- 1 Influence of activation conditions on textural
- 2 properties and performance of activated biochars
- 3 for pyrolysis vapors upgrading
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#### **ABSTRACT**

The main aim of the present study is to provide a comprehensive assessment of the effects of process activation conditions on the textural properties of the resulting activated carbons, which were produced from wheat straw-derived biochar through chemical activation (with  $K_2CO_3$  at different pressures and mass impregnation ratios) and physical activation (with  $CO_2$  at different temperatures and pressures). For chemically activated biochars, it was found that specific surface area and pore size distribution were both only positively affected by increasing the carbonate loading. However, physically activated biochars produced at the highest pressure and lowest temperature (1.0 MPa and 700 °C) had the highest surface areas and widest pore size distributions. The materials with the most appropriate textural properties were then tested as catalysts for steam and dry reforming of the aqueous phase of pyrolysis oil. The best catalytic performance (a total gas yield of 74% and a selectivity toward  $H_2$  of almost 40%) was observed for a physically activated biochar. This good performance was ascribed to the high availability of  $K^0$  on the catalyst surface, which could effectively promote the reactions involved in the upgrading process.

#### **Keywords**

- Biochar; chemical activation; K<sub>2</sub>CO<sub>3</sub>; physical activation; pressure; steam and dry reforming
- 31 of pyrolysis oil

#### 1. Introduction

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In recent years, the demand for activated carbons has increased due to their usefulness in a wide range of different applications such as gas and liquid purification [1], electrochemistry [2], soil remediation [3] and catalysis [4]. Thanks to their versatility to be adapted to specific applications, these materials are considered promising candidates to address environmental issues related to global warming and pollution [5]. Nevertheless, the current main drawback is that the production of activated carbons is still partly based on fossil fuels, which do not meet sustainability criteria [6,7]. Hence, much more efforts should be made to produce much more carbons from biomass. However, the direct conversion of biomass feedstock into the final product has a low yield. Thus, the production of activated carbons from biochar produced by slow pyrolysis seems to be more appropriate in terms of scalability, while at the same time these value-added materials can strengthen the value chain of existing biochar production systems. In general, pristine biochar has neither a well-developed surface area nor a hierarchical pore size distribution, which is mainly dominated by narrow pores with a diameter  $(d_p)$  lower than 0.7 nm (ultra-micropores). Hence, a subsequent activation step is required to improve the textural properties of the carbon material. The activation process involves the development and opening of the porosity of a char using an activation agent. Depending on the agent used, the procedure can be called chemical or physical activation. Chemical activation usually involves two steps. First, the biochar is impregnated with an aqueous solution of the chemical activation agent, or mixed with it in the dry state, and, in a second step, the blend is heated up to a given temperature at which oxidation, dehydration, aromatization and crosslinking reactions, among others, occur. Although the most widely used chemical reagents are KOH [8] and H<sub>3</sub>PO<sub>4</sub> [9], non-hazardous and relatively cheap alternative compounds such as K<sub>2</sub>CO<sub>3</sub> [10,11] have recently attracted considerable attention. Mai et al. [12] have recently reported that activation with potassium carbonate can result in carbons with a high percentage of structural defects and a well-balanced porosity between micro- and mesopores.

When activation is carried out by exposing the precursor to relatively high temperatures under an oxidizing atmosphere (e.g., CO<sub>2</sub> [13], H<sub>2</sub>O [14] and O<sub>2</sub> [15]), the process is called physical activation. Through either physical or chemical activation, it is possible to tune the textural features of the starting biochar by properly adjusting the activation conditions, mainly temperature, type and/or concentration of activation agent, as well as pressure. To the best of our knowledge, although both activation procedures are widely reported in the literature, a thorough study on the effects that the activation parameters and their possible interactions have on the textural properties of the resulting activated biochar, has never been reported so far. Such information could be very helpful to properly establish the most appropriate operating conditions to produce engineered carbon materials from biomass.

One of the most interesting fields of application of activated carbons is the upgrading of raw pyrolysis vapors, which contain both permanent gases (e.g., CO<sub>2</sub>, CO, CH<sub>4</sub>, and H<sub>2</sub>) and condensable compounds. The condensable fraction, which is generally called pyrolysis oil or bio-oil, is a mixture of hundreds of organic compounds (alcohols, ketones, acids, etc.) and water, and its composition strictly depends on the biomass composition and the pyrolysis operating conditions [16]. For biochar production systems based on slow pyrolysis, downstream processes aimed at upgrading pyrolysis vapors are required to avoid undesirable condensation of organic compounds and to increase simultaneously the quality of the gaseous product. For this purpose, combined steam and dry reforming of pyrolysis oil is a promising option [17,18], due to the presence of a relatively high amount of CO<sub>2</sub> and steam in the raw vapor phase. Heterogeneous catalysts based on transition metals such as Ni [19,20], Co [21], Pt and Rh [22] can be used to improve the overall conversion and the selectivity of the products. However, the main drawback of these catalysts is that the relatively expensive active phase can

easily be poisoned and/or deactivated by deposition of coke [23]. One possible solution to reduce the overall cost of the upgrading process is to use activated biochar as catalyst. Its porous structure and inherent inorganic contents (especially K, Mg and Ca) could result in a relatively good catalytic activity [24–29]. Furthermore, due to the presence in the reaction system of H<sub>2</sub>O and CO<sub>2</sub>, the carbonaceous support, as well as coke deposits, could be continuously partly gasified, thus creating new pores to avoid deactivation. In addition, the spent biochar can be burned to recover energy [30] or, depending on the metal loaded on the support, employed as soil conditioner [31].

Keeping in mind all the above, the main objective of this study was to perform a preliminary investigation about the effects of several activation conditions on the textural properties and the catalytic activity of the resulting activated biochars. To this end, wheat straw-derived biochar was activated chemically and physically with K<sub>2</sub>CO<sub>3</sub> and CO<sub>2</sub>, respectively, under different operating conditions. To objectively assess the effect of the selected operation conditions (temperature and mass ratio K<sub>2</sub>CO<sub>3</sub>/precursor for chemical activation, and temperature and pressure for CO<sub>2</sub> activation), we implemented two factorial designs of experiments, with two factors and three replicates at the center point. The specific surface areas and pore size distributions of the resulting activated biochars were then evaluated. The most promising activated biochars were finally tested as catalysts for steam and dry reforming of pyrolysis oil. Their performance was evaluated in terms of conversion of liquid into gas, product selectivity and resistance to deactivation.

# 2. Experimental Section

This study was divided into two main steps: in the first, we carried out physical and chemical activations under different process conditions in order to study their influence on the pore size distribution and specific surface area of the resulting activated biochars; in the second part, the most promising materials were then tested as catalysts in pyrolysis vapors

upgrading experiments. The methodology used in this study is summarized graphically inFig. A.1.

#### 2.1. Biochar production

The biochar used in this study was produced by slow pyrolysis of binder-free wheat straw pellets (9 mm OD and 10–13 mm long). Pyrolysis was carried out at atmospheric pressure in a fixed bed reactor, which was placed inside a furnace and heated up to 500 °C at an average heating rate of 5 °C min<sup>-1</sup> and using N<sub>2</sub> as carrier gas. More details on the pyrolysis device and the experimental procedure are available elsewhere [32]. The resulting biochar (i.e., "pristine") was ground and then thoroughly sieved to obtain particle sizes in the range of 0.212 to 1.41 mm. The pristine biochar was characterized by proximate analysis (performed in quadruplicate according to ASTM standards D3173 for moisture, D3174 for ash, and D3175 for volatile matter) and ultimate analysis by means of an elemental analyzer CHN628 from Leco Corporation (USA).

## 2.2. Activation of pristine biochar

#### 2.2.1. Chemical activation

For chemical activation, the pristine biochar was first impregnated with a 1 mol L<sup>-1</sup> aqueous solution of K<sub>2</sub>CO<sub>3</sub>. Three K<sub>2</sub>CO<sub>3</sub>: biochar mass impregnation ratios (1:1, 2:1 and 3:1) were achieved by adjusting the volume of solution. The heterogeneous mixture was then stirred for 2 h at 50 °C, filtered and dried overnight at 110 °C to remove the residual water. Afterwards, 10 g of the impregnated samples were heated up to 700 °C at a heating rate of 10 °C min<sup>-1</sup>, under an inert atmosphere (N<sub>2</sub>), and at three different values of absolute pressure (0.10, 0.55 and 1.00 MPa). For this purpose, a tubular fixed bed reactor (made of nickel-chromium alloy UNS N06600, 28.1 mm ID and 600 mm long) placed in a vertical furnace (model EVA 12/300 from Carbolite Gero, UK) was used. The relatively low activation temperature was chosen to avoid the evaporation of the metallic potassium derived from the decomposition of the

activation agent and, also, to ensure low activation extensions and clearly see the potential effects of the studied parameters. On the other hand, the range of activating pressures was chosen in order to allow this experimental setup to be easily scaled up, since the involved pressures were not too high (up to 1.0 MPa). A soaking time at the highest temperature of 60 min was set. The pressure within the reactor was adjusted using a downstream servo-controlled regulator valve. The gas hourly space velocity (GHSV) at the activation temperature was estimated to be  $7000 \, h^{-1}$ , considering the pressure applied and a bed void factor of 0.5. Thus, the mass flow rate of the inlet gas stream (N<sub>2</sub>) was properly adjusted as a function of the selected pressure and the highest temperature to achieve the aforementioned GHSV value.

As a last step, the carbons were rinsed to remove the unreacted reagent and other impurities from their surface. In this work, two different washing procedures were adopted to assess possible effects on the resulting surface area and the catalytic activity of the activated carbons produced. For this purpose, among all the unwashed chemically activated biochars, four of them were just washed with hot deionized water (100 °C), while the seven chemically activated biochars adopted in the design of experiments were washed with a 0.25 mol L<sup>-1</sup> solution of HCl followed by hot water. Both washing procedures were carried out until neutral pH. The resulting activated biochars were then dried overnight at 110 °C. The efficiency of the washing step was evaluated according to the washing yield, calculated from Eq. 1, in which  $m_c$  and  $m_w$  are the masses of activated carbon before and after the washing steps, respectively.

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$$Y_{wash} = \left(1 - \frac{m_c - m_w}{m_c}\right) 100 \tag{1}$$

2.2.2. Physical activation

Physically activated biochars were produced under an atmosphere of pure CO<sub>2</sub> at three different temperatures(700, 775 and 850 °C), and at three different absolute pressures (0.10, 0.55, and 1.00 MPa). The selected activation temperatures are within the range commonly reported in the literature, whereas the pressure values were established with the purpose to

compare the properties of resulting activated carbons produced under atmospheric and moderate pressures. Using the same device described in the previous section, 10 g of pristine biochar were heated under  $N_2$  atmosphere, at a heating rate of 10 °C min<sup>-1</sup>, until the target temperature was reached. Then, the gas supply was switched from  $N_2$  to  $CO_2$  at a constant GHSV of 7000 h<sup>-1</sup>. These conditions were maintained during the time required to reach a degree of burnout ( $\eta$ , defined as the percentage of mass loss) in the range of 30% to 60%.

## 2.2.3. Design of experiments

To objectively assess the effects of the activation conditions on the textural properties of activated biochars, an unreplicated two-level factorial design of experiments (with two factors and three replicates at the center point) was adopted for both chemical and physical activations. For chemical activation, the analyzed factors were the absolute pressure and the impregnation ratio of K<sub>2</sub>CO<sub>3</sub> to raw biochar, whereas in the case of physical activation, the temperature and the absolute pressure were the studied factors. The structure of the regression model used in the statistical analysis was the following:

$$\hat{\mathbf{y}} = \beta_0 + \beta_i A + \beta_i B + \beta_{ij} A B \tag{2}$$

where A and B corresponded to the assessed factors (normalized values in the range from -1 to 1), whereas  $\beta_0$ ,  $\beta_i$ , and  $\beta_{ij}$  were the intercept, linear, and interaction coefficients, respectively. The results obtained for the selected response variables (y) were analyzed using the Minitab v17 statistical package. A significance level of 5% was assumed and the adjusted coefficient of determination  $(R^2_{adj})$  was taken as an indicator of the quality of the fit. Table 1 summarizes the adopted designs of experiments and lists the names of the activated biochars produced. Briefly, activated biochar are referred as  $X_{-}Y_{-}P$ . X corresponds to the activation procedure (CB or PB for *chemically activated biochar* and *physically activated biochar*, respectively); Y is the activation temperature for PBs or the impregnation ratio for CBs; and P is the activation pressure.

#### 2.3. Catalytic pyrolysis vapors upgrading

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The liquid feed used here for pyrolysis vapors upgrading tests was the filtered aqueous phase of the pyrolysis oil formed during the production of pristine biochar derived from wheat straw. This resulting liquid product was characterized in terms of elemental composition, using the same CHN analyzer as mentioned in section 2.1, and water content, using a volumetric Karl-Fischer titrator Titrino plus 870 from Metrohm (Switzerland). A schematic overview of the experimental device used for the reforming tests is shown in Fig. A.2 (Appendix A). The reactor and furnace elements were the same as those previously described in section 2.2. The reforming operating conditions were selected based on the findings of a previous study [33], which aimed to establish the best process conditions to minimize deactivation by coke deposition. Briefly, activated biochar (2-5 g) was loaded into the reactor and heated under N<sub>2</sub> atmosphere at an absolute pressure slightly above 0.1 MPa. Once the bed temperature reached the target value of 750 °C, CO<sub>2</sub> was added to the inlet gas stream at a partial CO<sub>2</sub> pressure of 0.02 MPa. The liquid feed was then injected into the gas stream at the inlet of the reactor by means of a HPLC pump (model 521 from Analytical Scientific Instruments, USA). The liquid hourly space velocity (LHSV) was kept constant and equal to 2 h<sup>-1</sup> for all the experiments conducted. The duration of the experiments was 60 min. The reactor outlet stream, consisting of permanent gases, unreacted pyrolysis oil, and condensable side products, was forced to pass through an ice-bath condensation train. The permanent gases were analyzed using a dualchannel micro gas chromatograph (µ-GC 490 from Agilent, USA) equipped with TCD detectors and two analytical columns (a Molsieve 5 A and a PolarPlot U). The known amount of N<sub>2</sub> fed was used as a tracking compound to calculate the yield of produced gas.

**Table 1.** Matrix of the factorial designs adopted to assess the effects of the selected factors during both chemical and physical activations. For example, a physical activation carried out at A=-1 and B=1 means that the process was performed at 700°C and 1.0 MPa

		Chemical activation					
(thermal treatment under N <sub>2</sub> up to 700 °C and 1 h soaking time)							
		Factors	Response Variables				
	$\boldsymbol{A}$	B	$S_{2D ext{-}NLDFT}$ , $V_t$ , $V_{ultra}$ , $V_{micro}$ , $V_{meso}$				
Level	Pressure (MPa)	Mass ratio (K <sub>2</sub> CO <sub>3</sub> : raw biochar)					
Low (-1)	0.10	1:1					
Middle (0)	0.55	2:1					
High (+1)	1.00	3:1					
Runs			Activated biochar designation				
Kulis			(all washed with an acidic solution)				
1	0	0	CB_2_0.55				
2	-1	+1	CB_3_0.10				
3	+1	+1	CB_3_1.00				
4	0	0	CB_2_0.55				
5	0	0	CB_2_0.55				
6	-1	-1	CB_1_0.10				
7	+1	-1	CB_1_1.00				
		Physical activation					
		Factors	Response Variables				
	$\boldsymbol{A}$	B	$S_{2D ext{-}NLDFT}$ , $V_t$ , $V_{ultra}$ , $V_{micro}$ , $V_{meso}$				
Level	Temperature (°C)	Pressure (MPa)					
Low (-1)	700	0.10					
Middle (0)	775	0.55					
High (+1)	850	1.00					
Runs			Activated biochar designation				
1	-1	-1	PB_700_0.10				
2	-1	+1	PB_700_1.00				
3	+1	+1	PB_850_1.00				
4	0	0	PB_775_0.55				
5	0	0	PB_775_0.55				
6	+1	-1	PB_850_0.10				
7	0	0	PB_775_0.55				

The performance of the activated biochars tested was measured in terms of total gas yield  $(Y_g)$  and hydrogen yield  $(Y_{H2})$ , as defined by Eqs. (3) and (4), respectively.

$$Y_g = \frac{m_g}{m_L} 100 (3)$$

$$Y_{H2} = \frac{F_{H2 \text{ av}}}{F_{H2 \text{stoi}}} 100 \tag{4}$$

$$F_{\text{H2stoi}} = \left(2n + \frac{m}{2} - k\right) F_{\text{BO}} \tag{5}$$

$$S_i = \frac{n_i}{n_{\text{tot}}} 100 \tag{6}$$

In Eq. (3),  $m_g$  was the cumulative mass of the gas produced during the 60-min experiments, whereas  $m_L$  corresponded to the total mass of the liquid fed. In Eq. (4),  $F_{H2av}$  was the experimental average molar flow rate of hydrogen from minute 20 to minute 40.  $F_{H2stoi}$ , which was calculated according to Eq. (5), corresponded to the stoichiometric molar flow rate of hydrogen considering the contribution from steam reforming reaction.  $F_{BO}$  was the molar flow rate of the dry pyrolysis oil fed. The selectivity towards specific gaseous products ( $S_i$ ) was calculated according to Eq. (6), in which  $n_{tot}$  was the sum of the gaseous products in moles at the outlet stream.

## 2.4. Characterization of carbon materials

The textural properties of pristine and activated biochars were determined from both  $N_2$  and  $CO_2$  adsorption isotherms at -196 °C and 0 °C, respectively. Around 120 mg of sample were degassed under vacuum at 150 °C. ASAP 2020 and ASAP 2420 automatic adsorption analyzers (Micromeritics, USA) were used. The results obtained from the isotherms were treated using the MicroActive software. We determined the BET area ( $A_{BET}$ ) by application of the BET model, as well as the Gurvitch volume ( $V_{0.97}$ ). The enhanced 2D-NLDFT model [34] was employed using SAIEUS software (available at <a href="www.nldft.com">www.nldft.com</a>) to evaluate the pore size distributions (PSDs) and other related parameters: surface area ( $S_{2D-NLDFT}$ ), ultra-micropore

volume ( $V_{ultra}$ ,  $d_p < 0.7$  nm), micropore volume ( $V_{micro}$ ,  $d_p < 2$  nm), total pore volume ( $V_t$ ) and mesopore volume ( $V_{meso}$ ,  $d_p$  in the range of 2 to 50 nm). The latter was calculated as the difference between  $V_t$  and  $V_{micro}$ .

To evaluate the availability of residual potassium carbonate on the surface of chemically activated biochars, Fourier-transform Infrared Spectroscopy (FTIR) analyses were conducted using a Frontier MIR/NIR Spectrometer from PerkinElmer (USA).

Qualitative temperature-programmed desorption (TPD) analyses of pristine biochar and fresh catalysts used in reforming tests were also performed to assess the thermal stability of these samples. TPD measurements were carried out using a thermogravimetric analyzer coupled with a mass spectrometer (STA 449 F3 and QMS 403 Aëolos Quadro from Netzsch, Germany), by heating the sample up to 750 °C at a heating rate of 5 °C min<sup>-1</sup> under an Ar atmosphere.

A FEI XL30 SFEG scanning electron microscope coupled with energy-dispersive X-ray spectroscopy (Oxford Instrument EDS SDD XMAX detector) was used to characterize the surface of fresh and spent activated biochars. The secondary electron images were taken with an acceleration voltage of 3 or 5 kV to investigate the topology of the samples surface, while an acceleration tension of 10 kV was applied to carry out the chemical mapping of the materials by EDX analysis. Additionally, X-ray photoelectron spectroscopy (XPS) data of spent and fresh catalysts were recorded using an ESCAPlus OMICROM system equipped with a hemispherical electron energy analyzer, following the procedure detailed elsewhere [35].

#### 3. Results and discussion

The results of the textural characterization as well as the proximate and ultimate analyses of the pristine biochar are summarized in Table A.1. The reported values of  $A_{BET}$  and  $S_{2D-NLDFT}$  confirm that the porosity of the pristine biochar, as previously stated, was characterized by a very high fraction of ultra-micropores.

#### 260 3.1. Chemical activation

Although the use of  $K_2CO_3$  as catalyst in coal gasification processes has been widely reported in the literature [10–12,33], the mechanism explaining its interaction with the carbon matrix still remains unclear. The main reactions that probably occur during chemical activation of a carbonaceous material are the following:

$$K_2CO_3 \rightleftarrows K_2O + CO_2 \tag{7}$$

$$266 CO_2 + C \rightleftarrows 2CO (8)$$

$$267 K_2CO_3 + 2C \rightleftarrows 2K + 3CO (9)$$

$$268 K_2O + C \rightleftharpoons 2K + CO (10)$$

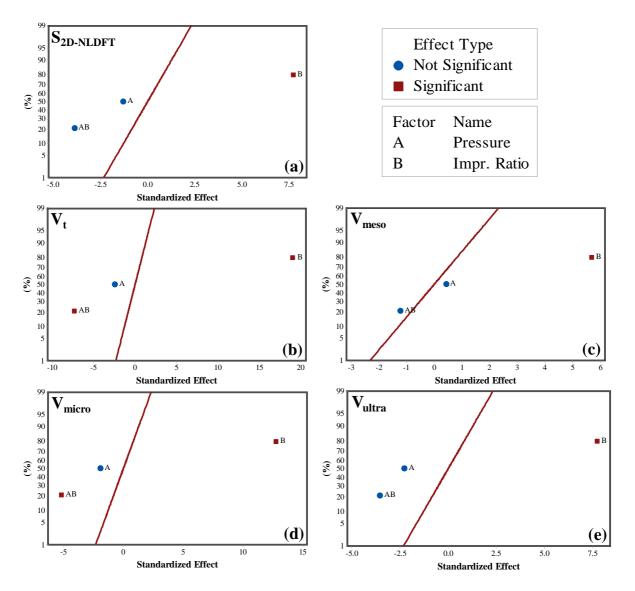
The increase in specific surface area could be ascribed to the CO<sub>2</sub> released from carbonate decomposition (Reaction 7), which can diffuse into the solid structure and subsequently react with carbon, thereby stimulating the production of CO by the reverse Boudouard reaction (Reaction 8) and creating vacancies in the solid structure [36]. Despite the fact that the decomposition of K<sub>2</sub>CO<sub>3</sub> mainly occurs at temperatures above 900 °C, its contact with the carbon matrix can promote its decomposition a relatively low temperatures [37,38]. The potassium oxide resulting from the decomposition of the carbonate, or the carbonate itself, can also react with the carbon-based material through Reaction 9 and 10 towards CO and metallic potassium [39].

The assessed textural properties of chemically activated biochars are listed in Table 2. Fig. 1 shows the normalized plots of the standardized effects for the response variables considered:  $S_{2D-NLDFT}$ ,  $V_t$ ,  $V_{ultra}$ ,  $V_{micro}$ , and  $V_{meso}$ . The PSDs obtained for biochars activated with different impregnation ratios and the  $N_2$  adsorption isotherms of such samples are shown in Figs. A.3a and A.4, respectively. More detailed statistical outcomes for the results reported in the present section can be found in Table A.2.

Table 2. Results of textural characterization and washing efficiency of chemically activated biochars

		ent specific face area			Washing yield			
Material	$A_{BET}$	$S_{2D ext{-}NLDFT}$	$V_{0.97}$	$V_t$	$V_{ultra}$	$V_{micro}$	$V_{meso}$	$Y_{wash}$
	r	$n^2 g^{-1}$		$\mathrm{cm^3~g^{-1}}$				%
CB_1_0.10	170	712	0.078	0.181	0.143	0.175	0.006 (3.19 %) <sup>a</sup>	77.0
CB_1_0.10_W <sup>b</sup>	142	700	0.068	0.180	0.135	0.169	0.011 (6.14 %) <sup>a</sup>	77.1
CB_1_1.00	180	761	0.081	0.195	0.149	0.188	0.007 (3.82%) <sup>a</sup>	76.4
CB_1_1.00_W <sup>b</sup>	100	700	0.049	0.168	0.144	0.160	0.008 (5.01%) <sup>a</sup>	78.1
CB_3_0.10	413	933	0.181	0.259	0.197	0.246	0.013 (4.96%) <sup>a</sup>	54.3
CB_3_0.10_W <sup>b</sup>	368	893	0.158	0.242	0.193	0.229	0.013 (5.35%) <sup>a</sup>	56.0
CB_3_1.00	337	834	0.148	0.230	0.169	0.218	0.012 (5.23%) <sup>a</sup>	56.9
CB_3_1.00_W <sup>b</sup>	316	851	0.138	0.229	0.180	0.217	0.012 (5.40%) <sup>a</sup>	60.6
CB_2_0.55	316	900	0.137	0.238	0.180	0.228	0.009 (3.92%) <sup>a</sup>	58.1
CB_2_0.55	310	872	0.136	0.233	0.177	0.222	0.011 (4.63%) <sup>a</sup>	54.4
CB_2_0.55	307	864	0.137	0.233	0.171	0.221	0.011 (4.86%) <sup>a</sup>	55.5

<sup>&</sup>lt;sup>a</sup> Calculated as  $\frac{Vmeso}{Vt}$  100 b Carbons just washed with water that were not included in the design of experiments.



**Fig. 1**. Normal plots of standardized effects ( $\alpha = 0.05$ ) for chemically activated biochars: specific surface area (a); total pore volume (b); mesopore volume (c); micropore volume (d); ultra-micropore volume (e).

Regarding the results obtained in terms of specific surface area ( $S_{2D\text{-}NLDFT}$ ), Fig. 1a clearly shows that a higher concentration of activation agent led to a significant increase in the porosity of the resulting activated biochars. In contrast, the absolute pressure proved to be irrelevant for the values of final specific surface area. Our results regarding the development of porosity when higher impregnation ratios were used confirm the relevance of  $K_2CO_3$  as a chemical reagent for the production of tailored activated carbons. All pore volumes analyzed in the

present study were also found to be strongly dependent on the amount of activating agent (see Fig. 1). The chemical activation process was able to slightly broaden the original pore size distribution of the pristine biochar, leading to a more hierarchical porous structure with a large contribution of ultra-micropores and a slightly increased amount of mesopores. Almost all the carried out  $N_2$  isotherms required more than 70 h to be completed, thus confirming the predominant narrow microporous structure of chemically activated biochars (see Fig. A.3a), which hindered the diffusion of  $N_2$  within the porosity. For both total and micropore volumes, a statistically significant combined effect of pressure and impregnation ratio (AB) was found. However, these effects were comparatively much weaker than that observed for the main effect of the impregnation ratio (B). From a thermodynamics point of view, an increased pressure shifts the equilibrium of the activation reactions to the left. The fact that the textural properties of the resulting activated carbons were practically independent of pressure could indicate that the chemical activation process was mainly kinetically controlled.

It should be pointed out that the overall curvature terms for  $S_{2D\text{-}NLDFT}$ ,  $V_t$  and  $V_{micro}$  were statistically significant (p-values below 0.05, as reported in Table A.2). This indicates that, in further studies, our adopted factorial design should be expanded to a central composite design to be able to evaluate the pure quadratic regression coefficients and then apply Response Surface Methodology (RSM) for optimization purposes.

As shown in Table 2, after the carbons were washed only with hot water, there was a slight decrease in both the specific surface area and the micropore volume, compared to those measured for carbons washed with the acidic solution. This can be explained by the fact that water could not completely remove the remaining carbonate (and other chemical species) from the solid surface, thus causing blockage of some micropores. By focusing on the difference between the washing procedures of activated carbons at the same loadings of K<sub>2</sub>CO<sub>3</sub>, the results reported in Table 2 indicate that a higher activation pressure led to an increased amount of

product not soluble in water, i.e., to higher  $Y_{wash}$  values. This finding agrees with results reported by Malekshahian *et al.* [40], who concluded that high gasification pressures can limit the volatilization of potassium.

#### 3.2. Physical activation

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30 min to obtain a degree of burnout of 57%.

It must be pointed out that an increase in absolute pressure also implied an increase in the partial pressure of the reactant (CO<sub>2</sub>). In the literature, the effects of absolute pressure and partial pressure of reactant are commonly assessed separately. Nevertheless, and as far as we know, no previous studies have addressed the question of whether activation under pressure can cause textural changes in the resulting physically activated biochars. Activation of biomass-derived carbon had only been studied using supercritical water at high pressure [41], but such conditions are too different from the present ones for allowing some comparison to be done. Table 3 lists the textural properties of the activated carbons produced, whereas Fig. 2 shows the normal plots of the standardized effects obtained for each response variable. More detailed statistical outcomes are reported in Table A.3. The most relevant PSDs and N2 adsorption isotherms are shown in Figs. A.3b and A.5, respectively. As previously mentioned in section 2.2.2, the activation times (also shown in Table 3) were varied to obtain degrees of burnout in the range of 30% to 60 %, since the gasification rate was strictly dependent on reaction temperature and reactant partial pressure. As expected, the reaction rate was minimal at the lowest temperature (700 °C). In fact, and whatever the pressure applied, an activation time of 180 min was required to reach the desired reaction extent. For activation temperatures of 775 and 850 °C, an activation time of 60 min was enough, except for the material PB 850 1.00, for which this reaction time led to an almost complete gasification of the sample ( $\eta = 85\%$ ). Hence, the activation time at the highest levels of temperature and pressure was finally set at

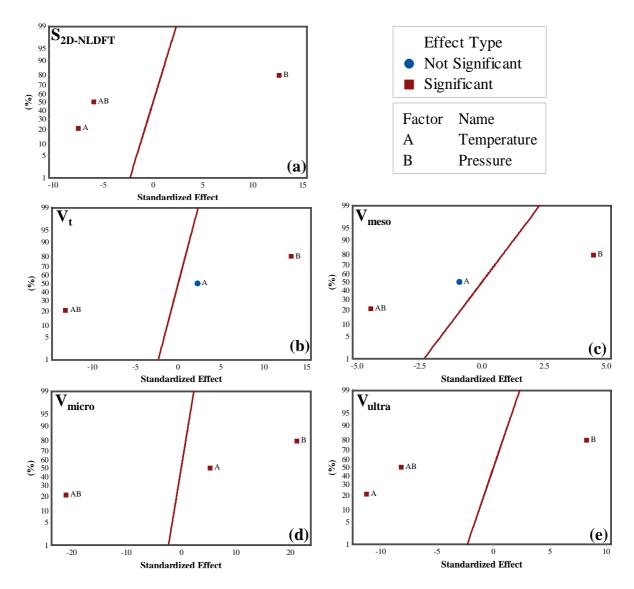
**Table 3.** Degrees of burnout activation times and textural properties for physically activated biochars

Material	Activation time	Burnout	Apparent specific surface area		Pore volumes				
	$t_{act}$	η	$A_{BET}$	$S_{2D\text{-}NLDFT}$	$V_{0.97}$	$V_t$	$V_{ultra}$	$V_{micro}$	$V_{meso}$
	min	%	$\mathrm{m^2~g^{-1}}$						
PB_700_0.10	180	30.1	552	833	0.237	0.243	0.184	0.227	0.016 (6.77%) <sup>c</sup>
PB_700_1.00	180	52.8	743	1008	0.333	0.333	0.226	0.301	0.032 (9.55%) <sup>c</sup>
PB_850_0.10	60	54.9	600	818	0.263	0.266	0.169	0.240	0.026 (9.64%) <sup>c</sup>
PB_850_1.00	30	56. 7	688	881	0.298	0.296	0.176	0.273	0.022 (7.57%) <sup>c</sup>
PB_775_0.50	60	43.7	719	931	0.314	0.312	0.176	0.285	0.027 (8.62%) <sup>c</sup>
PB_775_0.50	60	42.2	707	936	0.309	0.306	0.171	0.282	0.025 (8.02%) <sup>c</sup>
PB_775_0.50	60	42.4	707	949	0.308	0.306	0.174	0.282	0.023 (7.64%) °
PB_650_0.10 <sup>d</sup>	60	12.5	68.8	482	0.029	0.126	0.100	0.124	0.001 (1.07%) <sup>c</sup>
PB_650_1.00 <sup>d</sup>	180	25.0	447	750	0.191	0.214	0.154	0.200	0.014 (6.46%) <sup>c</sup>

<sup>&</sup>lt;sup>c</sup> Calculated as  $\frac{Vmeso}{Vt}$  100

It was clear that a high pressure accelerated carbon gasification, even at 700 °C. At this temperature, an increase in pressure from 0.1 to 1.0 MPa led to a marked increase in the burnout, from 30.1% to 52.8%, due to the higher reaction rate. In this sense, an increased CO<sub>2</sub> partial pressure resulted in a higher fraction of reactant adsorbed on the surface of the sample [42]. These outcomes are in agreement with the study conducted by Malekshahian *et al.* [42], but disagree with the results reported by Fermoso *et al.* [43], who observed that gasification of heartwood was enhanced using low partial pressures of CO<sub>2</sub>.

<sup>&</sup>lt;sup>d</sup> Additional carbons that were not included in the design of experiments.



**Fig. 2.** Normal plots of standardized effects ( $\alpha = 0.05$ ) for physically activated biochars: specific surface area (a); total pore volume (b); mesopore volume (c); micropore volume (d); ultra-micropore volume (e).

The specific surface areas reported in Table 3 were in line with expectations, given the present activation conditions. Even at the lowest temperatures (700 °C) it was possible to obtain relatively high surface areas, even greater than those measured for activated biochars produced at higher temperatures. As can be seen in Fig. 2a, the specific surface area was positively influenced by the absolute pressure and negatively affected by the activation temperature. Especially at 1.0 MPa, relatively low temperatures and longer activation times allowed the biochar to be gasified more homogeneously, thus resulting in a more developed porous

structure. Conversely, higher temperatures led to faster reaction rates, which can result in a more severe enlargement of micropores and the subsequent loss of surface area.

Fig. 2b shows that the total pore volume increased significantly when the pressure was raised from 0.1 to 1.0 MPa. In general, the total pore volumes for physically activated biochars, which were not significantly affected by the activation temperature, were higher than those measured for chemically activated carbons. As can be seen when comparing the data reported in Tables 2 and 3, physical activation led to porous carbons with slightly higher ultra-micropore volumes. An inspection of Figs. 2d, 2e and A.3b reveals that high temperatures can lead to some widening of the narrowest micropores, since the activation temperature had a negative effect on the ultra-micropore volumes and a positive one on the micropore volumes. With regards to the volume of mesopores, which was mainly affected by pressure (see Fig. 2c), it should be noticed that physical activation under pressure resulted in more hierarchical porous structures with higher contributions of mesopores (see values of  $V_{meso}$  and relative percentages in Table 3). The statistically significant overall curvature terms reported in Table A.3 for  $S_{2D}$ . NLDFT,  $V_t$ ,  $V_{meso}$ , and  $V_{micro}$  response variables also suggest that a central composite design could be required for optimization purposes.

Given the significant effects of pressure on the textural properties of carbons activated at 700–850 °C, we decided to perform two additional physical activations at 650 °C, leading to the materials called PB\_650\_0.10 and PB\_650\_1.00. The results obtained, which are also reported in Table 3, seem to confirm that activation with CO<sub>2</sub> under pressure is a very interesting way to produce biomass-derived porous carbons with high specific surface area and wide pore size distributions (including relatively high mesopore volumes), even at relatively low temperatures.

#### 3.3. Catalytic activity

Due to their relatively high specific surface area, both PB\_700\_1.00 and CB\_3\_0.10 activated biochars (one for each activation procedure) were selected as catalysts and tested during the upgrading process of the aqueous phase of a real pyrolysis oil.

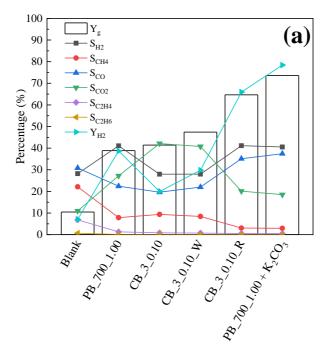
Elemental and moisture analyses of the aqueous phase of the pyrolysis oil revealed that the liquid sample had an average chemical formula of  $C_5 H_{6.7} O_2$  and a water content of 80 wt. %. Since the liquid was fed to the reformer without providing additional water, the steam to carbon molar ratio, S:C, was 4:1.

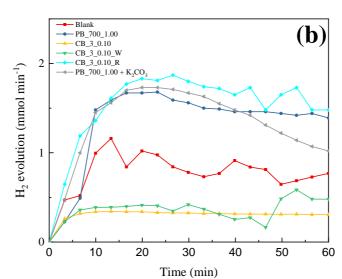
It is generally assumed that deactivation and/or instability of the catalyst can be attributed to two different phenomena: (1) the deposition of carbonaceous material (coke) on the surface of the catalyst, which clogs part of the available active sites; and (2) the extent of gasification reactions (both with steam and CO<sub>2</sub>) of the carbon-based catalyst, which leads to a loss of mass and a subsequent increase in LHSV (i.e., less contact time).

The results obtained in terms of total gas yield, selectivity towards specific gaseous species, hydrogen yield and hydrogen release over time are displayed in Fig. 3a. It should be noted that the poor results measured for the blank test (i.e., empty reactor) suggested that the reactor wall did not play a crucial catalytic role. The physically activated biochar (PB\_700\_1.00) exhibited performances comparable to those obtained in a previous study [33]. Using this material, which showed a good stability during the whole experiment (see Fig. 3b), a total gas yield of about 40% was obtained. Nevertheless, its relatively low hydrogen yield indicates that the extent of steam reforming was modest.

Regarding the performance of the chemically activated biochar (CB\_3\_0.10), a similar total gas yield was measured, compared to the physically activated one. However, selectivity to gaseous products were markedly different, leading to a decrease in the production of H<sub>2</sub> (the hydrogen yield was even lower than that of PB\_700\_1.00) and an increase in that of CO<sub>2</sub>. This

result could suggest that the decomposition of pyrolysis oil instead of steam and/or dry reforming, was the main process involved.





**Fig. 3**. Results obtained from combined steam and dry reforming of pyrolysis oil at 750°C: total and  $H_2$  yields ( $Y_g$  and  $Y_{H2}$ , respectively) as well as selectivities toward a given gaseous product,  $S_i$  (a); and evolution of the hydrogen production rate (b).

As can be deduced from Fig. 3a, similar results were also obtained for the chemically activated biochar that was washed with just water (CB\_3\_0.10\_W). The differences in the performance

between the physically and chemically activated biochars could be related to their textural properties. In this respect, the physically activated biochar exhibited a higher specific surface area and higher volumes of micro- and mesopores, thus providing more catalytic active sites of inherent alkali or alkaline earth metal species (AAEMs). The catalytic role of these species (especially K, Mg and Ca) in both steam reforming and gasification of carbon has been widely reported [25,44-46]. Furthermore, the more hierarchical pore size distribution of PB\_700\_1.00, with relatively high contributions of the mesopores, could shorten the diffusion path to reach the active sites. Fig. 3b clearly shows that H<sub>2</sub> release rates fluctuated over time for most of biochars tested. There are at least two possible explanations for this: First, the coke deposits, which gradually covered the catalyst surface, could have ted to large fluctuations in the extent of the reactions involved. Second, the structural modification of the activated carbons (the process temperature was higher than the activation temperature) could also led to unsteady hydrogen flow rates at the outlet. It should be noted that the reaction system studied was certainly complex, due the numerous competitive reactions that can affect the yields of the different gaseous products. Nevertheless, it can be assumed that methane is mainly produced by the cracking of the heaviest fraction of the pyrolysis oil, and that its yield will be correlated to the amount of coke produced [47]. Similarly, Fig. 3a reveals that the selectivity towards methane was almost constant for PB\_700\_1.00, CB\_3\_0.10, and CB\_3\_0.10\_W materials. Therefore, it can be concluded that the coke production in the process was quite similar for the above-mentioned carbons, suggesting that some structural modifications in the carbons could be accounted for the unsteady production of hydrogen. These modifications could be ascribed to the reverse Boudouard reaction [48,49], steam gasification and, in the case of the chemical activated biochars, potassium-catalyzed gasification. To support this argument, Fig. 4 shows the textural properties of fresh and spent activated biochars. Contrary to the loss of porosity observed for

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spent PB\_700\_1.00 and CB\_3\_0.10 catalysts, the spent CB\_3\_0.1\_W material showed an increase in pore volumes, especially for micropores. In addition, the results from TPD measurements, which are summarized graphically in Figs. 5 and A.6, indicate that the waterwashed chemically activated carbon exhibited a larger mass loss (with an increased release of CO<sub>2</sub> and CO, as shown in Fig. A.6) than that of the acid-washed carbon, thus confirming a greater extent of carbon gasification. This fact could be attributed to the residual amount of activation reagent available on the surface of the catalyst CB\_3\_0.10\_W, which might promote further biochar gasification during the experiment, as recently reported by Wang et al. [11]. To understand better the effect of the washing procedure on the performance of chemically activated biochars, an unwashed (raw) material (CB\_3\_0.10\_R) was also tested as catalyst. In this case, the total gas yield was 65% with enhanced selectivity towards both H<sub>2</sub> and CO (see Fig. 3a), thus indicating a higher extent of the steam reforming reactions (leading to a  $Y_{H2}$  of 66%). Fig. 6 displays the FTIR spectra obtained for fresh and spent chemically activated biochars as well as for pure K<sub>2</sub>CO<sub>3</sub>. In the case of acid and water-washed samples, no evident K<sub>2</sub>CO<sub>3</sub> content was detected on the surface, indicating the effectiveness of both washing procedures. On the other hand, most of the chemical activating agent available on the surface of the fresh CB 3 0.10 R catalyst disappeared after the catalytic test. The potassium carbonate (and other chemical species derived from its partial decomposition) available on the surface of the catalyst at the beginning of the reforming test (reductive environment) could progressively be reduced to metallic potassium  $(K^0)$ , leading to a further promotion of reforming reactions. Furthermore, a process temperature relatively close to the volatilization point of potassium can enhance the mobility and reactivity of the metal [50]. Generally speaking, activated biochars have relatively abundant oxygen and nitrogen-containing functional groups, which are not present in the materials shown in Fig. 6. The reason behind this could be the relatively high activation temperatures, since the majority of functional groups decompose below 800 °C [51].

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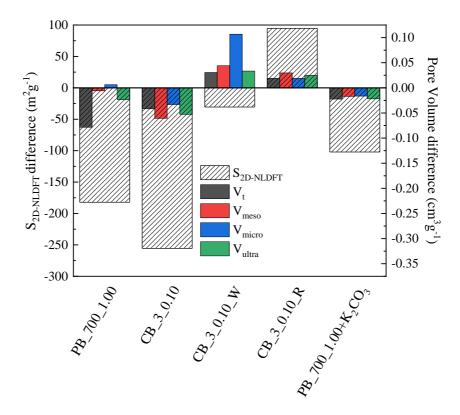
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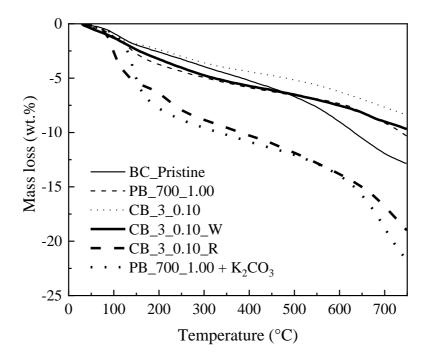
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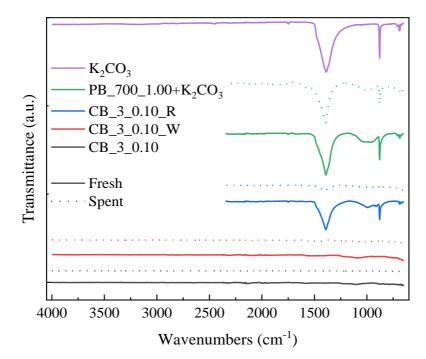
Furthermore, the relatively high amount of K<sub>2</sub>CO<sub>3</sub> loaded on the analyzed samples resulted in a very large peak, which may hide other peaks of interest, such as those related to oxygenated functionalities.



**Fig. 4**. Differences in textural properties between the fresh and spent activated biochars used as catalysts in steam and dry reforming tests.



486 Fig. 5. Results from TPD characterization (percentage of mass loss vs. temperature) for pristine487 and activated biochars.



**Fig. 6**. Comparison of the FTIR spectra obtained for pure K<sub>2</sub>CO<sub>3</sub> and some activated biochars tested in the present study.

The results from TPD measurements (see Fig. 5) showed that the mass loss under an inert atmosphere was much higher for the unwashed catalyst. In addition, the release of CO during TPD also increased for the material CB\_3\_0.10\_R (see Fig. A.6c). The increase in both mass loss and CO production when heating in N<sub>2</sub> up to 750 °C could be due to the reaction of K<sub>2</sub>CO<sub>3</sub> with carbon, which results in an additional release of CO, especially at temperatures near 700 °C and above, and the formation of catalytically active COK and CK complexes [52]. Hence, a certain gasification of carbon during the catalytic upgrading tests should be expected. In line with this, Fig.4 shows that the spent CB\_3\_0.10\_R exhibited more developed textural properties than the fresh one (for instance, the specific surface area increased from 29 m<sup>2</sup> g<sup>-1</sup> to 124 m<sup>2</sup> g<sup>-1</sup>), thus confirming the extent of gasification reactions. Despite the loss of catalyst through carbon gasification, which results in a higher LHSV, the production rate of H<sub>2</sub> for this catalyst was relatively stable during the experiment. This could be explained by the fact that the carbon gasification catalyzed by K<sub>2</sub>CO<sub>3</sub> contributes to creating new pores and then counterbalancing the deposition of coke, which induces micropore blockage and active sites coverage. Based on the findings discussed above, it seems reasonable to assume that an activated biochar having a wider hierarchical pore size distribution and some availability of K<sub>2</sub>CO<sub>3</sub> on its surface appears as an excellent candidate for catalytic pyrolysis vapors upgrading. With this in mind, a new biochar-derived material, designated as PB\_700\_1.00+K<sub>2</sub>CO<sub>3</sub>, was prepared by impregnating the material PB\_700\_1.00 with K2CO3 at a mass ratio of carbonate to precursor of 3:1. It should be expected that the better textural properties of the starting physically activated biochar could lead to a more homogeneous dispersion of the active phase on the carbon support, thus resulting in an improved performance of the catalyst. The results obtained for the catalyst PB\_700\_1.00+K<sub>2</sub>CO<sub>3</sub> confirmed our expectation. The

total gas yield reached a maximum value of 74%, whereas the hydrogen yield was the highest

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reported in the present study (78%). From the FTIR spectra given in Fig. 6, it can be seen that a fraction of K<sub>2</sub>CO<sub>3</sub> was still present on the catalyst surface after running the test.

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XPS analyses were carried out for both PB\_700\_1.00 and PB\_700\_1.00+K<sub>2</sub>CO<sub>3</sub> materials to get additional insight into the role of K<sub>2</sub>CO<sub>3</sub> during the pyrolysis vapors upgrading process. From the summarized XPS results given in Table 4, it can be observed that the atomic composition of PB\_700\_1.00 remained relatively constant after the catalytic test, even though the simultaneous increase of the peaks related to CII and OII seemed to indicate the formation of phenolic groups on the catalysts. The comparison of the atomic compositions between the fresh and spent PB\_700\_1.00+K<sub>2</sub>CO<sub>3</sub> materials indicates that the extent of carbon gasification reactions was much higher than that of the PB\_700\_1.00 material. Furthermore, from the spectra displayed in Fig. A.7, an additional peak at 283 eV, which is attributed to KC<sub>x</sub> carbides, was observed for the spent PB 700 1.00+K<sub>2</sub>CO<sub>3</sub>. On the other hand, the K2p3/2 region presented only a contribution and suggested the presence of potassium on the surface as K<sub>2</sub>O or K<sub>2</sub>O<sub>2</sub>, according to Li et al. [53]. This fact was also confirmed by the shift to lower binding energies of the OII and OIII peaks (see Fig. A.7). The marked difference in the potassium content between fresh and spent PB\_700\_1.00+K<sub>2</sub>CO<sub>3</sub> was probably due to the migration of the potassium-based compounds from the internal pores to the external surface, which can lead to the formation of a coating layer composed of K<sub>2</sub>O, as confirmed by SEM-EDX images reported in Fig. 7. The presence of potassium oxides on the surface of the catalyst can be ascribed to (i) the direct decomposition of the carbonate (Reaction 7) and/or (ii) the formation of metallic potassium. The latter can migrate from the inner structure of the catalyst to its surface during the course of the pyrolysis vapors upgrading tests and finally be oxidized when the catalyst has been removed from the reactor. This fact can also justify the significant decrease in the specific surface area reported in Fig. 4 for the spent PB\_700\_1.00+K<sub>2</sub>CO<sub>3</sub> catalyst (from 130 to 27.7  $\text{m}^2\text{g}^{-1}$ ).

From the SEM-EDX analyses, it can also be deduced that a relatively homogeneous dispersion of K<sub>2</sub>CO<sub>3</sub> was achieved for the fresh catalyst. Nevertheless, both fresh and spent catalysts exhibited some clusters and snail shell-like agglomerates, which could be attributed to the relatively high load of carbonate. However, no visible surface carbon layers was found on the superficial K<sub>2</sub>O of the spent catalyst, thus suggesting that the extent of the carbon gasification was probably sufficient to prevent the deposition of coke on the surface.

**Table 4.** Surface composition measured by XPS and peak contributions of C1s, O1s and K2p.

Sample	Surface concentration (at. %)					Bindir	ng energi	es (eV) and	d relative	peak are	eas (%) <sup>e</sup>	
						C1s region				O1s region		K2p3/2 region
	C	О	K	CI	CII	CIII	CIV	Carbide	OI	OII	OIII	K-O
PB_700_1.00 (Fresh)	87.3	11.3	1.4	284.4 (68.5)	285.7 (23.8)	287.6 (6.3)	289.2 (1.3)		531.1 (30.7)	532.8 (55.4)	534.5 (13.9)	293.3 (100)
PB_700_1.00 (Spent)	82.8	15.1	2.1	284.5 (62.7)	285.5 (31.3)	287.6 (4.1)	289.4 (1.9)		531.3 (20.0)	532.9 (60.2)	534.5 (19.8)	293.7 (100)
PB_700_1.00+K <sub>2</sub> CO <sub>3</sub> (Fresh)	72.6	21.6	5.8	284.6 (47.3)	285.8 (40.6)	287.6 (11.0)	289.3 (1.1)		531.3 (12.0)	532.7 (50.5)	534.5 (37.5)	293.7 (100)
PB_700_1.00+K <sub>2</sub> CO <sub>3</sub> (Spent)	42.0	27.8	30.2	284.5 (64.2)			288.7 (22.5)	283.1 (22.5)	531.1 (57.9)			292.8 (100)

<sup>&</sup>lt;sup>e</sup>C1s binding energies in carbon materials: **CI**, hydrocarbons, aromatics, aliphatics (284.5 eV); **CII**, single bond C-O associated to alcohols, phenols, carboxyls (286 eV); **CIII**, double bond C=O in carbonyl, quinone (287.5 eV); **CIV**, carboxyl and carbonate groups (288.7 eV).

O1s binding energies in carbon materials: **OI**, C= O quinone type groups (around 531 eV); **OII**, C-OH phenol groups and/or C-O-C ether groups (532.5 eV), **OIII**, chemisorbed oxygen (COOH carboxylic groups) and/or water (535 eV).

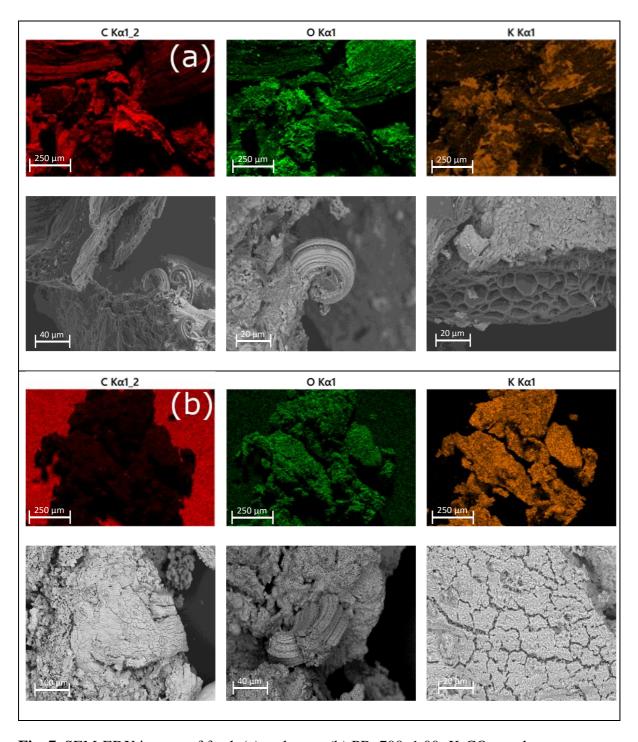


Fig. 7. SEM-EDX images of fresh (a) and spent (b) PB\_700\_1.00+K<sub>2</sub>CO<sub>3</sub> catalyst.

# 4. Conclusions

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From the results obtained in this study, the following conclusions can be drawn:

1. The chemical activation of the pristine biochar led to carbonaceous solids with a relatively high specific surface area and well-developed microporous structure. The

most important activation parameter was found to be the amount of activation agent, which accounted for the development of the porous structure. Activation pressure, on the other hand, was not significant for the outcomes of the procedure. Regarding the washing efficiency, acid washing resulted to be effective in cleaning the surface of the samples, whereas washing with water was not sufficient to remove completely the residual reactants, partially clogging the porosity of the samples.

- 2. Physical activation was found to be more sensitive to the process parameters. It was possible to obtain acceptable values of specific surface area even at low temperature (650 °C) by increasing the activation pressure to 1.00 MPa. A negative effect of the activation temperature on the results was also observed. In fact, the surface area decreased with the increase in the activation temperature, probably due to the widening of the smaller pores by a too rapid (and thus less controlled) gasification.
- 3. The most promising material produced by physical or chemical activation (PB\_700\_1.00 and CB\_3\_0.10) were tested as catalysts in the upgrading of the aqueous fraction of a real pyrolysis oil. Both activated biochars, which were initially characterized by a well-developed porosity, did not exhibit good performance, showing a huge decrease in the specific surface area, probably due to coke deposition. Nevertheless, the addition of K<sub>2</sub>CO<sub>3</sub> to the best physically activated biochar boosted the overall performance towards the production of more hydrogen, as a consequence of the enhanced reforming of pyrolysis oil. This finding can be related to the availability of K<sup>0</sup>, which was formed from the decomposition of the K<sub>2</sub>CO<sub>3</sub> previously deposited on the activated biochar.

## Acknowledgments

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## Appendix A. Supplementary data

- 593 Further details on characterization of pristine and activated biochars. Statistical outcomes from
- the adopted designs of experiments.

#### Nomenclature

596	$A_{BET}$	Brunauer-Emmett-Teller area (m <sup>2</sup> g <sup>-1</sup> )
597	$d_p$	Pore diameter (nm)
598	$F_{BO}$	Molar flow rate of dry bio oil (mol min <sup>-1</sup> )
599	$F_{H2Av}$	Experimental average H <sub>2</sub> molar flow rate (mol min <sup>-1</sup> )
600	$F_{H2\ Stoi}$	H <sub>2</sub> stoichiometric molar flow rate (mol min <sup>-1</sup> )
601	$m_0$	Initial mass of biochar before activation (g)
602	$m_c$	Biochar mass before the washing step (g)
603	$m_f$	Final mass of biochar after activation (g)

604	$m_g$	Total mass of produced gas during the upgrading process (g)
605	$m_l$	Mass of liquid fed into the upgrading reactor (g)
606	$m_w$	Biochar mass after the washing step (g)
607	$n_i$	Produced amount of a given gaseous specie i (mol)
608	$n_{tot}$	Total amount of produced gas (mol)
609	$S_{2D ext{-}NLDFT}$	2D-NLDFT specific surface area (m <sup>2</sup> g <sup>-1</sup> )
610	$S_i$	Selectivity toward a given gaseous specie $i$ (%)
611	$V_{0.97}$	Gurvitch pore volume (cm <sup>3</sup> g <sup>-1</sup> )
612	$V_{meso}$	Volume of mesopores (cm <sup>3</sup> g <sup>-1</sup> )
613	$V_{micro}$	Volume of micropores (cm <sup>3</sup> g <sup>-1</sup> )
614	$V_t$	Total pore volume (cm <sup>3</sup> g <sup>-1</sup> )
615	$V_{ultra}$	Volume of ultra-micropores (cm <sup>3</sup> g <sup>-1</sup> )
616	$Y_g$	Total gas yield (%)
617	$Y_{H2}$	Hydrogen yield (%)
618	$Y_{wash}$	Washing yield (%)
619	η	Degree of burnout (%)
620	Acronyms	
621	FTIR	Fourier-Transform Infrared spectroscopy
622	GHSV	Gas hourly space velocity
623	LHSV	Liquid hourly space velocity

624	PSD	Pore size distribution
625	RSM	Response surface methodology
626	S:C	Steam to carbon molar ratio
627	STP	Standard temperature and pressure
628	TPD	Temperature-programmed desorption
629	XPS	X-ray photoelectron spectroscopy
630	μ-GC	Micro gas chromatograph
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**Appendix A: supplementary material** 

Influence of activation conditions on textural

properties and performance of activated biochars for

pyrolysis vapors upgrading

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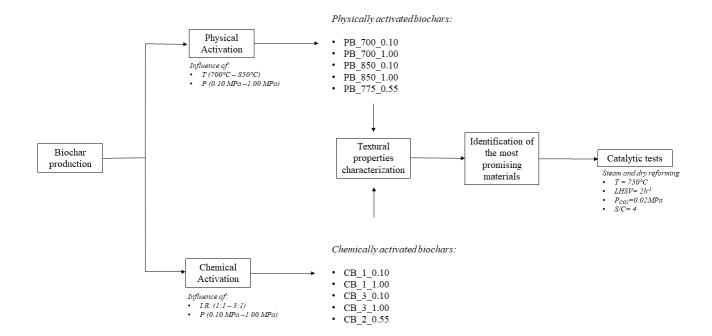
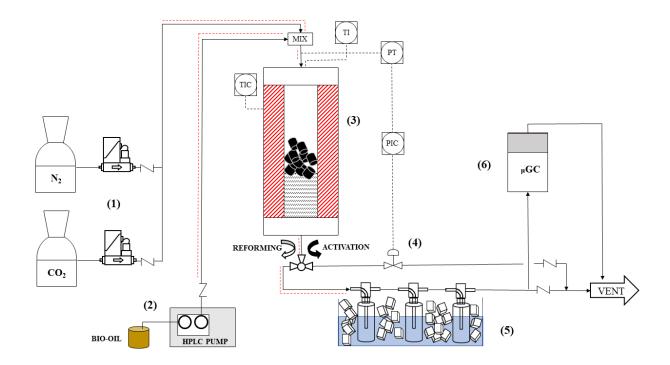


Fig. A.1. Schematic representation of the methodology followed in this study.



**Fig. A.2**. Overview of the experimental device used for activation and reforming tests: Feeding system (1); HPLC pump (2); fixed-bed reactor and furnace (3); Servo-controlled valve (4); condensation train (5); and  $\mu$ -GC analyzer (6).

**Table A.1.** Results of textural characterization, proximate analysis and elemental analysis of pristine biochar derived from wheat straw

Proximate analysis (wt. %)					
Moisture	$2.00 \pm 0.08$				
Volatile matter	$10.8 \pm 0.63$				
Ashes	$12.7 \pm 0.21$				
Fixed carbon	$74.5 \pm 0.55$				
Ultimate (v	wt. % on a dry ash-free basis)				
C	$92.6 \pm 0.07$				
H	$4.02\pm0.05$				
N	$2.01 \pm 0.08$				
0	1.32				
Molar H:C ratio	0.52				
Tex	ctural characterization				
$A_{BET}$ (m <sup>2</sup> g <sup>-1</sup> )	2.90				
$S_{2D\text{-}NLDFT}$ (m <sup>2</sup> g <sup>-1</sup> )	341				
$V_{0.97}  ({\rm cm}^3  {\rm g}^{-1})$	0.0026				
$V_{tot}$ (cm <sup>3</sup> g <sup>-1</sup> )	0.084				
$V_{ultra}~({ m cm}^3~{ m g}^{-1})$	0.083				
$V_{micro}$ (cm <sup>3</sup> g <sup>-1</sup> )	0.083				
$V_{meso}$ (cm <sup>3</sup> g <sup>-1</sup> )	0.001 ( <b>1.28 vol. %</b> )				

**Table A.2.** Regression coefficients and quality-of-fit statistics for the design of experiments on chemical activation. The values in brackets correspond to the p-values resulting from the t-tests. The significant terms are marked in bold

	Response variable						
	$S_{2D ext{-}NLDFT}$	$V_t$	$V_{meso}$	$V_{micro}$	$V_{ultra}$		
βο	810.1	0.21605	0.009516	0.2065	0.16447		
	(0.000)	(0.000)	(0.003)	(0.000)	(0.000)		
$\beta_A$ (Pressure)	-12.24	-0.00364	0.000215	-0.00385	-0.00553		
	(0.326)	(0.132)	(0.715)	(0.190)	(0.150)		
$\beta_B$ (Mass ratio)	73.32	0.02801	0.002904	0.02511	0.01871		
	(0.016)	(0.003)	(0.030)	(0.006)	(0.016)		
$oldsymbol{eta_{AB}}$	-36.90	-0.1089	-0.000625	-0.01026	-0.00865		
	(0.060)	(0.018)	(0.346)	(0.035)	(0.070)		
$R^2_{adj}$ (%)	94.05	98.78	83.90	97.38	93.27		
Curvature	(0.042)	(0.015)	(0.350)	(0.029)	(0.090)		

**Table A.3.** Regression coefficients and quality-of-fit statistics for the design of experiments on physical activation. The values in brackets correspond to the p-values resulting from the t-tests. The significant terms are marked in bold

	Response variable					
	$S_{2D ext{-}NLDFT}$	$V_t$	$V_{meso}$	$V_{micro}$	$V_{ultra}$	
βο	884.9	0.29181	0.023189	0.268623	0.19055	
	(0.000)	(0.000)	(0.001)	(0.000)	(0.000)	
$\beta_A$ (Temperature)	-35.46	0.00381	-0.000811	0.004623	-0.01445	
	(0.017)	(0.155)	(0.461)	(0.034)	(0.008)	
$\beta_B$ (Pressure)	59.54	0.02250	0.004000	0.018500	0.01050	
	(0.006)	(0.006)	(0.047)	(0.002)	(0.015)	
$oldsymbol{eta_{AB}}$	-28.08	-0.02250	-0.004000	-0.018500	-0.01050	
	(0.027)	(0.006)	(0.047)	(0.002)	(0.015)	
$R^2_{adj}$ (%)	98.06	98.47	86.47	99.42	98.22	
Curvature	(0.018)	(0.025)	(0.328)	(0.008)	(0.013)	

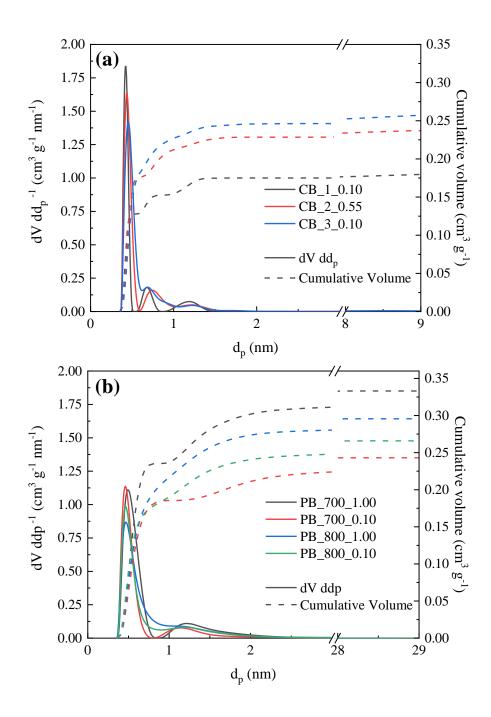
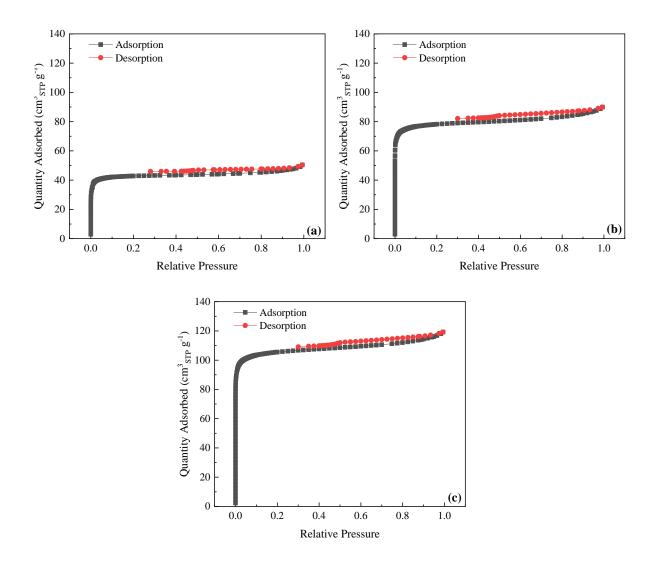
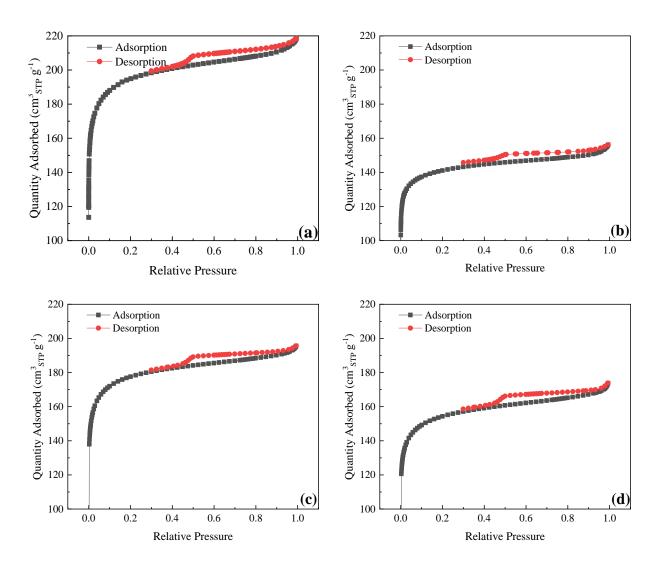


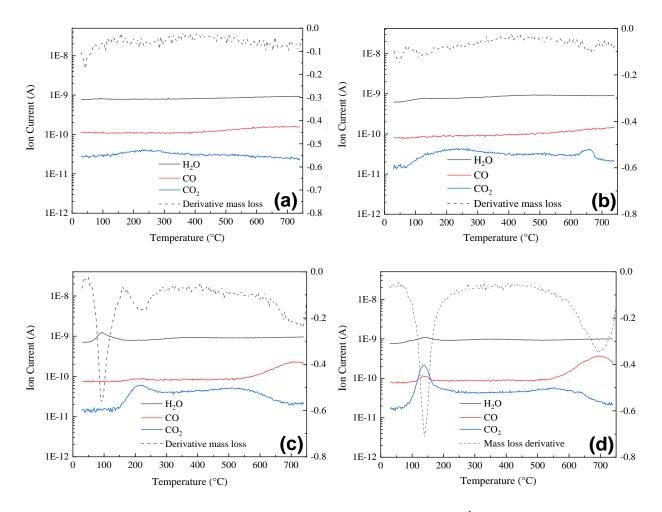
Fig. A.3. PSD and cumulative pore volume of chemical (a) and physical (b) activated biochars.



**Fig. A.4**.  $N_2$  adsorption/desorption isotherms of the following chemically activated biochars:  $CB_3_0.10$  (a);  $CB_2_0.55$  (b); and  $CB_3_0.10$  (c).



**Fig. A.5**. N<sub>2</sub> adsorption/desorption isotherms of the following physically activated biochars: PB\_700\_1.00 (a); PB\_700\_0.10 (b); PB\_850\_1.00 (c); and PB\_850\_0.10 (d).



**Fig. A.6**. Differential thermogravimetric curves (in % of mass min<sup>-1</sup>) and profiles of released species (H<sub>2</sub>O, CO<sub>2</sub>, and CO) from the TPD measurements conducted for the following materials: CB\_3\_0.10 (a); CB\_3\_0.10\_W (b); CB\_3\_0.10\_R (c); and PB\_700\_1.00+K<sub>2</sub>CO<sub>3</sub> (d).

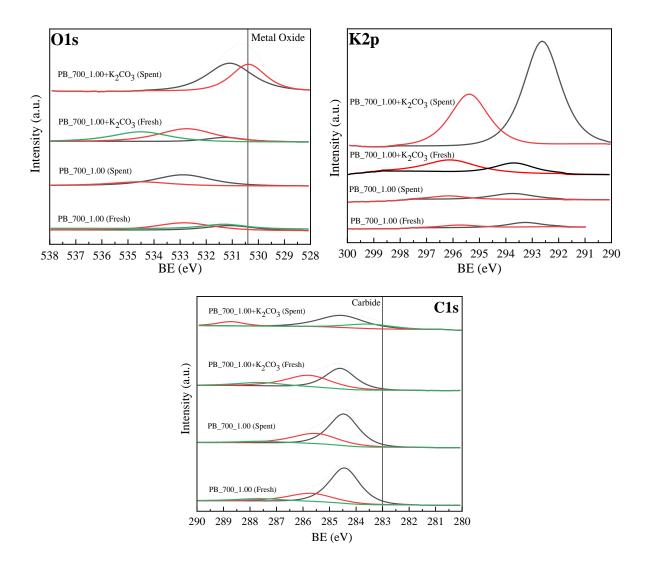


Fig. A.7. C1s, O1s and K2p spectra of fresh and spent PB\_700\_1.00 and PB\_700\_1.00+K<sub>2</sub>CO<sub>3</sub>.