to lack of large amount of safety studies on burdock. More *in vivo* studies are in particular needed on *A. lappa* to further evaluate its therapeutic potential and safe application window. Due to the presence of sesquiterpene lactones, the use of *Arctium* species should be avoided in patients with hypersensitivity to Asteraceae/Compositae (Chan et al., 2011).

SUMMARY

In summary, the volatile and non-volatile secondary metabolites present in different parts of *Arctium* species showed pharmacological potential in the treatment of various diseases. The literature existing on extracts of different parts of *A. lappa* and isolated compounds demonstrates antipyretic, antimicrobial, diuretic, diaphoretic, hypoglycaemic, antioxidant, anti-inflammatory, anti-hepatotoxicity, antiulcer, antimutagenicity, and antitumour activities. Hence, *Arctium* species display a broad therapeutic potential but further studies are needed on potential risks associated with their application.

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All authors listed have made a substantial, direct and intellectual contribution to the work, and approved it for publication.

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